Coric



Twist-on efocus Miniaturized Fluorescence Microscope

doriclenses.com

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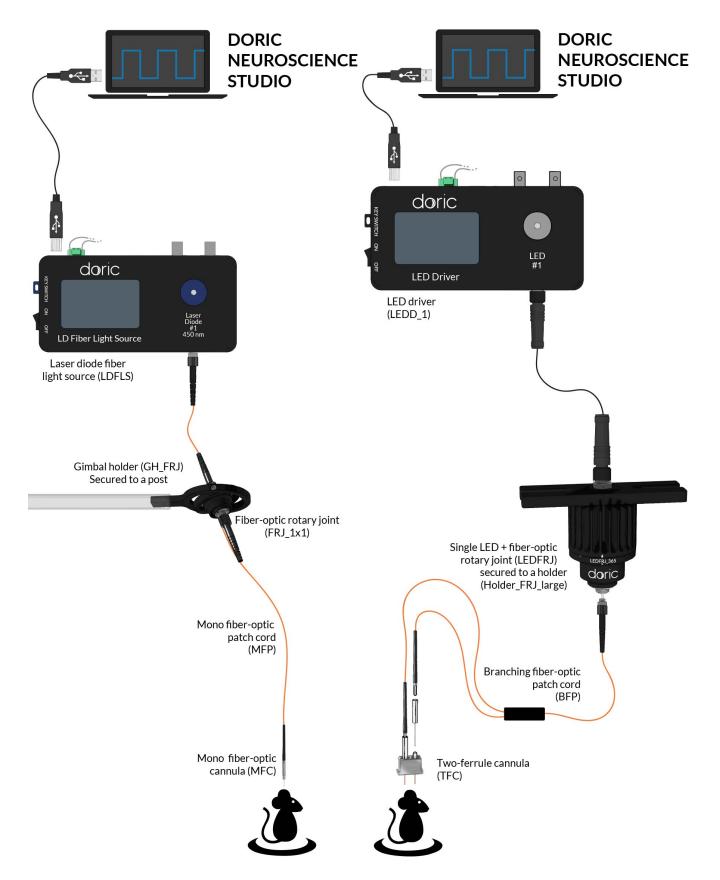
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Overview of Neuro-photonics products catalog

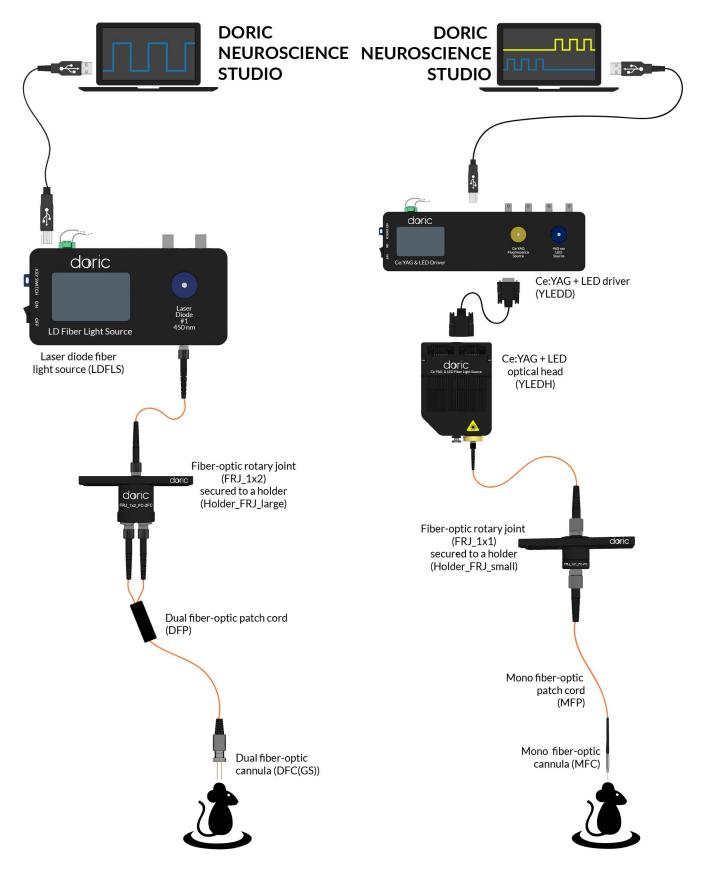
Over the years, this catalog has grown beyond its optogenetics roots to become a reference for neuro-photonics products. Initially, it covered only the hardware that was used for light stimulation and/or control of cells marked with genetically encoded light-sensitive proteins. In these experiments, the light from a laser or an LED source is sent through an optical fiber to a slice of brain tissue, or to the brain of a head-fixed or freely-moving animal. Overtime, this simple optical link has evolved into more complex circuitry, resembling the early days fiber-optic telecommunication networks. This fiber-to-the-brain network consists of fiber coupled light sources and their drivers, light shutters or modulators, rotary joints for experiments with freely-moving animals, beam-splitters, fiber-optic patch cords, various fiber-optic cannulas and much more. In addition to delivering light pulses to the tissue, this network monitors the interaction of tissue with light, sends and records electrical signals and if required administers different fluids.

Optogenetics uses light to control certain brain cells by opening/closing the ion channels in the membranes of genetically modified brain cells expressing light-sensitive ion channels. However, it does not generate any optical signal. On the other hand, fiber photometry and miniature fluores-cence microscopy are perfect tools for monitoring changes in brain cell activity of freely-moving animals. Both use excitation light to generate fluorescence signal which is an indicator of neuronal activity. The difference between the two is that fiber photometry detects overall optical signal from a group of labelled cells, whereas fluorescence microscopy records an image of optical signal from within the field of view.

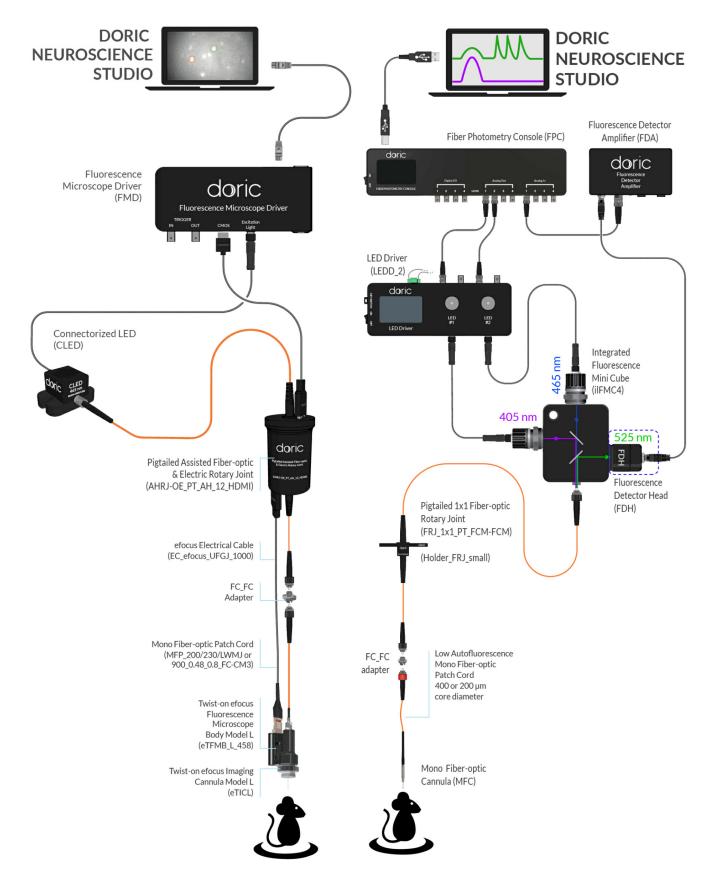
The aim of this catalog is to present each neuroscience application in the context of required optical hardware. In addition, there are several products combining these methods amongst themselves and with well-proven methods like electrophysiology. The following pages show system examples involving fiber-based optogenetics, fiber photometry, fluorescence microscopy systems and electrophysiology.



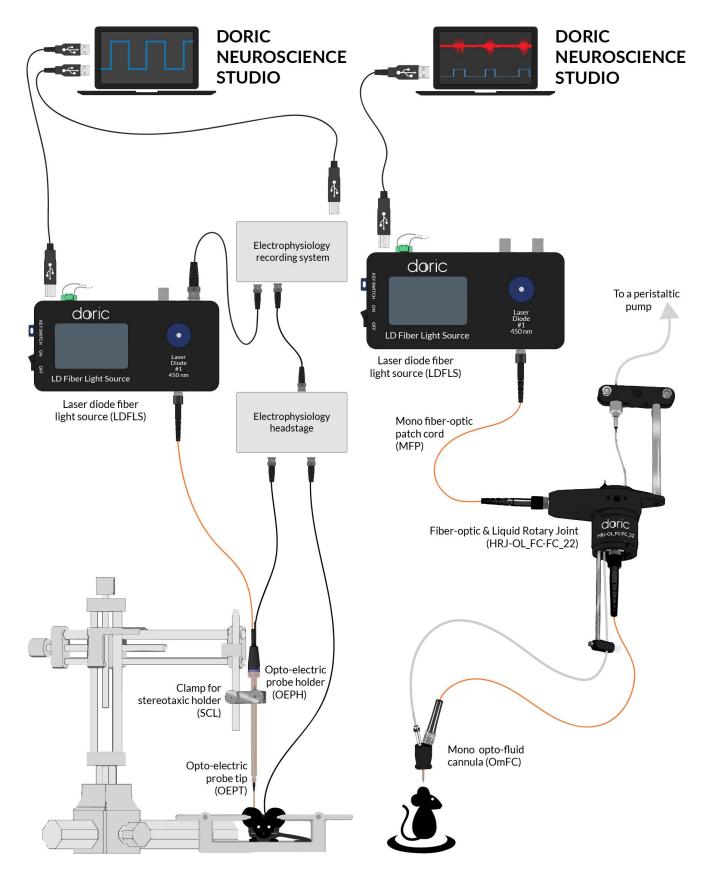
(Left) Optogenetic stimulation with a Laser Diode Fiber Light Source. (**Right**) Bilateral optogenetic stimulation with a Connectorized LED with Fiber-optic Rotary Joint.



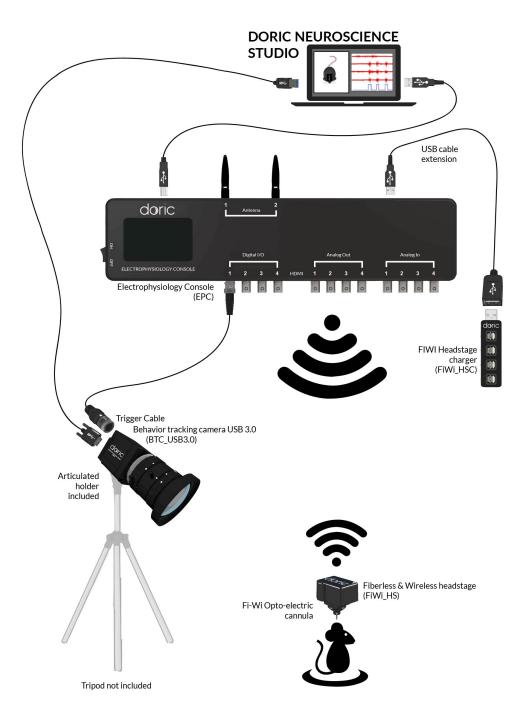
(Left) Dual optogenetic stimulation with a Laser Diode Fiber Light Source. (**Right**) 2-color optogenetic stimulation with a Ce:YAG + LED Fiber Light Source.



(Left) Miniaturized Fluorescence Microscopy System used for calcium imaging with GCaMP6. **(Right)** Fiber Photometry System with a 4 port Integrated Fluorescence Mini Cube and a Fiber Photometry Console.



(Left) Optogenetic stimulation and electrophysiology recordings with an Opto-electric Probe Tip. (Right) Optogenetic stimulation and fluid delivery with a Mono Opto-fluid Cannula.



Fiberless & Wireless Optogenetically Synchronized Electrophysiology System and Behavior Tracking Camera

Light Generation & Control

LED Illumination

Light emitting diodes (LEDs) coupled in an optical fiber are suitable for neuroscience experiments which need to bring light into the brain. LED light allows to control the excitation, inhibition or signalling of specific cells in optogenetic experiments. The uniform illumination of an LED makes it the preferred light source for miniature fluorescence microscopy and fiber photometry. Our compact *Connectorized LEDs* or multiple color combined LEDs are used with programmable *LED Drivers*. We also offer LED Fiber Light Sources integrating 1, 2, or 4 independently controlled LEDs into the driver housing.

LED Modules

Connectorized LEDs

Doric *Connectorized LEDs* couple high brightness LEDs into an FC receptacle compatible with an FC connectorized fiberoptic patch cord. Each *Connectorized LED* is actively aligned for optimum output power and its optical design provides the maximum fiber-coupling efficiency into multimode optical fibers.



Connectorized LED

Each *Connectorized LED* includes an EPROM memory enabling its identification by the driver. The wavelength is recognized and the maximum current is automatically set to avoid accidental overdrive. Doric *Connectorized LEDs* are easily screwed on an optical table for a basic passive cooling suitable for low power and pulsed operations. During high power Continuous Wave (**CW**) applications, an active cooling is obtained by connecting the internal fan with a micro-USB power supply. This is essential to maximize the device life span and to obtain stable performances in terms of output power.

Notes:

- A micro-USB power supply is included with each *Connectorized LED*.
- A *Connectorized LED* does not include the corresponding LED Driver. See Table 4 for available LED Driver models.
- An Optical Breadboard for Connectorized LED (LEDB; see Table 115) is available to mount systems including two Connectorized LEDs.

LED		TYPICAL OUTPUT POWER @1000 mA (mW) [*]			Overdrive @2000 mA (pulsed)	
Central Wavelength (nm)	Bandwidth FWHM (nm)	Core 200 μm 0.53 NA	Core 400 μm 0.53 NA	Core 960 μm 0.63 NA		
365	~12	6.0	23	100	*	
385	~12	6.0	23	100	*	
405	~15	5.0	23	100	*	
420	~15	5.5	23	100	*	
450	~25	8.0	23	100	x1.7	
465	~25	7.5	23	100	x1.7	
505	~30	3.0	12	50	x1.6	
515	~40	3.0	9.5	40	x1.5	
560	~100	2.0	8.5	40	-	
595	~20	2.0	8.5	40	x1.2	
625	~20	3.5	14	70	x1.6	
635	~20	6.5	25	100	x1.6	
840	~35	6.0	22	40	-	
940	~35	2.0	10	40	-	
5500K	-	4.5	17	80	-	

Table 1: Typical Connectorized LED Output Power vs Optical Fiber Core Diameter

The power is given for *Connectorized LEDs* and *LEDs* with *Fiber-optic Rotary Joints* (LEDFRJ)[†]. Contact us for power levels for other *LED* products.



Connectorized LED male pinout

^{*}All power values taken at a maximum current of 1000 mA, except for 365, 385, 405 and 420 nm LEDs (500 mA). [†]In overdrive mode, LED drivers can produce current pulses of up to 2000 mA.

Color	Central Wavelength (nm)	LED Color Code
Near UV	365	365
Near UV	385	385
Near UV	405	405
Violet	420	420
Royal	450	450
Blue	465	465
Cyan	505	505
Green	515	515
Lime	560	560
Amber	595	595
Orange	625	625
Red	635	635
Infrared	840	840
Infrared	940	940
White	5500K	W55

Table 2: Connectorized LEDs Color Codes

ORDERING CODE: CLED_

LED color code (see Table 2) ——

LED Drivers

Doric programmable *LED Drivers* are available in 1-, 2-, and 4-channel versions. When connected to a Connectorized LED having an eprom memory, the *LED Driver* recognizes the LED wavelength and automatically sets the maximum current value.



Two-channel LED Driver

In stand-alone mode, all LED Drivers allow cw

operation and external analog modulation through an input BNC connector for each channel. For each channel, there is also a current monitoring BNC output allowing data acquisition or triggering of other devices. When using Doric Neuroscience Studio Software, more advanced operating modes are available such as TTL modulation and software defined illumination sequences, thus eliminating the need for a function generator. In low-duty cycle pulsed mode, the software allows to overdrive the LED sources if a higher power is needed. For multiple channel driver versions, each channel is controlled independently. Although not mandatory for LED sources, our *LED Drivers* come with a safety interlock connector and a main key switch. These safety features are of interest for UV and near infrared LEDs.

Note: The renewed line of Doric *LED Drivers* has a new connector pinout that does not include pins for fan power. It is thus essential to use a *Fan Power Adapter* (**FPA**; see Table 114) when using *Combined LEDs* or *Combined LEDs with a Fiber-optic Rotary Joint*. This power adapter is suitable for up to 4 channels and is sold with corresponding M8 cables.

SPECIFICATION	VALUE
Maximum current	1 A (2 A overdrive)
Input BNC modulation	0 - 5 V TTL or analog (400 mA/V)
Output BNC monitoring	0 - 5 V (2.5 V/A)
Output LED connector	M8 4-pin female



LED Driver female pinout

Table 4: LED Drivers	Ordering Codes
----------------------	----------------

Number of Channels	Ordering Code
1	LEDD_1
2	LEDD_2
4	LEDD_4
8	LEDD_8 [*]

¹⁴

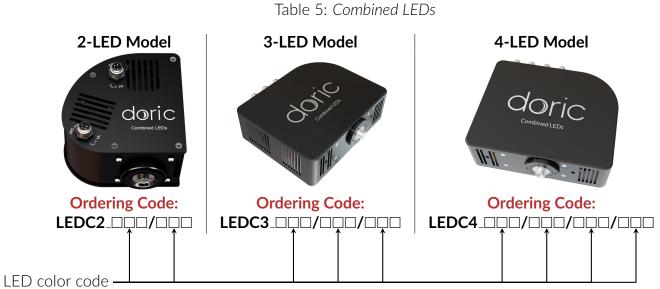
^{*8-}channel LED Driver is available on request

Combined LEDs

Doric *Combined LEDs* merge the light from multiple LEDs of different colors into a single output connector by using a patent pending regular pentagon mirrors configuration. The coupling efficiency for respective colors is near those of our Connectorized LEDs. Each LED of the *Combined LEDs* is driven independently via M8 cable when connected to any of our driver(s).

Notes:

- A compatible holder is included to secure the Combined LEDs.
- *Combined LEDs* do not include the corresponding LED Driver. See Table 4 for available LED Driver models.
- The renewed line of Doric LED Drivers has a new connector pinout that does not include pins for fan power. When using the *LEDC2*, it is thus essential to use a *Fan Power Adapter* (**FPA**; see Table 114).



(see Table 2)

LED + Fiber-optic Rotary Joint

Connectorized LED with Fiber-optic Rotary Joint

It is a common practice to connect an LED with a rotary joint via a fiber-optic patch cord. If the tips of the patch cord are not coated, which is usually the case, at least 8% of the light power is lost from the Fresnel reflections, in addition to other connection losses. One way of getting around these losses is to integrate the LED source and the fiber-optic rotary joint in a single device, thus eliminating one fiber-optic patch cord. That is the purpose of Doric Connectorized LED sources with fiber-optic rotary joint.



Connectorized LED + Fiber-optic Rotary Joint

Notes:

- A compatible holder is included with the *Connectorized LED* + *Fiber-optic Rotary Joint* (**Holder_-FRJ_large**; see Table 117).
- An optional gimbal holder allows pivoting the rotary joint along two additional axes, further reducing the mechanical stress on the animal (**GH_FRJ**; see Table 119).
- A *Connectorized LED with Fiber-optic Rotary Joint* does not include the corresponding LED Driver. See Table 4 for available LED Driver models.

Combined LEDs with Fiber-optic Rotary Joint

Combined LEDs with Fiber-optic Rotary Joint are perfect for the light activation of multiple opsins (*e.g.* channelrhodopsin and halorhodopsin). Other combinations of LED wavelengths are available as long as their spectra do not overlap.

New types of opsins are frequently emerging from ongoing research. Doric *Combined LEDs with Fiber-optic Rotary Joint* are easily customized to most sets of activation wavelengths. Our patent pending assemblies provide the possibility to combine up to four distinct wavelengths and couple them into a single output rotary joint.



Table 6: Combined LEDs with Fiber-optic Rotary Joint

Notes:

- A compatible holder is included to secure the Combined LEDs with Fiber-optic Rotary Joint.
- A Combined LEDs with Fiber-optic Rotary Joint does not include the corresponding LED Driver. See Table 4 for available LED Driver models.
- The renewed line of Doric LED Drivers has a new connector pinout that does not include pins for fan power. It is thus essential to use a *Fan Power Adapter* (**FPA**; see Table 114) when using *Combined LEDs with Fiber-optic Rotary Joint*. This power adapter is suitable for up to 4 channels.

LED Fiber Light Sources



2-channel LED Fiber Light Source

The LED Fiber Light Source is an assembly of one or multiple independent LEDs and their driving electronics into a compact housing. Each LED has its own output FC connector.

The functionalities and software of Doric LED Fiber Light Sources are identical to those of LED Drivers. When ordering multi-channel models, any combination of LED wavelengths can be chosen according to the following ordering codes.

ORDERING CODE:
1-channel model LEDFLS_DDD
2-channel model LEDFLS_DDD_DDD
4-channel model LEDFLS
LED color codes (see Table 2)

Note: The typical output power of each LED is shown in Table 1.

Laser Diode Illumination

Connectorized Laser Diode Modules

Our miniature Connectorized Laser Diode Modules have FC/APC receptacles compatible with FC/APC connectorized multimode optical fibers having 50 μ m or larger core diameters and at least 0.22 NA. With laser diode sources, using FC/APC connectors is essential to avoid optical feedback and the corresponding intensity noise. The laser diode module size is 24.6 x 36.8 x 12.0 mm³, excluding the base plate and the electric cable. The base plate is used as a passive heat sink and can be used to secure the module on an optical table for an even better thermal stability. The module connects only to Doric Laser Diode Module Driver over the M8 electrical cable.



Connectorized Laser Diode Module

Each module contains an EPROM memory allowing the Laser Diode Module Driver to recognize the device and set the corresponding maximum current, thus preventing accidental overdrive of the laser diode by the user. The available wavelengths and fiber-coupled output power values are given in Table 7.

 Central Wavelength(nm)	Bandwidth FWHM (nm)	Power (mW) [*]	Laser Diode Code
405	<3	100	405/100
450	<3	75	450/075
473	<3	70	473/070 [†]
488	<3	50	488/050 [†]
520	<3	60	520/060
638	<3	80	638/080
638	<3	120	638/120

Table 7:	Connec	torized	Laser	Diode	Modules	Codes
----------	--------	---------	-------	-------	---------	-------

ORDERING CODE: CLDM_000/000

Laser diode code (see Table 7) —

^{*}Power coupled into 50 µm core, NA 0.22 optical fiber

[†]The unit prices of the 473 nm and 488 nm laser diode modules are significantly higher.



Connectorized Laser Diode Module male pinout

Note: A Connectorized Laser Diode Module does not include the corresponding Laser Diode Module Driver. See Table 8 for available Laser Diode Module Driver models.

Laser Diode Module Drivers

The Laser Diode Module Driver available in 1-, 2- and 4-channel models is controlled manually or by a computer via USB. Each channel has a BNC input connector for up to 10 kHz TTL/analog modulation of the driving current and a BNC output connector for monitoring the driving current or for the synchronization with other devices. Doric drivers laser safety features include a rear panel interlock connector, a master key switch and



Laser Diode Module Driver: 2-channel model

white LED illuminated control knobs indicating laser diode operation. Unlike most commercial laser diode drivers, our linear driving electronics eliminates the leakage current and the corresponding residual light output when the current is set to zero. In order to avoid unwanted channel activity in optogenetics experiments it is of crucial importance to eliminate any light output when the driving current is set to zero. The *Laser Diode Module Driver* recognizes *Connectorized Laser Diode Modules* and automatically sets the corresponding maximum driving currents, thus preventing accidental overdrive.



Laser Diode Module Driver female pinout

Number of channels	Ordering Code
1	LDMD_1
2	LDMD_2
4	LDMD_4

Table 8: Laser Diode Module Drivers Ordering Codes

Laser Diode Fiber Light Sources



Laser Diode Fiber Light Source: 2-channel model

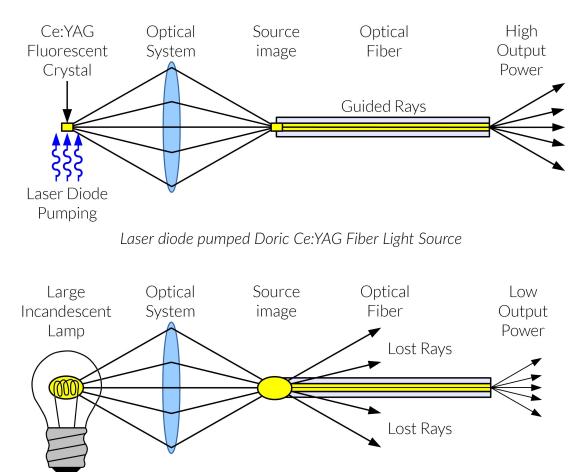
The Laser Diode Fiber Light Source is a more compact alternative to the combination of the Connectorized Laser Diode Modules and Laser Diode Module Drivers. Available in 1-, 2- and 4-channel models, the source is fully compatible with Doric free operating software. Each channel has a BNC input connector for up to 10 kHz TTL/analog modulation of the driving current and a BNC output connector for monitoring the driving current or for synchronization with other devices. Its laser safety features include a rear panel interlock connector, a master key switch and white LED illuminated control knobs indicating laser diode operation. Also, each FC/APC optical connector has a metal dust cap that acts as protective mechanical shutter in absence of optical fiber. Unlike most commercial laser diode drivers, our linear driving electronics eliminates leakage current and the corresponding residual light output when the current is set to zero. For optogenetics experiments it is of crucial importance to eliminate any light output when the driving current is set to zero. The available wavelengths and fiber-coupled power values are given in the table below. For multichannel models, any wavelength combination can be chosen at time of ordering.

ORDERING CODE:

1-channel model	
2-channel model	
4-channel model Laser diode codes	
(see Table 7)	

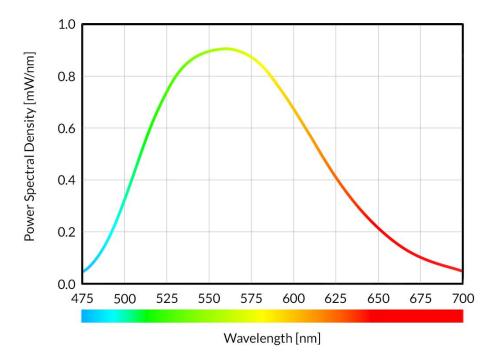
Ce:YAG Fluorescent Illumination

As LED lighting made it obvious, white light can be generated by blue LED pumping of phosphors or fluorescent crystals such as Cerium-doped YAG crystals (Ce:YAG). However, the relatively large emitting area of blue LEDs and their highly divergent light beams result in a fluorescent light source of very large optical etendue (emitter area times light beam divergence) unsuitable for effective fluorescence coupling into small core optical fibers. Optogenetics and other life science applications require tens of milliwatts of suitable bandwidth into the small core diameter of optical fibers. Consequently, we designed fluorescent light sources, called the *Ce:YAG Fiber Light Sources*, in which a Ce:YAG crystal is pumped over a very small area with multiple high-power blue laser diodes instead of LEDs. As shown in the figure below, this patent pending laser diode pumping geometry creates a small area fluorescence light emitter. This is optimized for efficient coupling into the small core diameter of optical fibers, unlike the LED based light sources and other technologies such as arc lamps and incandescent lamps.



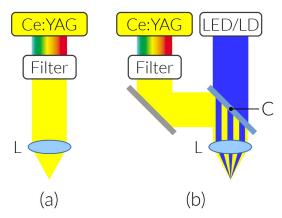
Conventional fiber light source

The Ce:YAG Fiber Light Source emits incoherent light in the green-yellow-red part of the spectrum (see the figure below) with brightness levels far exceeding those of LED based light sources. Unlike lasers, the output of the Ce:YAG Fiber Light Source is speckle-free due to the incoherent nature of fluorescence. Also, the Ce:YAG Fiber Light Source can be electronically modulated through its pumping laser diodes without the noisy intensity spiking encountered with most diode-pumped solid state (DPSS) lasers emitting in the same spectral range.



Output power spectral density (PSD) of a Ce:YAG Fiber Light Source using a 200 μm, 0.53 NA optical fiber. Power and irradiance specifications are respectively given in Table 13 and 14.

The optical head of the Doric Ce:YAG Fiber Light Source is offered in two models schematically shown in the figure on the right: (a) the Ce:YAG Optical Head and (b) the Ce:YAG + LED Optical Head or the Ce:YAG + Laser Diode Optical Head. For both models, a removable filter (see figure) can select the wavelength range within the broad emission band of the Ce:YAG fluorescence. Standard bandpass optical filters are given in Table 11 with their corresponding ordering codes. The optical head model shown in (b) includes a dichroic beam combiner C and a blue light source which is either a 465 nm LED, a 450 nm laser diode or a 473 nm laser diode. When combined with an LED or a LD source, the Ce:YAG source and the LED or LD can be modulated independently using Doric Ce:YAG Drivers. For both optical head models, a fiber coupling lens L focus the output beam into an FC receptacle for optimum fiber coupling.



Schematic representation of (a) the Ce:YAG Optical Head and (b) the Ce:YAG + LED Optical Head or Ce:YAG + Laser Diode Optical Head

Optical Heads of the Ce:YAG Fiber Light Source

A Ce:YAG Fiber Light Source is an optical head and an electronic driver linked by an HDB15 cable (see the section Drivers of the Ce:YAG Fiber Light Source). Optical heads of Ce:YAG Fiber Light Sources are optimized for optical fibers core diameters of 200 μ m to 400 μ m and numerical aperture NA = 0.53. The fiber-coupled output power increases with the core diameter up to about 600 μ m.

The optical output is thus well optimized for unilateral and bilateral activation/silencing in optogenetics experiments and for Doric Optogenetically Synchronized Fluorescence Microscopy Systems. A Ce:YAG optical head is also included with each 2-color Fluorescence Microscope System (using a different driver).



Ce:YAG Optical Head

The optical specifications of Ce:YAG optical heads are given in Table 9.

			Т	TYPICAL OUTPUT POWER (mW)				
	Central Wavelength (nm)	Bandwidth FWHM (nm)	Core 100 μm (0.22 NA)	Core 200 μm (0.53 NA)	Core 400 μm (0.53 NA)	Core 960 μm (0.63 NA)		
	Full spectrum	~110	8.6	94	230	350		
	525	~30	2.1	21	55	94		
ט	559	~34	2.7	29	72	108		
Ce:YAG	582	~75	4.7	51	125	181		
U I	593	~40	2.5	28	67	93		
	612	~69	3.0	33	79	112		
	475	07		E 4*	1.0*	40*		
LED	465	~27	-	5.4*	13*	43*		
LD	450	<3	75	75	75	75		
LD	473	<3	70	70	70	70		

Table 9: Typical Ce:YAG Optical Heads Output Power (mW) vs Optical Fiber Core Diameter, NA

^{*}Specifications in continuous (cw) mode. In overdrive mode, the LED output power is multiplied by \sim 1.7.

Table 10: Ce:YAG Optical Heads Ordering Codes



Laser wavelength (nm) – **450** or **473** nm

Notes:

- A *Ce*:YAG *Optical Head* does not include the corresponding Ce:YAG Driver. See Table 12 for available Ce:YAG Driver models.
- Each Ce:YAG Optical Head is delivered with an empty Filter Holder for Ce:YAG Fiber Light Source (**YFH**; see Table 116). The available *Bandpass filters for Ce*:YAG Fiber Light Sources (**YBPF**) are presented in Table 11.

Bandpass Filters for Ce:YAG Fiber Light Sources

Each Ce:YAG Optical Head is delivered with an empty Filter Holder for Ce:YAG Fiber Light Source (**YFH**; see Table 116). This holder can accept up to 5 mm thick filters of 25 or 25.4 mm diameter. Doric standard *Bandpass filters* are sold already mounted in a filter holder (**YBPF**, see Table 11).



Bandpass Filter for Ce:YAG Fiber Light Sources in its holder

Table 11: Bandpass filters for Ce:YAG Optical Heads

Ordering Code	Bandwidth FWHM (nm)	Central Wavelength (nm)
YBPF_525/030	~30	525
YBPF_549/015	~15	549
YBPF_559/034	~34	559
YBPF_582/075	~75	582
YBPF_593/040	~40	593
YBPF_612/069	~69	612

Drivers of the Ce:YAG Fiber Light Source

All *Ce:YAG Driver* models can be controlled manually or using a computer via a USB port and Doric Neuroscience Studio Software. Drivers are offered in 3 models shown in Table 12. All models include a first channel for controlling the Ce:YAG source driving current. For Ce:YAG Optical Heads including an internal blue light source, either an LED or a laser diode (LD), the corresponding drivers include a second channel for the blue source. In these cases, both channels are controlled independently through software defined sequences or using the BNC input connector of each channel for an external control by analog or TTL signals. Each channel also includes a BNC output connector proportional to the driving current. This output signal can be used for the synchronization of other devices. Doric *Ce:YAG Drivers* safety features include a rear panel interlock connector, a master key switch and, for each channel, a white LED illuminated knob indicating if the corresponding source is activated. Unlike most commercial drivers, Doric driving electronics eliminates the leakage current and the corresponding light output when the current is set to zero. This is of crucial importance for optogenetics experiments.



Table 12: Ce:YAG Drivers Ordering Codes

Notes:

- A *Ce*:YAG *Driver* does not include the corresponding Ce:YAG Optical Head. See Table 10 for available Ce:YAG Optical Head models.
- The Ce:YAG + Laser Diode Driver is compatible with the Ce:YAG + 450 nm Laser Diode Optical Head and the Ce:YAG + 473 nm Laser Diode Optical Head.

		TYPICAL OUTPUT POWER (mW)								
	λ (nm)	$\Delta\lambda$ (nm)	Source	50 μm (0.22 NA)	100 μm (0.22 NA)	200 μm (0.53 NA)	400 μm (0.53 NA)	960 μm (0.63 NA)	Ordering Code	
	365	~12	LED	-	-	6.0	23	100	CLED_365	
	385	~12	LED	-	-	6.0	23	100	CLED_385	
	405	<3	LD	100	100	100	100	100	CLDM_405/100	
	405	~15	LED	-	-	5.0	23	100	CLED_405	
	420	~15	LED	-	-	5.5	23	100	CLED_420	
	450	<3	LD	75	75	75	75	75	CLDM_450/07	
	450	~25	LED	-	-	8.0	23	100	CLED_450	
	465	~25	LED	-	-	7.5	23	100	CLED_465	
	473	<3	LD	70	70	70	70	70	CLDM_473/070	
	488	<3	LD	50	50	50	50	50	CLDM_488/05	
	505	~30	LED	-	-	3.0	12	50	CLED_505	
	515	~40	LED	-	-	3.0	9.5	40	CLED_515	
	520	<3	LD	60	60	60	60	60	CLDM_520/060	
	525	~30	Ce:YAG	-	2.1	21	55	94	Ce:YAG_525/03	
	550	~110	Ce:YAG	-	8.6	94	230	350	Ce:YAG_550/00	
	559	~34	Ce:YAG	-	2.7	29	72	108	Ce:YAG_559/03	
	560	~100	LED	-	-	2.0	8.5	40	CLED_560	
	582	~75	Ce:YAG	-	4.7	51	125	181	Ce:YAG_582/07	
	593	~40	Ce:YAG	-	2.5	28	67	93	Ce:YAG_593/04	
	595	~20	LED	_	-	2.0	8.5	40	CLED_595	
	612	~69	Ce:YAG	-	3.0	33	79	112	Ce:YAG_612/06	
	625	~20	LED	-	-	3.5	14	70	CLED_625	
	635	~20	LED	-	-	6.5	25	100	CLED_635	
	638	<3	LD	80	80	80	80	80	CLDM_638/080	
	638	<3	LD	120	120	120	120	120	CLDM_638/120	
	840	~35	LED	-	-	6.0	22	40	CLED_850	
	940	~35	LED	-	-	2.0	10	40	CLED_940	
5	500K	-	LED	_	_	4.5	17	80	CLED_W55	

Table 13: Typical Light Sources Output Power vs Optical Fiber Core Diameter

	TYPICAL INTENSITY at FIBER TIP (mW/mm ²)							
Ordering Code	960 μm (0.63 NA)	400 μm (0.53 NA)	200 μm (0.53 NA)	100 μm (0.22 NA)	50 μm (0.22 NA)	Source	$\Delta\lambda$ (nm)	λ (nm)
CLED_365	138	183	190	-	-	LED	~12	365
CLED_385	138	183	190	-	-	LED	~12	385
CLDM_405/100	140	800	3 200	13 000	51 000	LD	<3	405
CLED_405	138	183	159	-	-	LED	~15	405
CLED_420	138	183	175	-	-	LED	~15	420
CLDM_450/075	100	600	2 400	9 500	38 000	LD	<3	450
CLED_450	138	183	254	-	-	LED	~25	450
CLED_465	138	183	238	-	-	LED	~25	465
CLDM_473/070	97	560	2 200	8 900	36 000	LD	<3	473
CLDM_488/050	69	400	1 600	6 400	25 000	LD	<3	488
CLED_505	69	95	95	-	-	LED	~30	505
CLED_515	55	75	95	-	-	LED	~40	515
CLDM_520/060	83	480	1 900	7 600	31 000	LD	<3	520
Ce:YAG_525/03	130	438	668	267	-	Ce:YAG	~30	525
Ce:YAG_550/00	484	1 830	2 992	1 095	-	Ce:YAG	~110	550
Ce:YAG_559/03	149	573	923	344	-	Ce:YAG	~34	559
CLED_560	55	67	63	-	-	LED	~100	560
Ce:YAG_582/07	250	996	1 633	596	-	Ce:YAG	~75	582
Ce:YAG_593/04	128	533	891	318	-	Ce:YAG	~40	593
CLED_595	55	67	63	-	-	LED	~20	595
Ce:YAG_612/06	155	629	1 050	382	-	Ce:YAG	~69	612
CLED_625	96	111	111	-	_	LED	~20	625
CLED_635	138	198	206	-	-	LED	~20	635
CLDM_638/080	110	640	2 500	10 000	41 000	LD	<3	638
CLDM_638/120	170	960	3 800	15 000	61 000	LD	<3	638
CLED_850	55	175	190	-	-	LED	~35	840
CLED_940	55	79	63	-	-	LED	~35	940
CLED_W55	110	135	143	_	_	LED	-	5500K

Table 14: Typical Light Sources Irradiance vs Optical Fiber Core Diameter

doric

OTPG_8

0

Modulators

Optogenetics methods use light pulses to modulate the activity of genetically engineered light sensitive cells. Long gone are the days when a continuous streak of blue light, sent along an optical fiber to a mouse's brain to make it run, provokes worldwide scientific sensation. These days, even the simplest optogenetics experiments require programmable TTL pulse generators to modulate LED or laser diode drivers and create a desired light pulse train. When a direct modulation of the light source is not possible, as in the case of some solid state lasers, the continuous light beam is modulated using shutters.

Optogenetics TTL Pulse Generators (OTPG)

Our miniaturized TTL Pulse Generators connects to a computer with a USB cable and to a light source driver or a shutter with a BNC cable. They seamlessly integrate with our other optogenetics products. The pulse train parameters and its triggering are controlled via Doric Neuroscience Studio Software with which it is possible to program a sequence at a determined frequency and repeat this sequence several times. The Optogenetics TTL Pulse Generators have 4 input/output BNC and the 8-channel has 4 supplemental output BNC.



Connectorized Mechanical Shutter Heads and Adapters

The modulation of the light signal is essential for optogenetics experiments. The light sources, like LEDs or laser diodes are well-suited for the direct electrical modulation, while DPSS or fiber laser types require external modulation via mechanical shutters or acousto-optic modulators.

The mechanical shutters are more popular with laser based optogenetics set-ups as they are cheaper and better suited for use with multimode fibers. The inconvenience of mechanical shutters is that they require parallel beams of light and subsequent coupling into an optical fiber can be tricky and unstable. To facilitate the use of mechanical shutters, we are providing connectorized adapters for the Stanford Research Systems Model SR475 and the Vincent Associates Uniblitz Model LS-2 shutter heads.

Stanford Research Systems Model SR475 - Shutter Head and Adapter

The Stanford Research System Model SR475 Shutter Head is a highprecision shutter system with minimal vibration and a 4 ms minimum pulse duration. The shutter head of the Stanford Research Systems Model SR475 can not produce pulses duration as short as the Vincent Associates Uniblitz LS-2 shutter head (2 ms) but its level of audible noise is much lower. The Doric Lenses Adapter allows the integration of the shutter with fiber connectorized devices. The adapter can be supplied alone or pre-installed on the shutter head.



Stanford Research Systems Shutter Head -Model SR475 + Doric FC Adapter

Table 15: Stanford Research Systems Model SR475 Shutter Head and Adapter - Specifications andOrdering Code

SPECIFICATION	VALUE
Typical input fiber configuration	200 µm core, NA=0.22
Typical output fiber configuration	200 μm core, NA=0.22
Wavelength range	450 - 650 nm
Collimated beam diameter	~2.0 mm
Coupling efficiency	>75%
Maximum optical power	500 mW
Minimum pulse duration	4 msec
Maximal operating frequency	100 Hz
PRODUCT	Ordering Code
Stanford Shutter Head + Doric FC Adapter Doric FC Adapter only	CMSA-SR475_FC SR475_FOA

Note: The Stanford Research Systems Model SR470 - Shutter Controller is compatible with the Shutter Head and Adapter Model SR475.

Vincent Associates Uniblitz Model LS-2 - Shutter Head and Adapter

The Vincent Associates Uniblitz Model LS-2 Shutter Head is a highprecision shutter system with high repeatability and a 2 ms minimum pulse duration. Despite its higher level of audible noise than the Stanford Research Systems Model SR475, the Uniblitz Model LS-2 is suitable for experiments requiring pulse duration as short as 2 ms. The Doric Lenses Adapter allows the integration of the shutter with fiber connectorized devices. We provide the shutter with the Doric adapter already installed, as precision optical alignment is necessary for optimal usage.



Vincent Associates Uniblitz Shutter Head - Model LS-2 + Doric FC Adapter

Table 16: Vincent Associates Uniblitz Model LS-2 Shutter Head and Adapter - Specifications andOrdering Code

SPECIFICATION	VALUE
Typical input fiber configuration	200 μm core, NA=0.22
Typical output fiber configuration	200 µm core, NA=0.22
Wavelength range	450 - 650 nm
Collimated beam diameter	~2.0 mm
Coupling efficiency	>75%
Maximum optical power	500 mW
Minimum pulse duration	2 msec
Maximal operating frequency	100 Hz
PRODUCT	Ordering Code
Vincent Associates Shutter Head + Doric FC Adapter	CMSA-LS2_FC

Note: The Vincent Associates Uniblitz Model VCM-D1 - Shutter Controller is compatible with the Shutter Head and Adapter Model LS-2.

Connectorized Mechanical Shutter Controllers

Stanford Research Systems Model SR470 - Shutter Controller

The Stanford Research Systems Model SR470 - Shutter Controller is compatible with the Shutter Head and Adapter Model SR475.

ORDERING CODE: MSC_SR470

Vincent Associates Uniblitz Model VCM-D1 - Shutter Controller

The Vincent Associates Uniblitz Model VCM-D1 - Shutter Controller is compatible with the Shutter Head and Adapter Model LS-2.

ORDERING CODE: MSC_VCM-D1



Vincent Associates Uniblitz Model VCM-D1 -Shutter Controller



Systems Model SR470 -Shutter Controller

Beam Splitters/Combiners

As multimode fiber optics is finding wider use in microscopy, optogenetics and life sciences in general, the need to combine or divide the light signals within fiber optic circuits is indispensable. Beam-splitters have been used in optics for many years and almost exclusively within the parallel beam of light and at 45 degrees angle of incidence. Since the light coming out of the optical fiber is divergent, it needs to be made parallel or collimated before the beam-splitters can be used. Combining or splitting of the light output from optical fibers requires good collimation lenses, beam-splitters with steep transition curves and precision positioning to get efficient coupling. Inspired by the microscopy cubes and the need for user friendly beam-splitting in the fiber-optics applications, we have developed a family of mini cubes and multiple splitters that integrate beam-splitting glass plates, collimation lenses and fiber-optic receptacles in a small connectorized or pigtailed packages. Apart from shrinking the size of the so called microscope cubes, we have introduced highly efficient beam-splitters with unprecedented balance of the s and p polarization reflection curves based on our low angle of incidence design.

Doric Mini Cubes

Doric Mini Cubes: Intensity Division

This *Doric Mini Cube* contains a beam splitter that separates a beam in two output beams of equal power. This cube can be used effectively only as a splitter. If used as a combiner the power will not be doubled. The input and output NA is 0.22.

ORDERING CODE: DMC_1x2i_VIS_FC

VIS for 450 to 650 nm^{*} –

Receptacle code[†] —



Doric Mini Cube for Intensity Division

^{*}Other ranges available as custom product †FC is standard, SMA available on request

Doric Mini Cubes: Wavelength Division

The wavelength division mini cube has no other filters except the dichroic mirror which combines or separates different wavelengths. The angle of incidence of the light to the dichroic mirror found inside the standard version of wavelength division *Doric Mini Cubes* is 22.5 degrees. More conventional cubes with a 45 degrees angle of incidence is available only as a custom product. The input and output NA is 0.22.



ORDERING CODE: DMC_1x2w_470/590_FC Wavelength 1 (nm) ______ Doric Mini Cube for separation of 470 nm and 590 nm

Receptacle code^{*} —

Example of custom assembly



Doric Mini Cube for separation of 470 nm and 530 nm band

^{*}FC is standard, SMA available on request

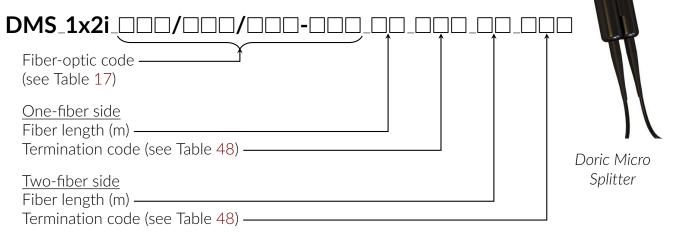
Doric Micro Splitters

To further reduce the body of the bulk optics splitters, they need to be pigtailed rather than connectorized. This product family we call *Doric Micro Splitters*. Their small size and low transmission losses make micro splitters a superior alternative to branching fiber-optic patch cords. When combined with these splitters, the standard FRJ_1x1, HRJ-OL, HRJ-OE and AHRJ rotary joints can be turned into bilateral optical stimulation ready joints. As an illustration of its performance, 1x1 fiberoptic rotary joint combined with *Doric Micro Splitter* has over 30% transmission per channel, less than 5% transmission difference between the channels and less than 5% power variation during rotation. They are second only to 1x2 FRJ. They can be also used in OEM devices whenever the space is limited.

Doric Micro Splitters: Intensity Division

This micro splitter separates an incoming beam into two output beams of equal intensity. Unlike Doric Mini Cube, *Doric Micro Splitter* has input and output fibers on the opposite sides of the device. The standard product is designed for visible light from 450 nm to 650 nm. The input and output NA is 0.22.

ORDERING CODE:



doric

Doric Micro Splitters: Wavelength Division

The wavelength division splitters have the same inside-out appearance as intensity division splitters, the only difference is in their respective dichroic filter. The input and output NA is 0.22.

ORDERING CODE:

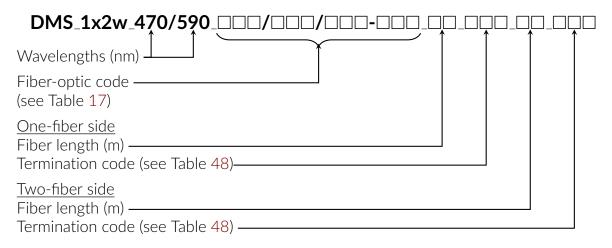


Table 17:	Micro	Solitters	Fiher-0	ntic	Codes
TADIE 17.	MICIO	Spincers	TIDEI-0	ριις	COUES

		Outer diameter (µm)			
Core (µm)	Cladding (µm)	Buffer	Jacket	NA	Fiber-optic Code
100	110	125	900	0.22	100/110/900-0.22
100	110	125	900	0.37	100/110/900-0.37
105	125	250	900	0.22	105/125/900-0.22
200	220	240	900	0.22	200/220/900-0.22
200	220	245	900	0.37	200/220/900-0.37
200	240	400	900	0.22	200/240/900-0.22

Doric Multiple Splitters/Combiners

Light Intensity Distributors

The fiber coupled laser sources typically offer high intensity within a relatively small fiber diameter. When running several simultaneous *in vivo* experiments with such light sources, it makes perfect sense to use the *Light Intensity Distributor* which is basically an intensity splitter. By doing this, the required number of modulation channels, drivers and optical sources can be reduced. Our patent pending *Light Intensity Distributor* provides a compact, connectorized package with the low insertion and polarization dependent loss (PDL), ideal for multimode fibers. The input and output NA is 0.22.



Light Intensity Distributor -4 channels

Table 18: Light Intensity Distributors Ordering Codes

Number of Channels	Ordering Code *
3	LID_1x3_VIS_FC
4	LID_1x4_VIS_FC

Note:

FC is standard. Contact us for custom requests. VIS stands for visible wavelength range from 450 to 650 nm. Other ranges available as custom product. The expected intensity percentage in each channel is typically 80% divided by the number of channels^{*}.

^{*}Our standard products assume the use of identical fiber diameters, receptacles and equal intensity for each channel. However, this can be customized if needed, at extra cost.

Light Spectrum Mixers

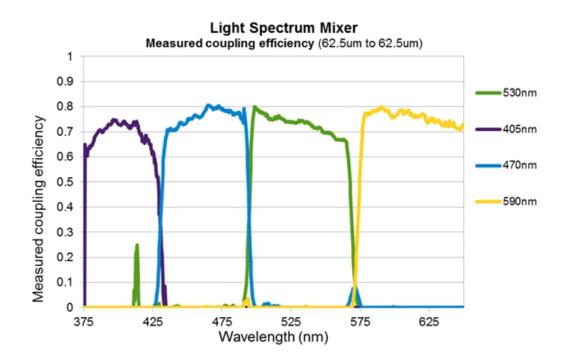
For *in vivo* optogenetics experiments there is a need to illuminate the tissue with specific pulses of spectrally different lights using the same fiber. To put it simply, the light from different fiber coupled LEDs or lasers needs to be combined into one beam and coupled to an optical fiber leading to a fiber-optic implant or cannula. Our patent pending *Light Spectrum Mixer* provides a compact, connectorized package with highly efficient coupling and low polarization dependent loss (PDL), ideal for multimode fibers. The input and output NA is 0.22.

The same device can be used in the opposite direction as a light spectrum separator. The concept of a spectrum mixer or splitter is analog to the concept of a wavelength division multiplexing and demultiplexing in optical telecommunication.



Light Spectrum Mixer

Number of Channels	Ordering Code [*]
3	LSM_1x3_470/530/590_FC
4	LSM_1x4_405/470/530/590_FC



^{*}FC and center wavelengths are standard. Contact us for custom requests.

Beam Modifiers

Filtering

Connectorized U-bracket

The attenuation or spectral filtering of the light within an optical fiber can be achieved with a simple *Connectorized U-bracket* and specific filter insert. To prevent dust entering the device, we recommend closing it with a filter insert at all times. For maximum transmission you can use the insert without a filter. For blocking the light use the insert without a hole. Unless some light loss is tolerated, it is necessary that NAs and diameters of input and output fibers are the same. The U-bracket comes with a blocking insert and an insert with hole but no filter. The specific filters have to be ordered separately.



Connectorized U-bracket and Filter Insert

Max fiber NA

Receptacle code^{*} ———

U-bracket Inserts

The inserts can be fitted with attenuating filters or spectral filters made from a variety of glass materials. As a matter of fact, we can fit any commercially available filter to our standard insert and engrave its code. In this way you can build your set using off-theshelf or custom filters. The narrow band filters can be useful for filtering the fluorescence excitation spectrum or for the fluorescence light.

ORDERING CODE: UBI

Example code: UBI_Semrock_FF01-474/23-25, UBI_Chroma_ET470/40x



Filter Insert

^{*}FC is standard, SMA available on request

NA Converter

Laser sources are valued for the large amount of power they deliver. However, one of their characteristics is that the beams they produce have small divergences. This can be a limitation for those who require a powerful illumination over a wide angle. To adress this issue, we have developped the *NA Converter*, that modifies the geometry of an input fiber guided light beam. Both the numerical aperture and the beam diameter are affected: their product is a constant, known as Lagrange invariant.



NA Converter



NA 2X magnification = Beam diameter 0.5X magnification

The typical application in optogenetics is when the laser source is coupled to an 0.22 NA fiberoptic while the fiber-optic cannula of interest is made of a fiber-optic with 0.48 NA. In this case, a magnification of x2 is well suited. In this example, if no NA converter is used, the fiber-optic cannula NA is not filled, and its output beam has an NA that is roughly 0.22.

ORDERING CODE: NAC	
Input NA	
Output NA ————	
Receptacle code [*] ————	

^{*}FC is standard, SMA available on request

Rotary Joints

Fiber-optic Rotary Joints

Fiber-optic Rotary Joints consist of a lens system and high precision bearings which allow a rotationinsensitive optical power transfer between optical fibers. The fixed part of the rotary joint allows the connection to a light source and the rotating part releases the twisting of the optical fiber connected to the animal. In neurosciences, freely-moving optogenetics experiments need a stable light input to the brain even if the animal is moving in a confined space. Fiber-optic Rotary Joints avoid the damaging of the optical fibers while minimizing light fluctuations when rotating. The nomenclature used for our rotary joints is FRJ_m X n where m and n represent the number of the fibers on the fixed and on the rotating side respectively.

1x1 Fiber-optic Rotary Joints

The 1x1 Fiber-optic Rotary Joint is the basic and the most popular type of rotary joints. It can either transmit the light from the sources to the sample and/or from the sample to a photodetector. When fiber-optic patch cord connectors are inserted in the rotary joint receptacles, the fiber tips are at the focal planes of the respective collimating lenses, and the beam is parallel between the lenses. 1x1 Fiber-optic Rotary Joints are typically used with optical fibers with a core diameter of 200 µm and an NA of up to 0.5.

Notes:

- The compatible holder for the 1x1 Fiber-optic Rotary Joints is sold separately (Holder_FRJ_small; see Table 117).
- The output fiber-optic patch cords are sold separately.
- An optional gimbal holder allows pivoting the rotary joint along two additional axes, further reducing the mechanical stress on the animal (**GH_FRJ**; see Table 119).



1x1 Fiber-optic Rotary Joint

SPECIFICATION	VALUE
Transmission*	> 85%
Maximum variation	\pm 3% of the mean
Start up torque	20 µN∙m
Input NA	up to 0.5
Output NA	up to 0.5
Optimized for 62.5 μm^{\dagger}	Ordering Code
No	FRJ_1x1_FC-FC
110	
Yes	FRJ_1x1_FC-FC_62.5
	FRJ_1x1_FC-FC_62.5
	FRJ_1x1_FC-FC_62.5
	FRJ_1x1_FC-FC_62.5

Table 20: 1x1 Fiber-optic Rotary Joints Specifications and Ordering Codes

Pigtailed 1x1 Fiber-optic Rotary Joints

Fiber photometry experiments detect small power variations from a fluorophore and for that reason the fiber-optic rotary joints within the setup require minimal transmission variation. Because of large core multimode fibers and connector tolerances (i.e. 400 μ m NA 0.48), transmission variation can only be minimized using a pigtailed version of 1x1 fiber-optic rotary joint. The pigtailed patch cords are made from 0.37 or 0.57 NA, 200 or 400 μ m diameter optical fiber with a lightweight metal jacket and FC connectors. The fixed input patch cord is 1 m long, while the output or rotating patch cord is 0.15 m long. Different length fiber-optic patch cords can be connected to the output using an FC/FC mating adapter.

Notes:

- The compatible holder for the *Pigtailed* 1x1 *Fiber-optic Rotary Joints* is sold ¹⁵ cm separately (**Holder_FRJ_small**; see Table 117).
- An optional gimbal holder allows pivoting the rotary joint along two additional axes, further reducing the mechanical stress on the animal (GH_-FRJ; see Table 119).
- An compatible FC/FC mating adapter (**ADAPTER_FC**; see Table 122) is sold separately and can be used to connect different patch cords to the optical fibers already linked to the rotary joint.

Pigtailed 1x1 Fiber-optic Rotary Joint

Tested with 200 μ m core NA 0.22 fiber-optic patch cords.

^{\dagger}Ideal for use with fiber-optic core from 62.5 μ m to 200 μ m. It is highly recommended to use our patch cords with these rotary joints to get appropriate coupling efficiency.

SPECIFICATION	VALUE
Transmission [*]	> 70%
Maximum variation	<1% peak-to-peak
Start up torque	20 μN·m
Input Fiber	200 or 400 μm core - NA 0.37 or 0.57
Output Fiber	200 or 400 μm core - NA 0.37 or 0.57

Table 21: Pigtailed 1x1 Fiber-optic Rotary Joints Specifications

ORDERING CODE:

F	RJ_1x1_PT// 1.0_F(Output optical fiber 1.0_F(200/220/LWMJ-0.37 1.0_F(200/230/LWMJ-0.57 1.0_F(400/430/LWMJ-0.57 1.0_F(400/440/LWMJ-0.37 1.0_F(ÇM_0.	15_FÇ	M
	Input fiber length (m) ———————————————————————————————————			
	Input receptacle code — FCM is standard (see Table 48).			
	Output fiber length (m) — 0.15 m is standard.]	
	Output receptacle code FCM is standard (see Table <u>48</u>).]

⁴¹

^{*}Tested with 400 μm core NA 0.48 fiber-optic patch cords.

1x2 Fiber-optic Rotary Joints

These rotary joints are used to divide the light coming from a single input optical fiber on a fixed side to two output optical fibers on a rotating side. We offer two distinct versions of this product, one for the intensity division and the other for the wavelength division of the light. Each version can be further customized if needed.

Notes:

- A compatible holder is included with the 1x2 Fiber-optic Rotary Joints (Holder_FRJ_large; see Table 117).
- The output fiber-optic patch cords are included.
- An optional gimbal holder allows pivoting the rotary joint along two additional axes, further reducing the mechanical stress on the animal (GH_FRJ; see Table 119)

Intensity division

The intensity division rotary joint sends half of the input light into each of the two output receptacles. This is particularly useful for bilateral stimulation experiments, where the illumination intensities must be the same in each channel.



1x2 Fiber-optic Rotary Joint -Intensity division

Table 00, 1,2 Fiber antis Datary	Lainte Intensity division	Considerations and	Ordering Codes
Table 22: 1x2 Fiber-optic Rotary	Joints - Interisity division	specifications and	Ordening Codes

	SPECIFICATION	VALUE
	Transmission [*] Maximum variation Start up torque Input NA	> 40% per channel ± 3% of the mean 30 μN⋅m 0.22
	Output NA	Ordering Code
	0.22 0.5	FRJ_1x2i_FC-2FC_0.22 FRJ_1x2i_FC-2FC_0.50
Input receptacle code		
Output receptacles co	de ———	

^{*}Tested with 200 μ m core NA 0.22 fiber-optic patch cords.

Wavelength division

The *wavelength division* rotary joint splits the spectral band originating from the input receptacle and sends each band to the corresponding rotating fiber receptacles. In some optogenetics experiments, it can be used for instance to separate the 473-488 nm blue light (activation signal) and the 590 nm orange light (inhibition signal). This rotary joint can also be used in the opposite direction as a spectral combiner.



1x2 Fiber-optic Rotary Joint - Wavelength division

Table 23: 1x2 Fiber-optic Rotary Joints - Wavelength division Specifications and Ordering Codes

SPECIFICATION	VALUE
Transmission* Maximum variation Start up torque Input NA	> 75% for each spectral band ± 3% of the mean 30 μN·m 0.22
Output NA	Ordering Code
0.22 0.5	$\begin{array}{c} FRJ_1x2w_\Box\Box/\Box\Box_FC-2FC_0.22\\ FRJ_1x2w_\Box\Box/\Box\Box_FC-2FC_0.50\\ \uparrow & \uparrow \\ \end{array}$
Output wavelengths (nm) ——— Connector A / Connector B	
Input receptacles code ————	
Output receptacle code ———	

Separate Light Path 2x2 Fiber-optic Rotary Joints

Separate Light Path 2x2 Fiber-optic Rotary Joints connect two arbitrary fiber-optic types on the stationary side of the rotary joint with their respective counterparts on the rotating side. This innovative patent pending technology offers unprecedented possibilities for laser or LED based optogenetics lighting requiring a compact and low loss dual channel fiber-optic rotary joint. This Separate Light Path 2x2 Fiber-optic Rotary Joint makes possible optogenetics and photometry experiments with an independent control of two different sites of illumination and/or detection of the light.



Notes:

- A compatible holder is included with the Separate Light Path 2x2 Fiber-optic Rotary Joints (Holder_FRJ_2x2; see Table 117).
- Two output fiber-optic patch cords are also included.

Separate Light Path 2x2 Fiber-optic
Rotary Joint

Table 24: Separate Light Path 2x2 Fiber	r-optic Rotary Joints Specifications
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SPECIFICATION	VALUE
Transmission*	> 80% for each channel
Maximum variation	\pm 3% of the mean per channel
Start up torque	< 3mN⋅m [†]
Input NA	0.22
Output NA	0.22

ORDERING CODE: FRJ_2x2_VIS_2FC-2FC

Wavelength range —

Input receptacles code —

Output receptacles code —

^{*}Tested with 200 μ m core NA 0.22 fiber optic patch cords.

[†]Start up torque too high for mice but acceptable for rats or larger animals.

1x4 Fiber-optic Rotary Joints

The 1x4 Fiber-optic Rotary Joint is used to send the light coming from a single optical fiber to 4 different regions on a moving animal via separate optical fibers. The fixed side consists of an FC receptacle and the rotating side of the joint is a 4-way optical connector specially developed for this application. A patch cord with a specific small footprint four-fiber connector designed to minimize the size and the inertia of the rotor is essential to the use of this rotary joint.

Notes:

- A compatible holder is included with the 1x4 Fiber-optic Rotary Joint (Holder_FRJ_large; see Table 117).
- One four-fold branching output patch cord is included. Contact us if spare patch cords are required to connect at the bottom of your 1x4 Fiber-optic Rotary Joint.
- An optional gimbal holder allows pivoting the rotary joint along two additional axes, further reducing the mechanical stress on the animal (**GH_FRJ**; see Table 119).



1x4 Fiber-optic Rotary Joint

SPECIFICATION	VALUE
Transmission*	20% per channel (-2% as function of used fiber)
Maximum variation	\pm 2% of the mean per channel
Start up torque	< 50 μN·m
Input NA	0.22
Output NA	0.22

ORDERING CODE: FRJ_1x4i_FC

Input receptacles code —

 $^{^*\}text{Tested}$ with 200 μm core NA 0.22 Fiber-optic Patch Cords.

Electrical Rotary Joints

Electrical Rotary Joints are used to transmit electrical signal from a moving sample to a fixed recording system (*e.g.* for *in vivo* electrophysiology experiments). Since it can be desirable to couple electrophysiological experiments with optogenetics stimulations, our Electrical Rotary Joints are designed with a central aperture (hollow bore) allowing the insertion (pass-through) of a fiber-optic patch cord. In this case, electrical and fiber-optic rotary joints (1x1 or 1x2) are used in tandem.

Electrical Rotary Joints

We have developed a passive *Electrical Rotary Joint* usable for electrophysiological experiments that can be combined with fiber-optic rotary joints (1x1 or 1x2) to bring light to and/or from the sample. Its 7.2 mm through-hole in the center is sufficient for passing fiber-optic patch cords with M3 connectors or ferrule/sleeve type connectors. It is also convenient for fluid tubing allowing drugs administration during electrophysiological experiments with freely-moving animals.

Our *Electrical Rotary Joint* has a torque as low as 0.9 mN·m (for 6 electrical contacts) or 1.8 mN·m (for 12 electrical contacts), acceptable for use with rats or larger animals. They are optimized to offer the best electric signal with the lowest torque, given that stable electrical transmission with small resistivity variations during rotation requires the increase of



Electrical Rotary Joint with a HDMI Connector

contact areas between each electrical contact. For small animals like mice, we recommend our *Assisted Electrical Rotary Joints* to remove the torque originating from the friction of the electrical contacts.

Notes:

- The number of electrical contacts does not necessarily equal the number of recording channels.
- Holders allowing the mounting of an *Electrical Rotary Joint* with a *Fiber-optic Rotary Joint* (1x1 or 1x2) are included (Holder_ERJ, Holder_FRJ_small and Holder_FRJ_large; see Table 118).
- If the *Electrical Rotary Joints* is used only for electrophysiology, without any additional fiberoptic rotary joints, a compatible holder is already included (**Holder_FRJ_large**; see Table 117).
- An optional horizontal cable holder keeping cables off-center can be added to increase the effective torque applied on the rotor (**HCH**; see Table 119). *Electrical Rotary Joints* come with the pre-installed adapter allowing the fixing on the optional horizontal cable holder.
- An optional gimbal holder allows pivoting the rotary joint along two additional axes, further reducing the mechanical stress on the animal (**GH_FRJ**; see Table 119).
- An optional adapter kit can be provided to allow a Harwin-connectorized rotary joint to be used with an Omnetics-connectorized systems (**ADAPTER_HO12**; see Table 120).

SPECIFICATION	VALUE
Number of contacts	6 or 12
Contact material	Gold
Maximum current	2 A per contact
Start up torque	0.9 mN⋅m (for 6 contacts)
	1.8 mN⋅m (for 12 contacts)
Contact resistance	$<$ 500 m Ω
Resistance variation during constant rotation	$< 100~{ m m\Omega}$ @ 5 VDC
Rotation speed	up to 300 rpm

 Table 26: Electrical Rotary Joints Specifications

Table 27: HDMI Electrical Connector Pinout for Non-assisted Rotary Joints

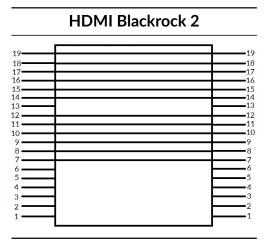


Table 28: Electrical Rotary Joints Ordering Codes

Connector type	Number of electrical contacts	Ordering Code
HARWIN [*]	6	ERJ_06_HARW
	12	ERJ_12_HARW
HDMI Blackrock pinout 2 (see table 27)	12	ERJ_12_HDMI-B2

Assisted Electrical Rotary Joints

The Assisted Electrical Rotary Joint is a rotary joint that transmits electrical signal over 12 or 24 channels. Compared to our passive Electrical Rotary Joint, the motor Assisted Electrical Rotary Joint is effectively frictionless, thus allowing its use with mice. The rotary joint is primarily used in electrophysiology with small animal subjects.

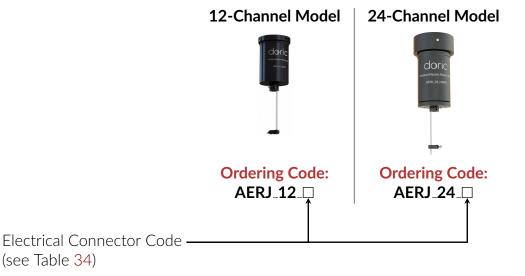
Notes:

- If the Assisted Electrical Rotary Joint is used only for electrophysiology, a compatible holder is also included (See Table 117).
- Holders allowing the mounting of a 12-channel Assisted Electrical Rotary Joint with a Fiberoptic Rotary Joint (1x1 or 1x2) are included (Holder_AERJ, Holder_FRJ_small and Holder_-FRJ_large; see Table 118).
- The torque sensor included with the *Assisted Electrical Rotary Joints* is also used as a cable holder with its rod and its clamp. The adapter to secure the rotary joint on the sensor is pre-installed.

SPECIFICATIONS	VALUE	
Start-up Torque	< 20 µN⋅m	
Electrical Contact Material	Go	ld
Number of Contacts	12 24	
Contact Resistance	$<$ 500 m Ω	$<$ 800 m Ω
Resistance Variation during rotation	$< 100~{ m m}\Omega$	$<$ 25 m Ω
Rotation Speed	< 300 RPM	< 60 RPM
Through-hole diameter	6.0 mm	4.7 mm

Table 29: Assisted Electrical Rotary Joint Specifications





Assisted Electrical Rotary Joint for Fluorescence Mini Cube

The Assisted Electrical Rotary Joint for Fluorescence Mini-Cube is an assisted electrical rotary joint designed to allow the direct integration of fluorescence mini cubes on its rotor. This reduced patch cord length between animal subject and mini cube maximizes light recovered. These are primarily used for fiber photometry or any application requiring the detection of very low-intensity light.

Notes:

- The holder for the Assisted Electrical Rotary Joint for Fluorescence Mini Cube is integrated directly onto the rotary joint.
- This rotary joint is always shipped with at least one *Ro-tary Fluorescence Mini Cube*, as well as any required *Fluorescence Detector Amplifiers*.
- The torque sensor included with the Assisted Electrical Rotary Joints is also used as a cable holder with its rod and its clamp. The adapter to secure the rotary joint on the sensor is pre-installed.
- The electrical rotary joint can be adapted to include the functions of an optical rotary joint. This is available on request.



Assisted Electrical Rotary Joint for Fluorescence Mini Cube

SPECIFICATIONS	VALUE
Start-up Torque	< 20 µN∙m
Light source Connectors	6xM8
Detector Connectors	4xM5
Number of Contacts	24
Contact Resistance	$<$ 800 m Ω
Resistance Variation during rotation	$<$ 25 m Ω
Rotation Speed	< 60 RPM

Table 31: Assisted Electrical Rotary Joint for Fluorescence Mini Cube Specifications

ORDERING CODE: AERJ_24_FMC

Fiber-optic & Electric Rotary Joints

The electrical rotary joints have long been used for *in vivo* electrophysiology recordings. The arrival of optogenetics in neurosciences created the need of rotary joints allowing optical stimulations and electrophysiological recordings. This combination requires an opto-electric hybridization in the connecting cables and the rotary joints.

Fiber-optic & Electric Rotary Joint

To facilitate *in vivo* experiments combining the light stimulation and electrophysiological recordings in optogenetics experiments, we have developed a passive low torque hybrid rotary joint with a number of electrical channels and one optical channel. The FC receptacles on both ends of the rotary joint allow the connection of the input and output fiber-optic patch cords. This product is more compact than the combination of the electrical rotary joint and the 1x1 fiber-optic rotary joint where the optical fiber is passed through the central hole of the electrical joint.



Fiber-optic & Electric Rotary Joint with a HDMI connector

Notes:

- The holder for the *Fiber-optic & Electric Rotary Joint* is included (**Holder_FRJ_**large; see Table 117).
- An optional horizontal cable holder keeping cables off-center can be added to increase the effective torque applied on the rotor and help the rotation (**HCH**; see Table 119). The *Fiber-optic & Electric Rotary Joints* come with the pre-installed adapter allowing the fixing on the optional horizontal cable holder.
- An optional gimbal holder allows pivoting the rotary joint along two additional axes, further reducing the mechanical stress on the animal (GH_FRJ; see Table 119).
- The output fiber-optic patch cords are sold separately.
- An optional adapter kit can be provided to allow a Harwin-connectorized rotary joint to be used with an Omnetics-connectorized systems (ADAPTER_-HO12; see Table 120).

SPECIFICATION	VALUE
Transmission [*]	80%
Maximum variation	2%
Start up torque	0.9 mN·m (for 6 contacts)
	1.8 mN·m (for 12 contacts)
Input NA	0.22
Output NA	0.22
Number of contacts	6 or 12
Contact material	Gold
Maximum current	2 A per contact
Contact resistance	$< 500 \ { m m}\Omega$
Resistance variation during rotation (constant rotation)	$< 100~{ m m}\Omega$ @ 5 VDC
Rotation speed	up to 300 rpm

Table 32: Fiber-optic & Electric Rotary Joints Specifications

Table 33: Fiber-optic & Electric Rotary Joints Ordering Codes

Connector Type	Number of electrical contacts	Ordering Code
HARWIN [†]	6	HRJ-OE_FC_06_HARW
	12	HRJ-OE_FC_12_HARW
HDMI Blackrock pinout 2 (see table 27)	12	HRJ-OE_FC_12_HDMI-B2

 $^{^*} Tested with 200 \ \mu m$ core NA 0.22 Fiber-optic Patch Cords. $^+ HARWIN$ 12 will be sold while stock lasts.

Assisted Fiber-optic & Electric Rotary Joint

The Assisted Fiber-optic & Electric Rotary Joint is electrically driven as it <u>senses and follows</u> the tethered animal's rotations. It detects the torsion of the optical cable during animal movement and releases it with a very high sensitivity. The assistance of this rotary joint helps to counter the frictional force of the internal slip-ring and offers quality transmission of electrical signal during any experiment with freely-moving small animals like mice. It comes with 12 or 24 electrical channels and one optical channel.

Notes:

- The 12-channel Assisted Fiber-optic & Electric Rotary Joints can be designed with two types of lenses. The 24-channel Assisted Fiber-optic & Electric Rotary Joints uses the aspheric (AH) lenses exclusively.
 - The achromatized doublets (AD) allow a near-equal focal distance for wavelengths between 450 nm and 650 nm, minimizing chromatic aberration. The AD models are designed for use with a 200 μ m core, 0.22 NA optical fiber.
 - The 0.50 NA aspheric (AH) is optimized for reduced optical aberration. It is designed for use at a wavelength of 470 nm.
- The holder for the Assisted Fiber-optic & Electric Rotary Joint is included (See Table 117).
- The output fiber-optic patch cords are also included.
- An optional adapter kit can be provided to allow a Harwin-connectorized rotary joint to be used with an Omnetics-connectorized systems (ADAPTER_-HO12; see Table 120).

Connector Type	# of Contacts	# of Channels	Maximum Current	Ordering Code
HARWIN [*]	12	12	2 A	HARW
HDMI Microscope	12	12	0.5 A	HDMI
HDMI Blackrock 2	12	12	0.5 A	HDMI-B2
USB-C	12	12	0.25 A	USB-C
HDMI	24	19	0.5 A	USB-C
USB-C	24	22	0.25 A	USB-C

Table 34: Assisted	Rotary Joint	Electrical	Connector	Codes

*Available while stocks last

Table 35: Assisted Opto-electric Rotary Joints

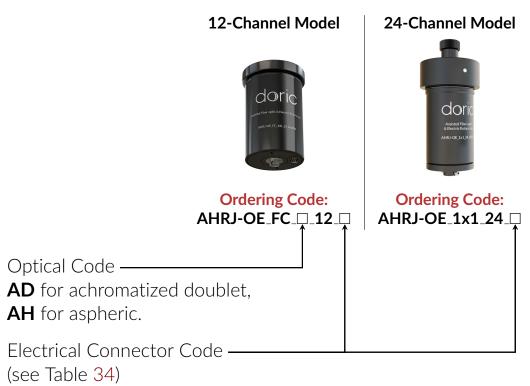


Table 36: Assisted Fiber-optic & Electric Rotary Joint Specifications

SPECIFICATIONS	VAL	UE
Start-up Torque	< 20 µN⋅m	
Input/Output NA	up to 0.5	
Electrical Contact Material	Gold	
Number of Contacts	12	24
Contact Resistance	$<$ 500 m Ω	$<$ 800 m Ω
Resistance Variation During Rotation	$< 100 \ { m m}\Omega$	$< 25 \ { m m}\Omega$
Transmission [*]	75%	85%
Maximum Variation (Peak to Peak)	2%	3%
Rotation Speed	< 300 RPM	< 60 RPM
Optical Connectors (Stator/Rotor)	FC/FC	FC/FC

 $^{^*\}text{Tested}$ with 200 μm core/0.22 NA optical fiber

Pigtailed Assisted Fiber-optic & Electric Rotary Joint

The *Pigtailed Assisted Fiber-optic & Electric Rotary Joint* is recommended for experiments that demand an extremely stable transmission. It is typically used in miniature fluorescence microscopy and fiber photometry.

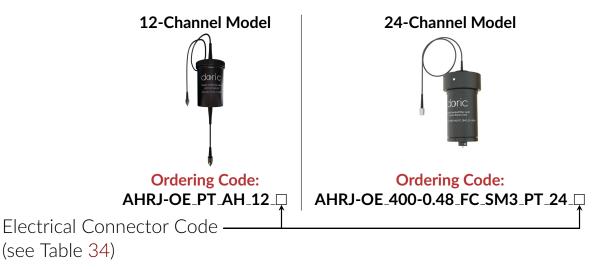
Notes:

- The holder for the *Pigtailed Assisted Fiber-optic & Electric Rotary Joint* is included (**Holder_ARJ**; see Table 117).
- A compatible FC/FC mating adapter (**ADAPTER_FC**; see Table 122) is sold separately and can be used to connect different patch cords to the pigtailed patch cord.

 Table 37: Pigtailed Assisted Fiber-optic & Electric Rotary Joint Specifications

SPECIFICATIONS	VAL	JE
Start-up Torque	< 20 µ	ιN∙m
Input/Output Fiber Core Diameter	200 or 4	00 µm
Input/Output Fiber NA	NA-0.37 or 0.57	
Electrical Contact Material	Gol	d
Number of Contacts	12	24
Contact Resistance	$<$ 500 m Ω	$< 800 \text{ m}\Omega$
Resistance Variation During Rotation	$< 100 \ { m m}\Omega$	$< 25 \text{ m}\Omega$
Transmission	45%*	70% [†]
Maximum Variation (peak to peak)	< 2%	< 1%
Rotation Speed	< 300 RPM	< 60 RPM
Optical Connectors (Rotor/Stator)	FC/FC	FC/M3

Table 38: 1-channel Assisted Rotary Joints



Assisted 1x2 Fiber-optic & Electric Rotary Joint

The Assisted 1x2 Fiber-optic & Electric Rotary Joint is an assisted rotary joint that splits a single fiber-optic channel into two. They are primarily used for bilateral optogenetic stimulation. Notes:

- The holder for the Assisted 1x2 Fiber-optic & Electric Rotary Joint is included (Holder_ARJ; see Table 117).
- The output fiber-optic patch cords are also included.



Assisted 1x2 Fiber-optic & Electric Rotary Joint

Table 39: Assisted 1x2 Fiber-optic & Electric Rotary Joint Specifications

SPECIFICATIONS	VALUE
Start-up Torque	< 20 µN⋅m
Input/Output NA	up to NA 0.22
Electrical Contact Material	Gold
Number of Contacts	24
Contact Resistance	$<$ 800 m Ω
Resistance Variation During Rotation	$<$ 25 m Ω
Transmission (per channel)	>40%
Maximum Variation (peak to peak)	3%
Rotation Speed	< 60 RPM
Optical Connectors (Rotor/Stator)	FC/2FC

ORDERING CODE: AHRJ-OE_ $1x2$
1x2 rotary joint type — i for intensity division, w for wavelength division.
Output wavelengths A/B For wavelength division only.
Input receptacle code — FC is standard.
Electrical connector code HDMI or USB-C. (see Table 34)

Rotary Joints

2x2 Assisted Fiber-optic & Electric Rotary Joint

The 2x2 Assisted Fiber-optic & Electric Rotary Joint allows frictionless rotation of 2 independent optical channels and 24 electrical contacts thanks to motorized assistance. The rotary joint detects torsion in optical fiber patch cords connected to a moving animal, which turns the rotor and allows the animal to move freely without discomfort. The two optical inputs use a standard FC connector, while the optical output uses a 1.0 mm guiding socket connector. Each optical channel is independent and features near zero crosstalk. The 24 electrical contacts are accessible using a standard USB-C connector (see Table 42). This rotary joint is optimal for use in optogenetics, allowing two regions of the brain to be activated independently.



2x2 Assisted Fiber-optic & Electric Rotary Joint

GND 5V

To assistance module

Note:

- The holder for the 2x2 Assisted Fiber-optic & Electric Rotary Joint is included (Holder_AHRJ-OE_2x2; see Table 117).
- The output patch cord with a 1.0 mm guiding socket connector is included.

Table 40: Assisted 2x2 Fiber-optic & Electric Rotary Joints SpecificationsTable 42: USB-C Pinout

SPECIFICATION	VALUE	Male (on rotary joint)		
Optical Channels	2 independent	Top USB-C	Bottom USB-C	
Maximum variation	<3 % of maximum	A1 • A2 •	• A1 • B11	
Optimal Wavelength	Achromatic	A3 •	• B10	
Start up torque	< 20 µN⋅m	A4 • A5 •	• A4 • A5	
Input Fiber Connector	2 FC	A6 • A7 •	• A6 • A7	
Output Fiber Connector	GS 1.0	A8 • • • • • • • • • • • • • • • • • • •	• A8	
Electrical Inputs	22 data contacts, 2 power contacts	A10 •	• B3 • B2	
Electrical Outputs	22 data contacts, 2 power contacts	A12 •	• A12	
Contact material	Gold	B1 • • • • • • • • • • • • • • • • • • •	• B1 • A11	
Contact resistance	$<$ 800 m Ω	B3 •	• A10	
Recommended Optical Fiber Patch Cord	200 μm/0.22	B5 •	• B5 • B6	
Transmission $(Typical)^*$	65%	B7 •	B7	
Table 41: 2x2 Assisted Fiber-optic & Ele	ectric Rotary Joints Ordering Code	B9 B10 B11 B12	• B9 • A3 • A2 • B12	

Connector Type	Ordering Code
USB-C	AHRJ-OE_2x2_AD_200-0.22_24_USB-C

^{*}Includes an output patch cord of the same type

Pigtailed 2x2 Assisted Fiber-optic & Electric Rotary Joint

This Pigtailed 2x2 Assisted Fiber-optic & Electric Rotary Joint is optimal for experiments requiring a highly stable output (e.g. fiber photometry), and allows multiple regions to be illuminated independently. These rotary joints have pigtailed input (1 m) fiber-optic patch cords of 200 or 400 μ m diameter (NA 0.37 or 0.57) with a light weight metal jacket.

<u>Note</u>: The holder for the *Pigtailed 2x2 Assisted Fiber-optic & Electric Rotary Joint* is included (**Holder_AHRJ-OE_2x2**; see Table 117).

♥ Pigtailed 2x2 Assisted Fiber-optic & Electric Rotary Joint

orio

SPECIFICATION	VALUE					
Optical Channels		2 indepe	endent			
Maximum variation		<1.5 % of	maximum			
Optimal Wavelength		530	nm			
Start up torque		< 20 µ	uN∙m			
Input Fiber Connector	2 FCM					
Output Fiber Connector	GS 1.0					
Electrical Inputs	22 data contacts, 2 power contacts					
Electrical Outputs	22	2 data contacts, 2	2 power contact	S		
Contact material		Go	ld			
Contact resistance	$< 800 \text{ m}\Omega$					
Fiber Core/NA	200 µm/0.37	200 µm/0.57	400 μm/0.37	400 µm/0.57		
Transmission (Typical)*	55%	50%	45%	35%		

Table 43: Pigtailed 2x2 Assisted Fiber-optic & Electric Rotary Joints Specifications

Table 44: Pigtailed 2x2 Assisted Fiber-optic & Electric Rotary Joints Ordering Codes

Fiber-optic Patch Cord	Ordering Code
200 µm core - NA 0.37	AHRJ-OE_2x2_PT_AH_200-0.37_24_USB-C
200 μm core - NA 0.57	AHRJ-OE_2x2_PT_AH_200-0.57_24_USB-C
400 μm core - NA 0.37	AHRJ-OE_2x2_PT_AH_400-0.37_24_USB-C
400 µm core - NA 0.57	AHRJ-OE_2x2_PT_AH_400-0.57_24_USB-C

⁵⁷

^{*}Includes an output patch cord of the same type

Fiber-optic & Liquid Rotary Joints

To get better insights of the brain functions, it is desirable to combine different methods for deep brain manipulation of neuronal activity. In order to allow for the delivery of light and fluid simultaneously in freely-moving animals, the rotary joint needs to combine functions of the fiber-optic and liquid rotary joints within one instrument.

Fiber-optic & Liquid Rotary Joint

Our Fiber-optic & Liquid Rotary Joint consists of an optical arrangement allowing the passage of fluid into a small tubing that minimize the perturbation of the light transmission during the rotation. Stainless steel fluid swivels from Instech Solomon are required for the use of this rotary joint. The 1-channel fluid swivel comes with the *Fiber-optic & Liquid Rotary Joint* and if more channels are needed, it can be adapted to work with the 2- or 5channel fluid swivel. Two versions of the product are available depending on the liquid tubing size (22 or 25 gauge).



Notes:

- The output fiber-optic patch cords are also included.
- The joint comes with a pre-installed metal tube for the insertion of plastic tubing and a box of 50 supplemental metal tubes. Eight different positions are possible on the rotary joint for the metal tubes.
- The package includes 1 m of plastic tubing.
- The 1-channel fluid swivel and its attachments are included.
- To prevent cross-contamination, we recommend to replace plastic and metal tubes with clean ones when

Fiber-optic & Liquid Rotary Joint - 1 channel liquid swivel changing liquid solutions. Spare metal tubes can be ordered in lots of 25 units.

Table 45: General Specifications

SPECIFICATIONS	VALUE	NOTES
Transmission	60-65%	With 200 μm core, 0.22 NA optical fiber
Maximum Variation	$\pm 5\%$	Additional +10% power drop when tubing crosses light path
Start-up Torque	$pprox$ 150 μ N·m	Without Fluid Swivel
	≈ 600 µN⋅m	With 1-channel Fluid Swivel *
Input/Output NA	0.22	-

Table 46: Fiber-optic & Liquid Rotary Joint Ordering Codes

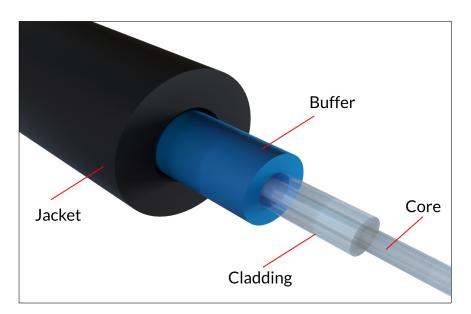
	Ordering Code					
	Tubing					
Gauge	Rotary joint	Metal	Flexible			
22 25	HRJ-OL_FC-FC_22 HRJ-OL_FC-FC_25					

^{*}See supplier specifications on the INSTECH website

Patch Cords

Fiber-optic Patch Cords

In the context of optogenetics experiments with the rotary joint, a *Fiber-optic Patch Cord* is needed to connect the light source and the rotary joint and yet another patch cord to connect the rotary joint and the fiber-optic cannula.



Structure of a Fiber-optic Patch Cord

The *core* and the *cladding* are two layers that make up the lightguide. However, the light travels inside the core of the fiber-optic, barely or not inside the cladding. For this reason, interconnected fiber-optics should have the same core diameter. Different cladding diameters have no influence on the coupling efficiency.

The *buffer* is a protective layer that tightly encircles the cladding. For patch cords, we usually recommend the use of another protective layer, called *jacket*, which is a loose tube covering the previously mentioned layers of the cable.

Mono Fiber-optic Patch Cords

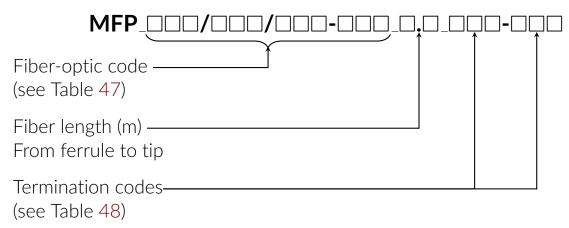
Standard Mono Fiber-optic Patch Cords



Standard Mono Fiber-optic Patch Cord

The simplest form of the patch cord is a piece of fiber with buffer coating and two ferrules on its ends. So far, the most popular fiber in optogenetics research is a fiber with a 200 μ m core diameter and NA 0.22.

ORDERING CODE:



Note:

• The fiber diameter and its numerical aperture (collection angle) limit the coupling efficiency into the fiber. Therefore, for higher coupling from sources like LEDs into an optical fiber of a specific diameter, please select a higher NA fiber and follow it all the way through to the fiber-optic cannula.

			Outer diameter (µm)			
	Core (µm)	Cladding (µm)	Buffer	Jacket [*]	NA	Fiber-optic Code
	50	125	250	900	0.22	50/125/900-0.22
	60	65	75	900	0.37	60/65/900-0.37
	62.5	125	250	900	0.27	62.5/125/900-0.27
	100	110	125	900	0.22	100/110/900-0.22
	100	110	125	900	0.37	100/110/900-0.37
	105	125	250	900	0.22	105/125/900-0.22
	200	220	240	900	0.22	200/220/900-0.22
	200	220	245	900	0.37	200/220/900-0.37
	200	230	500	900	0.48	200/230/900-0.48
g	200	240	400	900	0.22	200/240/900-0.22
Silica	300	330	360	900	0.37	300/330/900-0.37
S	300	330	370	900	0.22	300/330/900-0.22
	300	330	650	1000	0.48	300/330/1000-0.48
	300	335	650	1000	0.37	300/335/1000-0.37
	400	430	730	1100	0.37	400/430/1100-0.37
	400	430	730	1100	0.48	400/430/1100-0.48
	400	440	480	900	0.22	400/440/900-0.22
	550	600	1040	PVC 3 mm	0.22	550/600/3000-0.22
	600	630	1040	PVC 3 mm	0.37	600/630/3000-0.37
	600	630	1040	PVC 3 mm	0.48	600/630/3000-0.48
	600	660	710	1100	0.22	600/660/1100-0.22
	240	250	-	PVC 1 mm	0.63	240/250/900-0.63
stic	480	500	-	PVC 1 mm	0.63	480/500/900-0.63
Plastic	960	1000	-	PVC 2.2 mm	0.63	960/1000/2200-0.63
	1480	1500	-	PVC 3 mm	0.50	1480/1500/3000-0.50

Table 47: Standard Mono Fiber-optic Patch Cords Codes

^{*}Standard jacket; other jackets are also available, see Protective Jackets.

Description	Product	Termination Code
FC Connector with Zirconia Ferrule		FC
FC Connector with Metal Ferrule		FCM
FC/APC Connector with Zirconia Ferrule*		FCA
FC/APC Connector with Metal Ferrule [*]		FCMA
SMA Connector with Metal Ferrule		SMA
Zirconia Ferrule OD = 1.25 mm		ZF1.25
Zirconia Ferrule OD = 1.25 mm with Flange		ZF1.25(F)
Zirconia Ferrule OD = 1.25 mm with Peek Flange		ZF1.25(FP)
Metal Ferrule OD = 1.25 mm		MF1.25
Zirconia Ferrule OD = 2.5 mm		ZF2.5
Zirconia Ferrule OD = 2.5 mm with Flange		ZF2.5(F)
Zirconia Ferrule OD = 2.5 mm with Peek Flange		ZF2.5(FP)
Metal Ferrule OD = 2.5 mm		MF2.5
Slim Magnetic Connector		SMC
M3 Connector		CM3
M3 Connector Peek Plastic		CM3(P)
M2 Connector		CM2
M2 Connector Peek Plastic		CM2(P)

Table 48: Termination Codes for Fiber-optic Patch Cords

^{*}FC/APC Connectors available for Fiber-optic Patch Cords NA 0.22 only.

Attenuating Mono Fiber-optic Patch Cords

Optical fiber patch cords with an integrated attenuating filter are ideal for applications where optical power coupled into a fiber is too high, i.e. fiber photometry excitation. Addition of attenuating filter does not affect light distribu-



Attenuating Mono Fiber-optic Patch Cord

tion inside the optical fibre, only transmission is reduced. Different optical fibers or attenuating factors are possible.

ORDERING CODE:

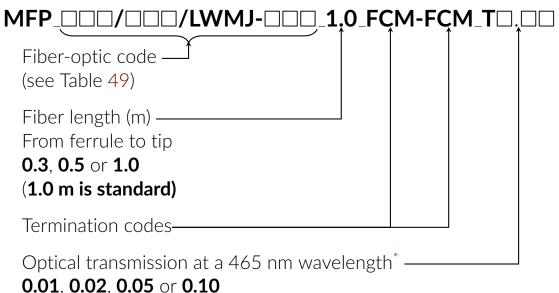


Table 49: Attenuating Mono Fiber-optic Patch Cords Codes

		Outer di	ameter (µm)		
Core (µm)	Cladding (µm)	Buffer	Jacket	NA	Fiber-optic Code
200	220	240	LWMJ	0.22	200/220/LWMJ-0.22
200	220	245	LWMJ	0.37	200/220/LWMJ-0.37
200	230	500	LWMJ	0.48	200/230/LWMJ-0.48
200	230	500	LWMJ	0.57	200/230/LWMJ-0.57
400	430	730	LWMJ	0.48	400/430/LWMJ-0.48
400	430	730	LWMJ	0.57	400/430/LWMJ-0.57
400	440	470	LWMJ	0.37	400/440/LWMJ-0.37
400	440	480	LWMJ	0.22	400/440/LWMJ-0.22

^{*}Optical transmission is specified for visible light, and measured at a 465 nm wavelength. Please note that for a 405 nm wavelength (UV), the transmission value is about half of the specification for visible light.

Low Autofluorescence Mono Fiber-optic Patch Cords

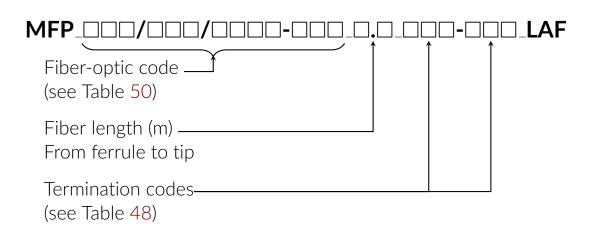
The Low Autofluorescence Mono Patch Cords are specially designed to minimize the fluorescence caused by photo-active compounds in the optical fiber and associated elements. Fibers with a low natural fluorescence are selected and used with glues, ferrules and protective coatings that have low or no-fluorescence. To ensure its minimal autofluorescence, each patch cord under-



Low Autofluorescence Mono Fiber-optic Patch Cord

goes a 12-hour photobleaching process that decreases their remaining natural fluorescence. Patchcords with NA = 0.37 use all glass fiber which have a very low autofluorescence level naturally. Consequently, it is not necessary to use bleaching process on LAF patchcords with NA=0.37.

ORDERING CODE:



Notes:

- The autofluorescence is dependent on the length of the patch cord and the fiber core diameter. In order to minimize autofluorescence, we recommend that the patch cord be as short as possible.
- The photobleaching process creates a temporary and reversible decrease of autofluorescence. It is recommended to repeat this process before each use to minimize autofluorescence. The instructions are provided with the product.
- The photobleaching process is much more efficient for fiber with hard polymer cladding (NA=0.48, 0.53, 0.57) than ones with all glass fiber (NA= 0.22, 0.37, 0.66). For this reason it is not necessary to use this process on LAF patch cords with NA=0.37.

Table 50: Low Autofluorescence	Fiber-optic Patch Cords Codes
--------------------------------	-------------------------------

		Outer d	iameter (µm)		
Core (µm)	Cladding (µm)	Buffer	Jacket*	NA	Fiber-optic Code
100	110	125	900	0.37	100/110/900-0.37
200	220	245	900	0.37	200/220/900-0.37
200	230	500	900	0.57	200/230/900-0.57
400	430	730	1100	0.57	400/430/1100-0.57
400	440	470	900	0.37	400/440/900-0.37

^{*}Standard jacket; other jackets are also available, see Protective Jackets.

Dual Fiber-optic Patch Cords

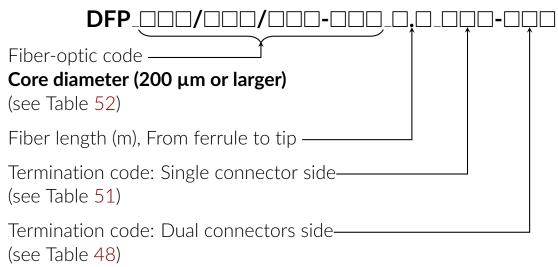
A *Dual Fiber-optic Patch Cord* has two optically isolated fibers. One side ends with a dual ferrule guiding pin or a guiding socket connector. The other side of the patch cord can also end with a dual ferrule connector or with separate FC connectors for each fiber. The dual fiber-optic patch cord can transmit in-



Dual Fiber-optic Patch Cord

dependent optical signals when used with a Separate Light Path 2x2 Fiber-optic Rotary Joint. Using dual patch cords with a 1x2 Fiber-optic Rotary Joint is optically more efficient but more expensive than using a branching patch cord with a 1x1 Fiber-optic Rotary Joint.

ORDERING CODE:



DF□.□
GS <u>□.□</u>

^{*}Recommended termination for Dual Fiber-optic Patch Cord

			Outer diameter (µm)			
	Core (µm)	Cladding (µm)	Buffer	Jacket [*]	NA	Fiber-optic Code
	200	220	240	900	0.22	200/220/900-0.22
	200	220	245	900	0.37	200/220/900-0.37
	200	230	500	900	0.48	200/230/900-0.48
	200	240	400	900	0.22	200/240/900-0.22
	300	330	360	900	0.37	300/330/1000-0.37
IJ	300	330	370	900	0.22	300/330/900-0.22
Silica	300	330	650	1000	0.48	300/330/1000-0.48
S	300	335	650	1000	0.37	300/335/1000-0.37
	400	430	730	1100	0.37	400/430/1100-0.37
	400	430	730	1100	0.48	400/430/1100-0.48
	400	440	480	900	0.22	400/440/900-0.22
	550	600	1040	PVC 3 mm	0.22	550/600/3000-0.22
	600	630	1040	PVC 3 mm	0.37	600/630/3000-0.37
	600	630	1040	PVC 3 mm	0.48	600/630/3000-0.48
	600	660	710	1100	0.22	600/660/1100-0.22
Plastic	240	250	-	PVC 1 mm	0.63	240/250/900-0.63
	480	500	-	PVC 1 mm	0.63	480/500/900-0.63

Table 52: Dual Fiber-optic Patch Cords Codes

^{*}Standard jacket; other jackets are also available, see Protective Jackets.

Low Autofluorescence Dual Fiber-optic Patch Cords

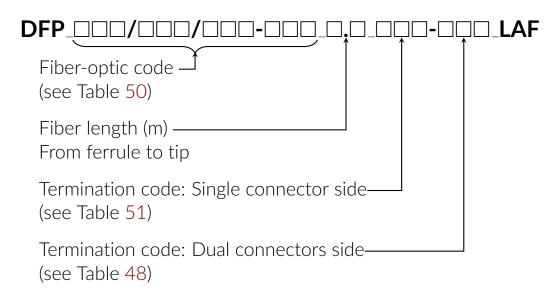
The Low Autofluorescence Dual Patch Cords are specially designed to minimize the fluorescence caused by photo-active compounds in the optical fiber and associated elements. Fibers with a low natural fluorescence are selected and used with glues, ferrules and protective coatings that have low or no-fluorescence. To ensure its minimal autofluorescence, each patch cord under-



Low Autofluorescence Dual Fiber-optic Patch Cord

goes a 12-hour photobleaching process that decreases their remaining natural fluorescence. Patchcords with NA = 0.37 use all glass fiber which have a very low autofluorescence level naturally. Consequently, it is not necessary to use bleaching process on LAF patch cords with NA=0.37.

ORDERING CODE:



Notes:

- The autofluorescence is dependent on the length of the patch cord and the fiber core diameter. In order to minimize autofluorescence, we recommend that the patch cord should be as short as possible.
- The photobleaching process creates a temporary and reversible decrease of autofluorescence. It is recommended to repeat this process before each use to minimize autofluorescence. The instructions are provided with the product.
- The photobleaching process is much more efficient for fiber with hard polymer cladding (NA=0.48, 0.53, 0.57) than ones with all glass fiber (NA= 0.22, 0.37, 0.66). For this reason it is not necessary to use this process on LAF patchcords with NA=0.37.

Branching Fiber-optic Patch Cords

The *Branching Fiber-optic Patch Cord* takes light entering a single fiber-optical connector and evenly splits said light into multiple different outputs. There are two types of branching patch cords-*Splitter* and *Bundle* branching patch cords.



Splitter Branching Fiber-optic Patch Cord

The *Splitter Branching Patch Cords* take light entering a single large-diameter optical fiber and split it evenly amongst two or more smaller-diameter optical fibers, each with their own connector. These patch cords are typically used in optogenetics or illumination applications.



Bundle Branching Fiber-optic Patch Cord, 2 Branches

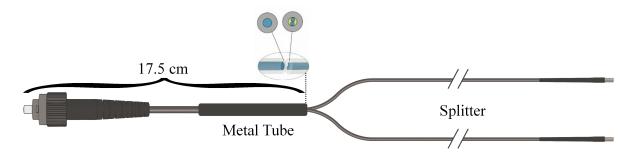
The Bundle Branching Fiber-Optic Patch Cords consist of two or more optical fibers bundled into a single optical connector with multiple branches on the opposite side. These patch cords are typically used in fiber photometry and similar lowautofluorescence applications.

Splitter Branching Fiber-optic Patch Cords

The Splitter Branching Patch Cords take light entering a single large-diameter and large numerical aperture optical fiber and split it evenly amongst several smallerdiameter optical fibers, each with their own connector. These patch cords are typically used in optogenetics, splitting a single powerful light source between several sites. Splitters are also effective at splitting and thereby increasing light uniformity when using a laser input. The splitter patch-cord also conserves light intensity (mW/mm^2) over all outputs. This means that each output will have the same intensity as a single optical fiber of the same type, allowing a single LED light source to be used for multiple sites with sufficient intensity.

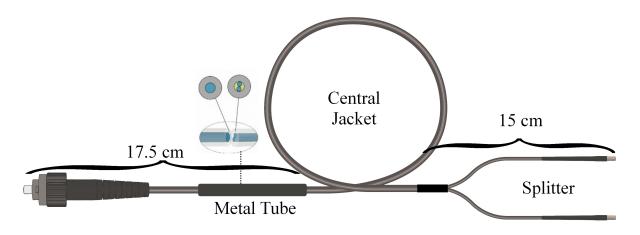
While the standard *Splitter Branching Fiber-optic Patch Cords* has 2 branches, more are available if requested. A two-branch patch cord splitter typically outputs between 16 and 20% of the power entering the input optical connector, with variations depending on fiber type and the area recovery ratio. This approach uses a 50 mm long metal tube with a 6.35 mm diameter to protect the passage from 1 to several fibers. The length of patch-cord from the metal tube to the patch-cord split, called the central jacket, can be protected using a different jacket type. Splitter patch cords are not suitable for short cables less than 30 cm long. This version allows improved splitting of laser light over older models. In all other applications, the *Splitter Branching Patch Cords* maintain the same high-quality performance as our legacy *Standard Branching Patch Cords*.

• The **Type-A** (AX identifier) patch cords have no central jacket, with the splitter starting at the metal tube. The split is done 17.5 cm from the input.



Type-A Splitter Branching Fiber-optic Patch Cord Layout, Without Central Jacket

• The **Type-B** (*BX* identifier) patch cords have a central jacket covering the length between the metal tube and the splitter. The split is done at 15 cm from the output.



Type-B Splitter Branching Fiber-optic Patch Cord Layout, With Central Jacket

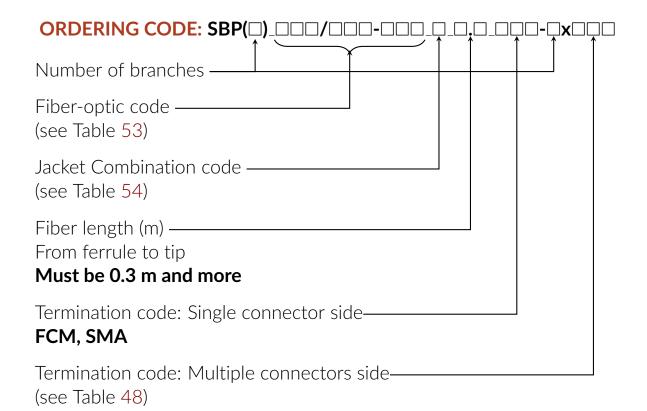
Note: The splitter makes use of multiple different jackets, as shown in the preceding figure. These result in two different patch cord types

	Out	er diamete	r (µm)		Standard Jacket	Fiber entie Code
	Core	Cladding	Buffer	NA	Combinations *	Fiber-optic Code
	50	125	250	0.22	A1 or B1	50/125-0.22
	60	65	75	0.37	A1 or B1	60/65-0.37
	62.5	125	250	0.27	A1 or B1	62.5/125-0.27
	100	110	125	0.22	A1 or B1	100/110-0.22
	100	110	125	0.37	A1 or B1	100/110-0.37
	105	125	250	0.22	A1 or B1	105/125-0.22
ŋ	200	220	240	0.22	A1 or B1	200/220-0.22
Silica	200	220	245	0.37	A1 or B1	200/220/900-0.37
S	200	230	500	0.57	A1 or B2	200/230-0.57
	400	430	730	0.48	A2 or B2	400/430-0.48
	400	440	470	0.37	A1 or B2	400/440-0.37
	400	430	730	0.57	A2 or B2	400/430-0.57
	400	440	480	0.22	A1 or B2	400/440-0.22
Plastic	240	250	-	0.63	A1 or B1	240/250-0.63
	480	500	-	0.63	A1 or B2	480/500-0.63

Table 53: Splitter Branching Fiber-optic Patch Cords Codes

Identifier	Central Jacket	Split Jacket	Jacket Combination Code
A1	None	900	900
A2	None	1100	1100
A3	None	2000	2000
A4	None	3000	3000
A5	None	LWMJ	LWMJ
A6	None	ARMO	ARMO
B1	900	900	900/900
B2	2000	1100	2000/1100
B3	LWMJ	LWMJ	LWMJ/LWMJ
Β4	LWMJ	900	LWMJ/900
B5	LWMJ	1100	LWMJ/1100
B6	ARMO	ARMO	ARMO/ARMO

Table 54: Jacket Combinations



<u>Note</u>: If Dual Fiber-optic Patch Cord connectors (**DF**... or **GS**...; Table 51) are required, remove the 2x before the termination code. Example: Splitter Branching Fiber-optic Patch Cord with dual connector and no central jacket: **BFP(2)**_-**200/230-0.57_900_1.5_FCM-GS1.5**

^{*}Standard jacket combination; other jacket combinations (AX and BX) can be found in Table 54

Bundle Branching Fiber-optic Patch Cords

The Bundle Branching Patch Cords use a bundle of multiple optical fibers rather than a single one. This bundle is together as the entry connector, and is split to multiple different connectors just after the input. This splits the light evenly amongst the different output connectors. There is no limitation in the choice of optical fiber, connector type or the number of branches. They are also assembled with transparent glue to prevent heat absorption in case of high intensity input light. This approach has many possible uses and many different configurations are available. These patch cords are typically used in fiber photometry, and are used to illuminate and recover light from multiple sites. Such bundles can also be used with an imaging camera to measure the input of each fiber at the same time. Other optical fiber and connector types can be made available on request. Different optical fiber types and more branches are possible on request.



Bundle Branching Fiber-optic Patch Cord , 2 Branches

Number of optical fibers ≤7 Optical Fibers	
Fiber-optic code (see Table 55)	
Fiber length (m) From ferrule to tip	
Termination code: Single connector side FCM, SMA	
Number of output connectors	
Termination code: Multiple connectors side	

Example:

Branching Fiber-optic Patch Cord with three fibers: **BBP(3)_200/230/900-0.57_-1.5_FCM-3xZF1.25**

			Outer diameter (µm)			
	Core (µm)	Cladding (µm)	Buffer	Jacket [*]	NA	Fiber-optic Code
	50	125	250	900	0.22	50/125/900-0.22
	60	65	75	900	0.37	60/65/900-0.37
	62.5	125	250	900	0.27	62.5/125/900-0.27
	100	110	125	900	0.22	100/110/900-0.22
	100	110	125	900	0.37	100/110/900-0.37
	105	125	250	900	0.22	105/125/900-0.22
	200	220	240	900	0.22	200/220/900-0.22
	200	220	245	900	0.37	200/220/900-0.37
	200	230	500	900	0.57	200/230/900-0.57
	200	230	500	900	0.48	200/230/900-0.48
	200	240	400	900	0.22	200/240/900-0.22
	300	330	360	900	0.37	300/330/1000-0.37
Silica	300	330	370	900	0.22	300/330/900-0.22
Sil	300	330	650	1000	0.48	300/330/1000-0.48
	300	335	650	1000	0.37	300/335/1000-0.37
	400	430	730	1100	0.48	400/430/1100-0.48
	400	440	470	900	0.37	400/440/900-0.37
	400	430	730	1100	0.57	400/430/1100-0.57
	400	440	480	900	0.22	400/440/900-0.22
	550	600	1040	PVC 3 mm	0.22	550/600/3000-0.22
	600	630	1040	PVC 3 mm	0.37	600/630/3000-0.37
	600	630	1040	PVC 3 mm	0.48	600/630/3000-0.48
	600	660	710	1100	0.22	600/660/1100-0.22
	240	250	-	PVC 1 mm	0.63	240/250/900-0.63
stic	480	500	-	PVC 1 mm	0.63	480/500/900-0.63
Plastic	960	1000	-	PVC 2.2 mm	0.63	960/1000/2200-0.63
	1480	1500	-	PVC 3 mm	0.50	1480/1500/3000-0.50

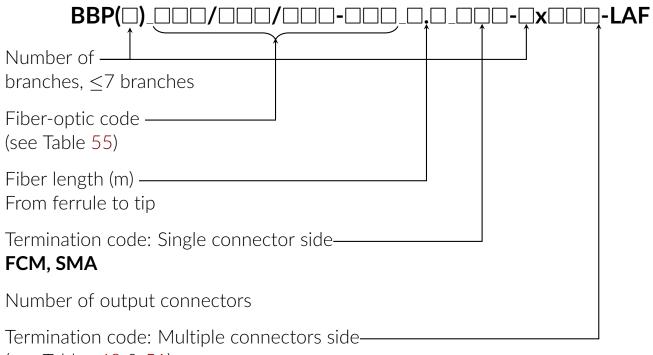
Table 55: Bundle Branching Fiber-optic Patch Cords

^{*}Standard jacket; other jackets are also available, see Protective Jackets.

Low Autofluorescence Bundle Branching Fiber-optic Patch Cords

The Low Autofluorescence Bundle Branching Fiber-Optic Patch Cords consist of two or more low autofluorescence optical fibers bundled into a single optical connector with multiple branches on the opposite side. There is no limitation in the connector type or the number of branches. The patch cord is assembled using low autofluorescence materials, such as high-absorption glue that minimizes cross-talk as well. To ensure minimal autofluorescence, and each patch cord undergoes a 12-hour photobleaching process that temporarily decreases the remaining natural fluorescence. This patch cord model is primarily used in multi-site fiber photometry, though it is useful in any application where low patch cord autofluorescence is required. Different optical fiber types and more branches are possible on request.

ORDERING CODE:



(see Tables 48 & 51)

Table 56: Low Autofluorescence Fiber-optic Patch Cords Codes

		Outer diameter (µm)			
Core (µm)	Cladding (µm)	Buffer	Jacket [*]	NA	Fiber-optic Code
200	230	500	900	0.57	200/230/900-0.57
400	430	730	1100	0.57	400/430/1100-0.57

*Standard jacket; other jackets are also available, see Protective Jackets.

Protective Jackets

For a better fiber protection, we also offer larger jackets made of PVC tubing. Metal jackets or jackets made of other materials are also available on request. If you want other jacket than those in Table 47, 52, 53 and 55, just replace corresponding jacket code with:

2000 for PVC jacket OD 2 mm

3000 for PVC jacket OD 3 mm

LWMJ for Lightweight metal jacket (black, OD 2.4 mm, 8 g/m)

ARMO for Armored jacket (OD 3 mm, 12 g/m)

Description	Outer Diamter (mm)	Mass (g/m)	Product	Jacket Code
900 µm	0.9	0.5		-
PVC Jacket-OD2	2.0	2.5		2000
PVC Jacket-OD3	3.0	5.8		3000
Lightweight Metal Jacket Armored Jacket	2.4 3.0	8.0 12.0		LWMJ ARMO

Table 57: Codes for Fiber-optic Patch Cord Jackets

Adapters

Low Profile Patch Cord Adapter

This adapter can be used to modify the direction path of a patch cord without bending the fiber cable. It can help to minimize stress constraints and allow to do a 90° bend within 6 mm radius.



Low Profile Patch Cord Adapter ZF1.25 to ZF1.25 - 90 °

SPECIFICATION	VALUE
Receptacle Size (W x L x H)	3.2 mm x 8.0 mm x 4.5 mm
Connection	ZF1.25, ZF2.5, MF1.25, MF2.5 ferrules
Fiber-optic Type	200/250-0.66; 400/430-0.66
Angle	Standard angles: 90°; Tolerance of +/- 0.5°
Material	Peek plastic/zirconia ferrule
Light transmission output	> 60%

Fiber-optic code	
Termination code: Patch cord to adapter side ZF1.25, ZF2.5, MF1.25, MF2.5	
Termination code: Adapter to cannula side ————————————————————————————————————	

Table 58: Low Profile Patch Cord Adaptor Specifications

Electrical Patch Cords

Electrical Patch Cords

The electrical patch cord or cable is offered to assure an interconnection with the electrical part of some of our opto-electric cannula. It could be used to interconnect a Mono Opto-electric Cannula to a recording headstage or to an electrical stimulator.



Mono Electric Patch cord

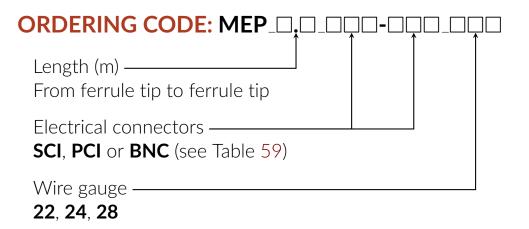


Table 59: (Connectors	for E	ectrical	Patch	Cords
-------------	------------	-------	----------	-------	-------

Description	Product	Termination Code
Socket Cooper Interconnect		SCI
Pin Cooper Interconnect		PCI
BNC		BNC

Opto-electric Patch Cords

Mono Opto-electric Patch Cords

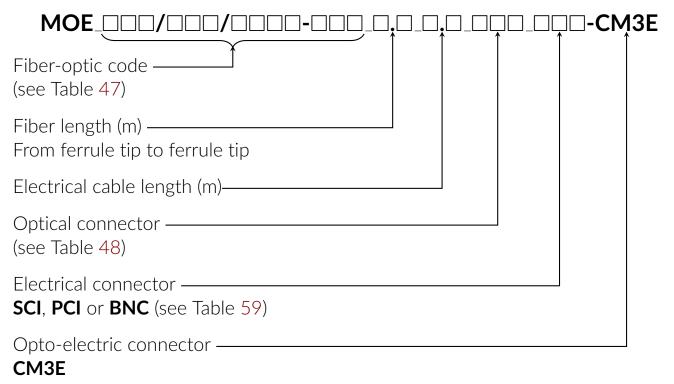
We offer an opto-electric patch cord that connects both modality on cannula with a single easy step. This M3E connection was inspired by our very popular M3 cannula connector, where we added an electrical pin within the connector. This patch cord is light and compact on the animal head and can be connected independently



Mono Opto-electric Patch Cord

on most common electrophysiological headstage and light sources. In order to minimize the electrical noise, we recommend short length between the cannula and the pre-amplification system, consequently the electrical wire length should be shorter as possible.

ORDERING CODE:



Cannulas

Fiber-optic Cannulas

A fluid cannula is an assembly of a metal tube and a fluid tube receptacle, used for administering fluids when the metal tube is inserted into the body. As an example, a venous cannula is inserted into a vein to obtain blood samples or to deliver medicines. The body of a cannula has a form that easily connects to or disconnects from the fluid tubing. The tubing is often disconnected while the cannula remains attached to the body surface with the hollow needle (tube) inserted into the body for the later use. Similarly, biomedical and optogenetics applications need *Fiber-optic Cannulas* to deliver the light into the body tissue and/or to collect fluorescence or scattered light coming from the tissue.

The illumination of neurons within the mouse's brain with the blue or amber light has become an essential tool for studying the processes within genetically modified photosensitive neurons. In the early days of optogenetics, a fluid cannula was used to insert the optical fibers into the brain tissue, where the metal tube was guiding the fiber to the point of interest. Occasionally, the optical fiber was removed from the fluid cannula only to be reinserted later. The optical fiber removal and re-entry often led to infections and clogging of the fluid cannula.

With some exceptions, the *Fiber-optic Cannula* is used without the metal tube of the fluid cannula. It consists of a fiber-optic ferrule with some sort of a fiber-optic receptacle on one side and the im-



Mono Fiber-optic Cannula - M3 slim

plantable fiber protruding from the other side. When the fiber-optic cannula is secured to the body and the fiber implanted, the light can be delivered to the tissue and the fluorescence or scatter from the tissue can be captured. In these experiments, it is imperative that the connection between the delivery fiber and the cannula is light, small and simple to connect and disconnect. For a mono fiber delivery, the connection between the ferrules of the light delivery fiber patch cord and the fiber-optic cannula is achieved, in its simplest form, via the fiber-optic sleeve.

The connector type connection is preferred but it is not always applicable. In some optogenetics experiments it is necessary to introduce two or more implantable fibers within a small and precise distance. Those applications call for the dual or multiple fiber-optic cannula easily connectable to the matching delivery fibers. The concept of *Fiber-optic Cannulas* with different optical fibers, receptacle types and fiber terminations is bound to be further fragmented. So far we carry Mono Fiber-optic Cannulas, Dual Fiber-optic Cannulas, and Two-ferrule Cannulas. In effect, we are developing hybrid cannulas that transmit a combination of light, liquid and electrical signals. Tables 61 and 62 show different possibilities for each cannula type.

Cannulas

Mono Fiber-optic Cannulas

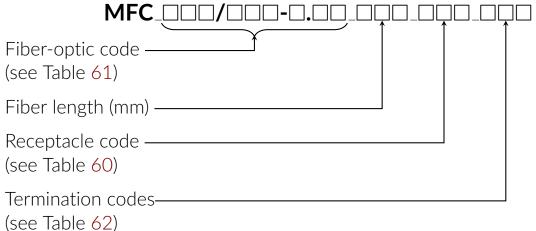
The Mono Fiber-optic Cannula is an assembly of a bare optical fiber, a fiber ferrule and a receptacle or a sleeve. One side of the ferrule is polished while the implantable part of the fiber protrudes from the opposite end of the ferrule. The ferrule is placed within the receptacle or sleeve to allow connection to the fiber-optic patch cord. The protruding fiber can be implanted into the body while the ferrule or the receptacle is attached to the skin. When the cannula is connected with the patch cord, it is possible to send the light signals to and from the tissue close to the fiber tip. It is imperative for *in vivo* optogenetics applications that the fiber-optic cannula allows for an efficient, plug and play type connection with the fiber-optic patch cord. A receptacle is a mechanical holder that defines the positions of the fiber tip and guides the connecting ferrule to the optical coupling position.



Mono Fiber-optic Cannula ZF2.5 with grooves

For *Mono Fiber-optic Cannulas* we offer zirconia sleeves as the simplest form of a receptacle, M2, M3 and rectangular magnetic receptacles.





Notes:

- The tolerance on the length of the protruding fiber is better than 0.1 mm.
- Sleeves required to connection with a patch cord are ordered separately (**SLEEVE**; see Table 122).
- Adapters (Receptacle adapters) are available for M2 and M3 receptacles (**FCA**; see Table 76).
- A Stereotaxic Cannula Holder (Stereotaxic Cannula Holders) is available for implantation to secure the Mono Fiber-optic Cannulas (**SCH**; see Table 73).

Description	Product	Receptacle Code
Zirconia ferrule OD 1.25 mm		ZF1.25
Zirconia ferrule OD 1.25 mm with grooves		ZF1.25(G)
Metal ferrule OD 1.25 mm		MF1.25
Zirconia ferrule OD 2.5 mm		ZF2.5
Zirconia ferrule OD 2.5 mm with grooves		ZF2.5(G)
Metal ferrule OD 2.5 mm		MF2.5
Receptacle with M2 thread Titanium		RM2
Receptacle with M2 thread Peek plastic		RM2(P)
Receptacle with M3 thread Titanium		SM3
Receptacle with M3 thread Peek plastic		SM3(P)
Slim Magnetic Receptacle Aluminum		SMR

Table 60: Receptacle Codes for Mono Fiber-optic Cannulas

	Core Diameter (µm)	Outer Diameter (μm)	NA	Buffer Color	Outer Layer	Fiber-optic Code
	44*†‡	50	0.66	clear	Borosilicate (fragile)	044/050-0.66
	50 [*]	70	0.22	yellow	Polyimide buffer	050/070-0.22
	60*	75	0.37	yellow	Polyimide buffer	060/075-0.37
	100	125	0.22	yellow	Polyimide buffer	100/125-0.22
	100	125	0.37	yellow	Polyimide buffer	100/125-0.37
	100	125	0.66	clear	Borosilicate (fragile)	100/125-0.66
	200	230	0.48	clear	Hard polymer cladding	200/230-0.48
	200	240	0.22	yellow	Polyimide buffer	200/240-0.22
	200	245	0.37	yellow	Polyimide buffer	200/245-0.37
	200	250	0.66	clear	Borosilicate	200/250-0.66
	200	260	0.22	clear	Silicone buffer	200/260-0.22
Silica	300	330	0.37	clear	Hard polymer cladding	300/330-0.37
Sil	300	360	0.37	yellow	Polyimide buffer	300/360-0.37
	300	370	0.22	yellow	Polyimide buffer	300/370-0.22
	400	430	0.37	clear	Hard polymer cladding	400/430-0.37
	400 [†]	430	0.48	clear	Hard polymer cladding	400/430-0.48
	400	430	0.66	clear	Borosilicate	400/430-0.66
	400	470	0.37	yellow	Polyimide buffer	400/470-0.37
	400	480	0.22	yellow	Polyimide buffer	400/480-0.22
	600	630	0.48	clear	Hard polymer cladding	600/630-0.48
	600	710	0.37	yellow	Polyimide buffer	600/710-0.37
	200	230	0.57	clear	Hard polymer cladding	200/230-0.57
	400	430	0.57	clear	Hard polymer cladding	400/430-0.57
<u>.</u>						
Plastic	240*,†,‡	250	0.63	clear	PMMA	240/250-0.63
Ē	480 ^{*,†§}	500	0.63	clear	PMMA	480/500-0.63

Table 61: Fiber-optic Codes for Cannulas

- [†]Not offered with a 45° mirror tip (**MA45**; see Table 62).
- [‡]Not offered with a diffuse layer tip (**DFL**; see Table 62).
- [§]Only for fiber lengths inferior to 5 mm

^{*}Not offered with a conical tip (**C45**, **C60**; see Table 62).

Cannulas

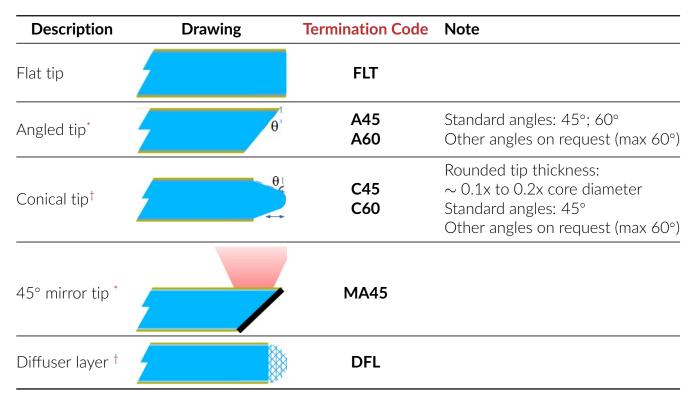


 Table 62: Fiber-optic Termination Codes for Cannulas

^{*} Angled and 45° mirror tips are not offered with Opto-fluid Cannulas (**OsFC**, **OmFC**, **iOFC**, **DiOFC**), Opto-electric Cannulas (**OEC**), Opto-electric Probe Tips (**OEPT**), Low Profile Cannulas (**MFC_LPB**) and Two-ferrule Low Profile Cannulas. See other exceptions on Table 61.

[†] Some Optical Fibers are not offered with this type of termination. See Table 61.

Low Profile Cannulas

Low Profile Cannulas are designed to minimize the height over the animal's head. The patch cord connection is in the same axis as the animal's body instead of hanging over its head. This cannula allows for the fiber optic implantation in a standard stereotaxic dorsoventral axis, but the interconnection is along the anteroposterior plane. This cannula design minimizes the pressure applied on the animal's head during the connec-

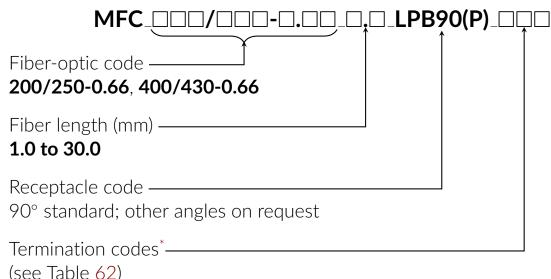




tion/disconnection of the patch cord. Furthermore, the *Low Profile Cannula* facilitates the motion of the animal's head in restraint areas. A shortened version is also available.

SPECIFICATION	VALUE
Receptacle Size (W x L x H)	2.4 mm x 6.0 mm x 2.0 mm
Connection	ZF1.25 ferrule (height 1.02 mm) and ZF1.25 patch cord
Fiber Type	200/250-0.66; 400/430-0.66
Angle	Angle Standard = 90 deg. Custom on request. Tolerance +/- 2 deg
Protrusion Length	1.0 to 30.0 mm. Tolerance +/- 0.2 mm
Material	Peek plastic/zirconia ferrule, Aluminium on custom demand
Light transmission output	> 60%

ORDERING CODE:



<u>Note</u>: A Low Profile Cannulas Holder (Low Profile Cannula Holder) is available for implantation to secure the Low Profile Cannula (**SCH_LP90**).

^{*}Angled (A45, A60) and mirror (MA45) tips are not offered with Low Profile Cannulas (see Table 62).

Dual Fiber-optic Cannulas

A *Dual Fiber-optic Cannula* features two implantable fibers at a prescribed distance and protrusion length held by a single ferrule. The tolerance on the protrusion for each fiber is better than 0.1 mm. These cannulas are perfectly suited for a bilateral brain stimulation or silencing. The alternatives to a dual fiber-optic cannula are two mono fiber cannulas. However, positioning two cannulas with stereotaxic equipment, one at a time, has a greater likelihood of positioning errors, prolongs the duration of the operation, complicates the fixation of the cannulas and increases the minimum possible distance between the two fiber tips. With *Dual Fiber-optic Cannula* the insertion of the fiber is faster (single shot), while the distance between the fiber tips and the protrusion depth are factory set. The precision fiber-to-fiber mating of the cannula with the corresponding fiber-optic patch cord is vital for good coupling and this is achieved by a guiding pin or by a guiding socket.



Patch Cord / Cannula connection with a guiding pin



Patch Cord / Cannula connection with a guiding socket

Dual Fiber-optic Cannula with a guiding pin

This cannula with a circular metal ferrule and two optical fibers has a guiding hole to insure precise alignment when connecting to the equivalent dual fiber-optic connector loaded with a guiding pin. The smallest optical fiber core diameter it can accept is 200 μ m while the fiber-to-fiber distance is in 0.7 to 1.7 mm range. For larger inter-fiber distances, please refer to dual fiber-optic cannula with a guiding socket or to Two-ferrule Cannulas. A typical transmission of the cannula with a guiding pin is higher than 70% for either fiber channel.



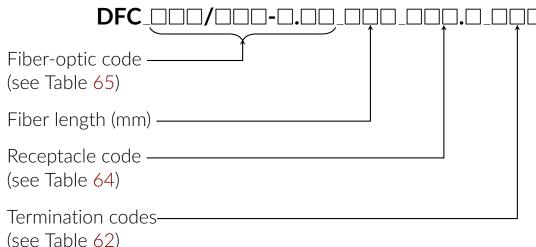
Dual Fiber-optic Cannula with a guiding pin

Dual Fiber-optic Cannula with a guiding socket

In spite of the huge popularity of our dual fiber-optic cannulas with a guiding pin, the need for a stronger and more user friendly connection led to development of a new *Dual Fiber-optic Cannula with a guiding socket*. In a way, it resembles M3 cannulas with its screw-in connecting feature. The guiding socket assures the orientation of the fiber tips of the corresponding connector. The fiber ferrule is an assembly of precision ground zirconia ferrules

that can have fiber holes as small as $125 \,\mu$ m diameter. These cannulas have thread diameter of 3.2 mm and provide the possibility of larger pitch distances. The *Dual Fiber-optic Cannula with a guiding socket* has a typical transmission higher than 75% for each fiber channel. It is important to note that the maximum pitch of 1.7 mm relates to standard 3.2 mm outside thread. If larger diameter thread or cannula studs are permitted, then the pitch between two fibers can be greater.

ORDERING CODE:



Notes:

- The *Dual Fiber-optic Cannula* with guiding pin is implanted using the Stereotaxic Cannula Holders 2.5, while the guiding socket model requires a specific adapter (available for 1.25 or 2.5 Stereotaxic Cannula Holders).
- Angled and conical tips (Axx and Cxx) are offered to facilitate the insertion of the fiber-optic in the tissue (see Table 62). However, they have little influence on the light spread.



Dual Fiber-optic Cannula with a guiding socket

^{*}Recommended receptacle for Dual Fiber-optic Cannulas

Table 64: Receptacle Codes for Dual Fiber-optic Cannulas

Description	Product	Receptacle Code
Dual ferrule with a guiding pin Pitch from 0.7 mm to 1.7 mm		DF□.□
Guiding socket [*] Pitch from 0.7 mm to 1.7 mm		GS□.□

	Core Diameter (µm)	Outer Diameter (µm)	NA	Buffer Color	Outer Layer	Fiber-optic Code
	200	230	0.48	clear	Hard polymer cladding	200/230-0.48
	200	240	0.22	yellow	Polyimide buffer	200/240-0.22
	200	245	0.37	yellow	Polyimide buffer	200/245-0.37
	200	250	0.66	clear	Borosilicate	200/250-0.66
ŋ	200	260	0.22	clear	Silicone buffer	200/260-0.22
Silica	300	330	0.37	clear	Hard polymer cladding	300/330-0.37
S	300	360	0.37	yellow	Polyimide buffer	300/360-0.37
	300	370	0.22	yellow	Polyimide buffer	300/370-0.22
	400	430	0.37	clear	Hard polymer cladding	400/430-0.37
	400†	430	0.48	clear	Hard polymer cladding	400/430-0.48
	400	430	0.66	clear	Borosilicate	400/430-0.66
	400	470	0.37	yellow	Polyimide buffer	400/470-0.37
	400	480	0.22	yellow	Polyimide buffer	400/480-0.22
	200	230	0.57	clear	Hard polymer cladding	200/230-0.57
	400	430	0.57	clear	Hard polymer cladding	400/430-0.57

Two-ferrule Cannulas

The *Two-ferrule Cannula* provides two implantable fibers, each within its own ferrule, at a precise distance exceeding 1.7 mm. The tolerance on the protrusion for each fiber is less than 0.1 mm. These cannulas are perfectly suited for the applications where two brain centers at a distance larger than 1.7 mm from each other are optically stimulated or controlled. The positioning of one mono fiber cannula at a time with the stereotaxic equipment has a greater likelihood of 3D positioning errors (lateral and depth). With a two-ferrule cannula the insertion of the

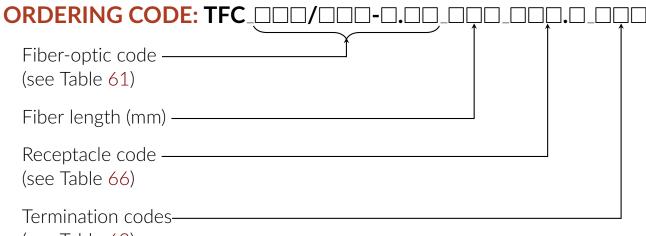


Two-ferrule Cannula

fiber is faster (single shot), the distance between the fiber tips is predefined and the protrusion depth is assured.

Two types of receptacles are currently available for the two-ferrule cannula (see pictures on next page). They both consist of precision machined holders that house zirconia ferrules and determine the spacing between the ferrule centers. The first type of two-ferrule cannula connects to a pair of patch cords terminated with 1.25 mm ferrules by using two zirconia sleeves (ID 1.25 mm). In the other case, the holder also includes a pair of magnets, so that the cannula can connect to a pair of rectangular magnetic connectors.

The two-ferrule cannula can be made for distances larger than 1.7 mm. For shorter distances between the brain centers, please refer to Dual Fiber-optic Cannulas.



(see Table <mark>62</mark>)

Notes:

• Unless otherwise specified, an aluminum housing and 1.25 mm zirconia ferrules are being used.

• The *Two-ferrule Cannula* is implanted using the Stereotaxic Cannula Holders 1.25 mm. For pitch less than 2 mm, ask for a customized stereotaxic cannula holder.

Center To Center Distance Between Ferrules (mm)	Product	Receptacle Code
Sleeve Co	onnection	
(x)		TFx *
2.0 mm		TS2
2.5 mm		TS2.5
3.0 mm		TS3
3.5 mm	\mathbf{x}	TS3.5
4.0 mm		TS4
Other (x)		TSx
Magnetic (Connection	
3.0 mm 4.0 mm Other (x)		TSM3 TSM4 TSMx

Table 66: Receptacle Codes for Two-ferrule Cannulas

Two-ferrule Low Profile Cannulas

On custom basis, we can do *Two-ferrule Low Profile Cannulas* with the appropriate fiber pitch distance, up to 10 mm.[†]

Note: A Low Profile Cannulas Holder (Low Profile Cannula Holder) is available for implantation to secure the Two-ferrule Low Profile Cannula (**SCH_LP90**).



Two-ferrule Low Profile Cannula - 90°

ORDERING:

Contact our sales department (sales@doriclenses.com)

^{*}Will be sold while stock lasts.

[†]Angled (**A45**, **A60**) and mirror (**MA45**) tips are not offered with Two-ferrule Low Profile Cannulas (see Table 62).

Fiber-optic Array Cannulas

Optogenetics experiments that target multiple excitation sites require cannulas with multiple fibers arranged within an fiber array. A one dimensional fiber array has several parallel fibers within the same plane at various distances and protrusions. Two dimensional fiber arrays are also possible (m x n). Currently, only the cannulas with well-arranged fiber-optic arrays on the tissue side and a bundled fibers on the side connecting to fiber patch cord are being offered. In this way similar levels of illumination are obtained around each fiber tip. The *Fiber-optic Array Cannula* where each fiber within the cannula is connected to corresponding fiber in the patch cord array is a possibility.

Array of m x n Fibers

For experiments that require specific spatial brain targets, we are able to offer customized fiber arrays to reach those places. It is possible to determine center-to-center distance between the fibers within a few microns and protrusion length of those fibers with a precision of 100 μ m. Such components are available with all standard fiber diameters and numerical aperture of 0.22 or 0.37.

Description	Product
Eight fibers in-line as a patch cord array termination	
One dimensional three fiber array with different pitch and length	
12 fibers divided into 2 groups of 2 x 3 fibers	

Examples of Fiber-optic Array Cannulas

ORDERING:

Contact our sales department (sales@doriclenses.com)

Fiber-optic Cannulas with LED

Fiber-optic Cannula + Single LED

We have developed an assembly where the LED is the integral part of the fiber-optic cannula thus providing a lightweight optical source attached to the head of the animal suitable for a deep brain illumination. The protruding optical fiber is implanted into the skull. In order to keep the assembly small and light there is no heat sink. To avoid heating, the maximum current should be limited.



Fiber-optic Cannula + Single LED

SPECIFICATION	VALUE	NOTE
Maximum current	150 mA 300 mA	Continuous (CW) 10 msec pulse, 10% duty cycle
Dimensions Mass Interface Pinout	6 x 6 x 6 mm 200 mg 3 pins header, 1.27 mm Pin 1, Pin 3 = Cathode (-) Pin 2 (centre) = Anode (+)	Without cable Sullins #GRPB031VWVN-RC

Table 67: Fiber-optic Cannula + Single LED Specifications

Color	Central Wavelength (nm)	Typical Output Power @150 mA (mW) [*]	LED Color Code
Blue	465	8.0	465
Green	525	3.5	525
Amber	595	1.5	595
Red	635	4.0	635

Note: A bare cable connection is provided to interface with a current source.

 $^{^*}$ Power coupled into 200 μm core, NA 0.66 optical fiber.

LED Color Code -(see Table 68)

Fiber-optic code^{*} _____ 200/250-0.66 or 480/500-0.63

Fiber length (mm)–

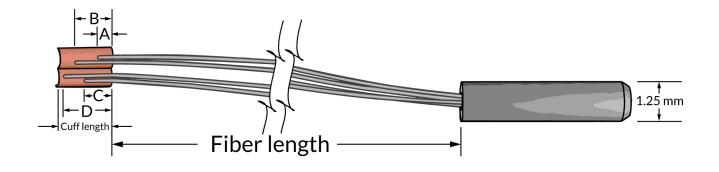
Optical Fiber Cuffs

Optical Fiber Cuffs are designed to perform optogenetic excitation/inhibition on muscles and/or nerve fibers. Light and flexible plastic optical fibers terminated with an angled mirror (MA45 tip) allow multiple illumination spots around fibrous tissues. Our design provides a polyimide based cuff composed of two half-cylindrical parts that can surround the muscle or the nerve. At



Optical Fiber Cuffs

the opposite end, all fibers are bundled into a standard ferrule to allow connection to a patch cord.



Optical Fiber Cuffs Schematic

Note: Complete the ordering code below. Use the PDF on our website to indicate the depth of each optical fiber in the cuff and the distance from the edge of the cuff to the end of the fiber.

^{*}Contact us if different specification is required

	;□_□ 1	ZF1	.25
Number of fibers 2, 4 or 6			
Fiber-optic code			
Fiber length (mm) ——————————————————————————————————			
Cuff length (mm) 1.0 to 10.0 (± 0.5)]		
Cuff internal diameter (mm) ———— 1.0 , 1.5 or 2.0			
Connector type ————————————————————————————————————			

Mono Opto-electric

Cannula with a

ZF 1.25 receptacle

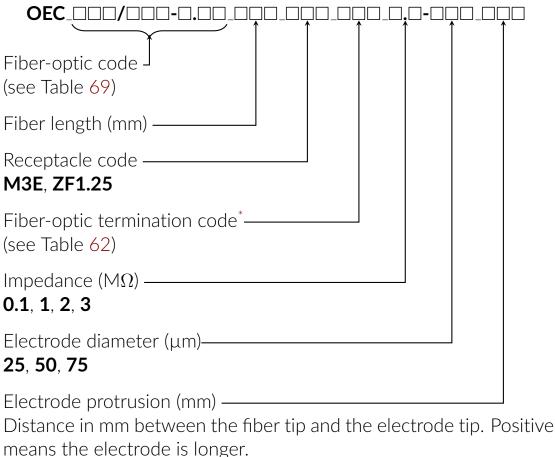
Opto-electric Cannulas

Indispensable when combining optogenetics and electrophysiology in freely moving animals, these chronically implanted cannulas enable optical and electrical contact with a specific group of neurons.

Mono Opto-electric Cannulas

The simplest configuration is made-up of a single 200 μ m core diameter optical fiber of 0.22 NA and 75 μ m diameter and a 3 M Ω impedance metallic wire as electrode. The fiber and the wire are held together by a grooved 1.25 mm zirconia ferrule with a fiber in the central hole and the electrode imbedded in the groove. The optical and electrical contacts with corresponding opto-electric patchcord are maintained with the help of a bronze sleeve.

ORDERING CODE:



^{*}Angled (A45, A60) and mirror (MA45) tips are not offered with Mono Opto-electric Cannulas (see Table 62).

<u>Note</u>: Stereotaxic Cannula Holders and Ferrule Receptacle Adapter are available for implantation to secure the Mono Opto-electric Cannulas (**SCH** and **FCA**; see Tables 73 and 77).

	Core Diameter (µm)	Outer Diameter (μm)	NA	Buffer Color	Outer Layer	Fiber-optic Code
	50	70	0.22	yellow	Polyimide buffer	050/070-0.22
	60	75	0.37	yellow	Polyimide buffer	060/075-0.37
	100	125	0.22	yellow	Polyimide buffer	100/125-0.22
	100	125	0.37	yellow	Polyimide buffer	100/125-0.37
	100	125	0.66	clear	Borosilicate (fragile)	100/125-0.66
	200	240	0.22	yellow	Polyimide buffer	200/240-0.22
_	200	245	0.37	yellow	Polyimide buffer	200/245-0.37
Silica	200	250	0.66	clear	Borosilicate	200/250-0.66
Sil	200	260	0.22	clear	Silicone buffer	200/260-0.22
	300	360	0.37	yellow	Polyimide buffer	300/360-0.37
	300	370	0.22	yellow	Polyimide buffer	300/370-0.22
	400	430	0.66	clear	Borosilicate	400/430-0.66
	400	470	0.37	yellow	Polyimide buffer	400/470-0.37
	400	480	0.22	yellow	Polyimide buffer	400/480-0.22
	600	710	0.37	yellow	Polyimide buffer	600/710-0.37

 Table 69: Fiber-optic Codes for Mono Opto-electric Cannulas

Opto-electric Cannula Arrays for Recording and/or Stimulation

Our Opto-electric Cannula Arrays combine optogenetics and/or photometry with multi-site electrophysiology in freely-moving animals. Our opto-electric cannula arrays consist of an optical fiber (200/250 μ m NA 0.66) surrounded by up to 32 electrodes. This product allows electrophysiological recording and/or stimulation depending on the electrode type. We offer tungsten electrodes (diameter of 25 or 50 μ m) for single-, multi- or field potential recording and platinium/iridium electrodes (diameter of 100 μ m) for electrical stimulation. Custom configurations are available upon



Opto-electric Cannula Arrays

request, like the type of electrodes, the number of optical fibers and the disposition of the optical fibers and electrodes.

ORDERING:

Contact our sales department (sales@doriclenses.com)

Opto-fluid Cannulas

As the convergence of different techniques for cell monitoring like optogenetics, electrophysiology and fluorescence gathers speed, the cannula hybridization and fluid administration becomes imperative. For classification purposes we consider a cannula as port of entry that can be chronically implanted while injectors connect to and disconnect from cannulas. The simplest way to allow passage of liquids, optical and electrical signals in and out of the body is by using universal guiding cannula with a plastic body in the shape of a receptacle and a shaft.

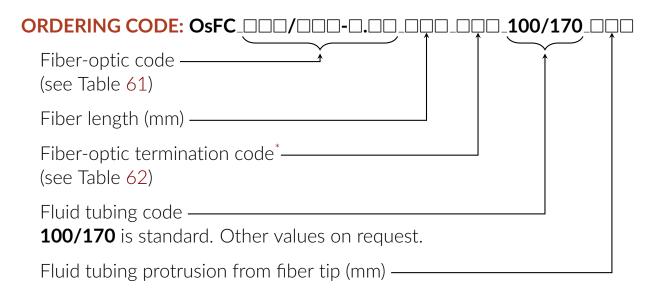
Mono Opto-fluid Cannulas

Single-shot Fluid Injection Cannulas

Optogenetics experiments often require introduction of virus born opsins near targeted cells or neurons that will be subsequently activated or silenced by light. The *Mono Optofluid Cannula* for *Single-shot Fluid injection* has an optical fiber and a side tubing that should be pre-loaded with a virus. The virus is injected after the cannula implantation surgery. Upon the first injection, the liquid passage is often clogged and for this reason the second injection is not recommended and should not be planned. This cannula connects to the liquid delivery system with a plastic tube that attaches to the metal tube on the cannula.



Single-shot Fluid Injection Cannula

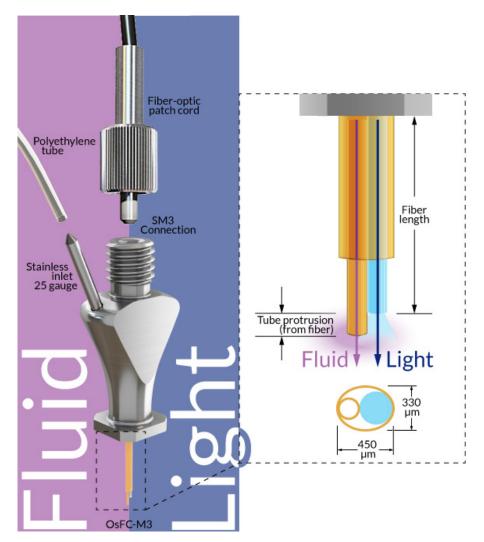


^{*}Angled (A45, A60) and mirror (MA45) tips are not offered with Mono Opto-fluid Cannulas (see Table 62).

Cannulas

Notes:

- Stereotaxic Cannula Holders and Receptacle adapters are available for implantation to secure the Single-shot Fluid Injection Cannula (SCH and FCA; see Tables 73 and 76).
- A 2-meter length of polyethylene tube is sold separately to connect a fluid delivery system to the cannula (**PT_OFC_2**, see Table 121).



Connection diagram and protrusion geometry of the Single-shot Fluid Injection Cannula

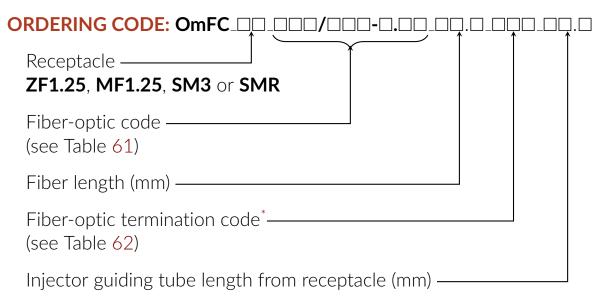
Multiple Fluid Injections Cannulas

This cannula is used for repeated drug or light sensitive dye injections and has a continous fluid path that permits multiple insertion of a liquid loaded micro-injector. The length of optical fiber and micro-injector can be precisely defined to reach targeted brain region. The post surgery fluid injection requires the use of an external micro-injector needle, pre-loaded with pharmacological agents, viruses or plasmids. The fluid delivery can start when the injector is fully inserted into the cannula guiding tube. The fluid injector consists of a 1.25 mm ferrule and corresponding sleeve connector. The cannula includes a plug with ZF sleeve to fill the guiding tube when the micro-injector is not



Multiple Fluid Injections Cannula

in place. The plug is 100 μ m longer than the guiding tube protrusion.



Notes:

- A specially designed holder is available for implantation to secure the Multiple Fluid Injections Cannula (SCH_OmFC).
- The Fluid Injector for the Multiple Fluid Injections Cannulas must be ordered separately (FI_OmFC).
- A 2-meter length of polyethylene tube is sold separately to connect a fluid delivery system to the cannula (**PT_OFC_2**, see Table 121).

^{*}Angled (A45, A60) and mirror (MA45) tips are not offered with Mono Opto-fluid Cannulas (see Table 62).

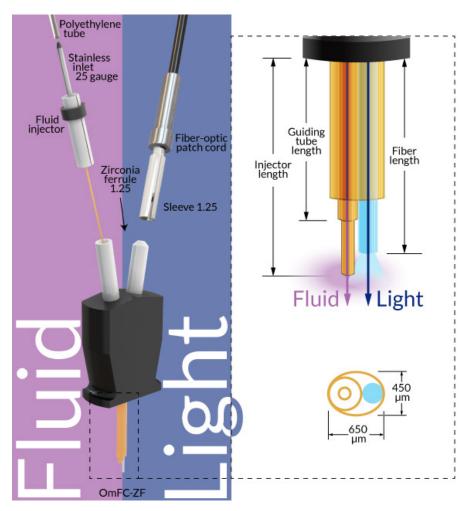
	Core Diameter (µm)	Outer Diameter (μm)	NA	Buffer Color	Outer Layer	Fiber-optic Code
	50*	70	0.22	yellow	Polyimide buffer	050/070-0.22
Silica	60*	75	0.37	yellow	Polyimide buffer	060/075-0.37
	100	125	0.22	yellow	Polyimide buffer	100/125-0.22
Sill	100	125	0.37	yellow	Polyimide buffer	100/125-0.37
	100	125	0.66	clear	Borosilicate (fragile)	100/125-0.66
	200	240	0.22	yellow	Polyimide buffer	200/240-0.22
	200	245	0.37	yellow	Polyimide buffer	200/245-0.37
	200	250	0.66	clear	Borosilicate	200/250-0.66
	300	360	0.37	yellow	Polyimide buffer	300/360-0.37
	300	370	0.22	yellow	Polyimide buffer	300/370-0.22
	400	430	0.66	clear	Borosilicate	400/430-0.66
	400	470	0.37	yellow	Polyimide buffer	400/470-0.37
	400	480	0.22	yellow	Polyimide buffer	400/480-0.22
	600	710	0.37	yellow	Polyimide buffer	600/710-0.37
U						
Plastic	240* † ‡	250	0.63	clear	PMMA	240/250-0.63
Pla	480*,† ‡	500	0.63	clear	PMMA	480/500-0.63

Table 70: Fiber-optic Codes for Multiple Fluid Injection Cannulas

^{*}Not offered with a conical tip (**C45**, **C60**; see Table 62).

^{\dagger}Not offered with a 45° mirror tip (**MA45**; see Table 62

[‡]Only for fiber lengths inferior to 5 mm



Connection diagram and protrusion geometry of the Multiple Fluid Injections Cannula

Fluid Injector for Multiple Fluid Injections Cannulas	
ORDERING CODE: FI_OmFC-	Ť
Receptacle ZF, SM3 or SMR	
Injector length (mm)	

Fluid Injector for Multiple Fluid Injections Cannulas

Opto-fluid Cannula with interchangeable injectors

The Opto-fluid Cannula with interchangeable injectors provides a simple way to use both light and fluid injection when they are not required at the same time. The interchangeable configuration saves space and weight and can be used with optical and fluid injector of different lengths. The threaded body ensures a secure connection for the injectors.

Each cannula ships with a plug to prevent the guiding tube from clogging at implantation and between uses. Since both the optical and fluid injector does not stay inside the brain for a prolonged time, the plug is placed back when the cannula is not in use to keep it free of biological debris.



Opto-fluid Cannula with interchangeable injectors

Optical injector	Guidir			
Fiber diameter (μm) (see table 72)	Inner diameter (µm)	Outer diameter (µm)	Guiding Tube Code	
70 to 125	250	350	250/350	
230 to 250	320	430	320/430	
360 to 500	530	660	530/660	

Table 71: Guiding Tube Codes for Opto-fluid Cannulas with interchangeable injectors

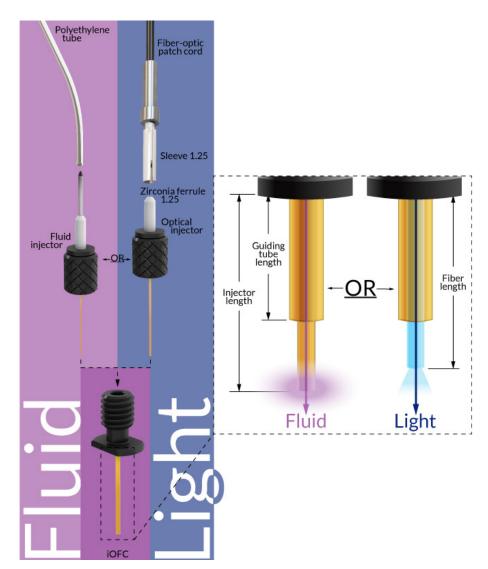
ORDERING CODE: iOF	C_M3/	
		· · ↑

Guiding tube code – (see Table 71)

Guiding tube length from receptacle (mm) —

Notes:

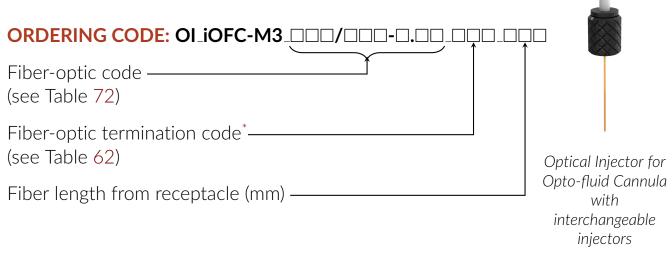
- The tubing internal diameter must match the optical injector external diameter (see Tables 71 and 72).
- A Stereotaxic Cannula Holder is available for implantation to secure the Optofluid Cannula with interchangeable injectors. The plug (included with the cannula) is used to fix the holder on the Opto-fluid Cannula (**SCH**; see Table 73).



Connection diagram and protrusion geometry of the Opto-fluid Cannula with interchangeable injectors

- Optic and Fluid Injectors for Opto-fluid Cannula with interchangeable injectors must be ordered separately (**OI_iOFC** and **FI_iOFC**).
- A 2-meter length of polyethylene tube is sold separately to connect a fluid delivery system to the cannula (**PT_OFC_2**, see Table 121).

Optical injector for Opto-fluid Cannula with interchangeable injectors



	Core Diameter (µm)	Outer Diameter (µm)	NA	Buffer Color	Outer Layer	Fiber-optic Code
	50	70	0.22	Yellow	Polyimide buffer	050/070-0.22
	60	75	0.37	Yellow	Polyimide buffer	060/075-0.37
	100	125	0.22	Yellow	Polyimide buffer	100/125-0.22
	100	125	0.37	Yellow	Polyimide buffer	100/125-0.37
	100	125	0.66	Clear	Borosilicate (fragile)	100/125-0.66
	200	240	0.22	Yellow	Polyimide buffer	200/240-0.22
Silica	200	245	0.37	Yellow	Polyimide buffer	200/245-0.37
Sill	200	250	0.66	Clear	Borosilicate	200/250-0.66
	300	360	0.37	Yellow	Polyimide buffer	300/360-0.37
	300	370	0.22	Yellow	Polyimide buffer	300/370-0.22
	400	430	0.66	clear	Borosilicate	400/430-0.66
	400	470	0.37	yellow	Polyimide buffer	400/470-0.37
	400	480	0.22	Yellow	Polyimide buffer	400/480-0.22
<u>.</u>						
Plastic	240†	250	0.63	Clear	PMMA	240/250-0.63
Pla	480†	500	0.63	Clear	PMMA	480/500-0.63

Table 72: Fiber-optic Codes for Optical Injectors

^{*}Angled (**A45**, **A60**) and mirror (**MA45**) tips are not offered with Mono Opto-fluid Cannulas (see Table 62). [†]Only for fiber lengths inferior to 5 mm

Fluid Injector for Opto-fluid Cannula with interchangeable injectors

ORDERING CODE: FI_iOFC-M3_100/170_

Injector length from receptacle (mm) -

Dual Opto-fluid Cannulas

Dual Opto-fluid Cannula with interchangeable injectors

The precise pitch of the *Dual Opto-fluid Cannula with interchangeable injectors* guarantees an optimal bilateral implantation where both light and fluid injection can be used. The interchangeable configuration saves space and weight and can be used with multiple lengths of optical and fluid injector.

Each cannula ships with two plugs to prevent the guiding tubes from being clogged at implantation and between uses. Since both the optical and fluid injectors do not stay inside the brain for a long time, this cannula has the additional benefit of keeping them free of biological debris.

Depending on the pitch between the two optical fibers, two receptacle types are offered: small (S) and large (L). The model S is required when the pitch is between the guiding tube outer diameter and 1.7 mm. For pitch more than 1.7 mm, the model L is needed.

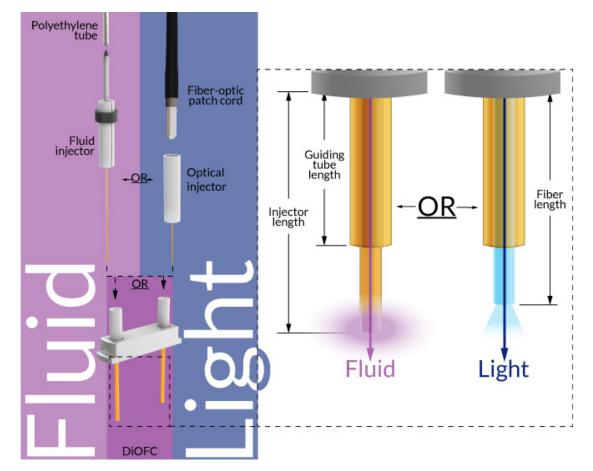
Receptacle type — S or L	
Center-to-center pitch (mm) Between the 2 guiding tubes	
Guiding tube code —————————— (see Table <mark>71</mark>)	
Guiding tube length from receptacle (mm) —	

Fluid Injector for Opto-fluid Cannula with interchangeable injectors



Dual Opto-fluid Cannula (L) with interchangeable injectors

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Connection diagram and protrusion geometry of the Dual Opto-fluid Cannula with interchangeable injectors

Notes:

- For the model S, the minimum possible distance between the tubes is the guiding tube diameter.
- Optical and Fluid Injectors for Dual Opto-fluid Cannulas must be ordered separately (OI_DiOFC and FI_DiOFC). Be careful to order the right amount of injectors.
- The tubing internal diameter must match the optical injector external diameter (see Tables 71 and 72).
- A 2-meter length of polyethylene tube is sold separately to connect a fluid delivery system to the cannula (**PT_OFC_2**, see Table 121).

Optical Injector for Dual Opto-fluid Cannula with interchangeable injectors	5
Receptacle type S or L (depending on DiOFC receptacle)	Optical Injector for
Fiber-optic code(see Table 72)	Dual Opto-fluid Cannula with interchangeable
Fiber-optic termination code [*]	injectors
Fiber length from receptacle (mm)	

Fluid Injector for Dual Opto-fluid Cannula with interchangeable injectors

Receptacle type _____

S or **L** (depending on DiOFC receptacle)

Injector length from receptacle (mm) ------

^{*}Angled (A45, A60) and mirror (MA45) tips are not offered with Dual Opto-fluid Cannulas (see Table 62).

Stereotaxic Tools

Stereotaxic arm is valued for its positioning precision. However, when it comes to positioning the fiber-optic cannula, some of the precision is lost when attaching or detaching the cannula to/from the arm. To simplify the implantation of the cannula and maintain the precision, we have developed a *Stereotaxic Cannula Holder* and *Fiber-optic Cannula Adapters* for attaching our cannulas to the stereotaxic arm.

Stereotaxic Cannula Holders



Stereotaxic Cannula Holder 1.25

Notes:

- The diameter of the *Stereotaxic Cannula Holder* is 6.35 mm. Its length is 7.9 cm and an adapter of 10 cm long can be added at one end (**SIA**; see Table 74).
- An optional clamp (**SCL**) can be used to fix the Stereotaxic Cannula Holder on stereotaxic apparatus.
- The Stereotaxic Cannula Holder 1.25 mm allows the implantation of Two-ferrule Cannula with a pitch of 1.8 mm.

Table 73: Stereotaxic Cann	ula Holders Ordering Codes
----------------------------	----------------------------

Ferrule Diameter (mm)	Ordering Code	
1.25	SCH_1.25	
2.5	SCH_2.5	

Cannulas

OmFC Cannula Holder

The OmFC Cannula Holder is an adapter designed to maintain the OmFC cannula protrusion straight during the implantation. The upper part is compatible with our *In-line Adapters*. The bottom side holds the OmFC cannula steady with a screw. The configuration of the holder allows the use of a fluid injector and/or an optical connection during the implantation.

Notes:

- An adapter of 10 cm long can be added at one end of the holder (SIA; see Table 74).
- An optional clamp (SCL) can be used to fix the Stereotaxic Cannula Holder on stereotaxic apparatus.

ORDERING CODE: SCH_OmFC_

Receptacle of the cannula — ZF or SM3

Low Profile Cannula Holder

This stereotaxic holder is related to low profile cannulas. It is used to keep the fiber along the dorso-ventral axis during the implantation. Other inclined holders are available to be compatible with custom angle cannulas. The diameter of the holder is 7.9 mm.

Notes:

- An adapter of 10 cm long can be added at one end of the Low Profile Cannula holder (SIA; see Table 74).
- An optional clamp (**SCL**) can be used to fix the Stereotaxic Cannula Holder on stereotaxic apparatus.

ORDERING CODE: SCH LP90

OmFC ZF Cannulas Holder

Holder - 90°





Adapters for Stereotaxic Cannula Holders

We offer adapters to attach our stereotaxic holders on stereotaxic frames that use 7.9 mm or 5 mm diameter rods as standard. We can offer other adapters on request.

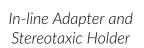
In-line Adapter

This adapter consists of a rod, with 8-32 threads at one end. Notes:

- The In-line Adapters are 10 cm long and compatible with our Stereotaxic Cannula Holders.
- An optional clamp (**SCL**) can be used to fix the Stereotaxic Cannula Holder on stereotaxic apparatus.
- For diameter 7.9 mm, there is a 1/4'' threaded hole at the other end.

Table 74: In-line Adapters Ordering Codes

Diameter (mm)	Ordering Code	
5	SIA_5	
6.35	SIA_6.35	
7.9	SIA_7.9	



Clamp

This adapter is a double clamp, designed to hold a 5.0 or 7.9 mm diameter rod on one side and a 6.35 mm diameter rod on the other side.



Clamp

Note: The Clamp is compatible with our Stereotaxic Cannula Holders (**SCH**), OmFC Cannulas Holder (SCH_OmFC), Low Profile Cannulas Holder (SCH_LP90) and Inline Adapters (SIA).

Table 75: Clamp Orderir	ng Codes
-------------------------	----------

Diameter (mm)	Ordering Code	
5.0	SCL_5	
7.9	SCL_7.9	

Receptacle adapters

M2, M3 or GS Receptacle Adapters

This adapter allows the use of M2, M3 or GS Cannula Receptacles with a Stereotaxic Cannula Holder.



Receptacle Adapter SM3 1.25 mm for Fiber-optic Cannula

	Receptacle Adapter			
Attachment Diameter (mm)	RM2	SM3	GS	
1.25		FCA_1.25_SM3		
2.5	FCA_2.5_RM2	FCA_2.5_SM3	FCA_2.5_GS	

Table 76: Receptacle Adapters Ordering Codes

Note: GS for a Dual Fiber-optic Cannula with a guiding socket.

Ferrule Receptacle Adapter

This adapter allows an extension of the ferrule in cases where the clearance on the receptacle is insufficient to use the *Stereotaxic Cannula Holder*. This adapter can be useful to implant all types of Opto-electric Cannula Receptacles.



Ferrule Receptacle Adapter

Table 77: Ferrule Receptacle Adapters Ordering Codes

Diameter (mm)	Ordering Code
1.25	FCA_1.25_1.25
2.5	FCA_2.5_2.5

In vitro and In vivo (head-fixed animal) Illumination

Optical Fiber Probes

Instead of cannulation, *in vitro* and *in vivo* head-fixed animal optogenetics experiments require thin and long optical probes that easily connect to micro-manipulator probe holders and have minimal obstruction of the observation site.

Optical Fiber Probe Holder

Our Optical fiber probe holder is a stainless steel rod having an FC receptacle on one end that allows a light delivery patch cord to be plugged in and at the other end an M3 receptacle where a probe can be screwed on. The two receptacles are mutually connected with an internal optical fiber housed within the 6.35 mm diameter rod that fits most popular micro-manipulators. To avoid unnecessary optical losses, the selected optical fiber parameters such as the core diameter and NA should match those of



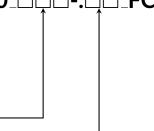
Optical Fiber Probe Holder

the connecting fibers. The type of fiber within the holder is marked with a color code. The fiber NA is engraved on the holder.

Note: An optional clamp (**SCL**) can be used to fix the holder on stereotaxic apparatus.

ORDERING CODE: OFPH_150_□□□-.□□_FC

Rod length (mm) **150** mm is standard. 100 mm is also available. Fiber-optic core diameter (μm) -

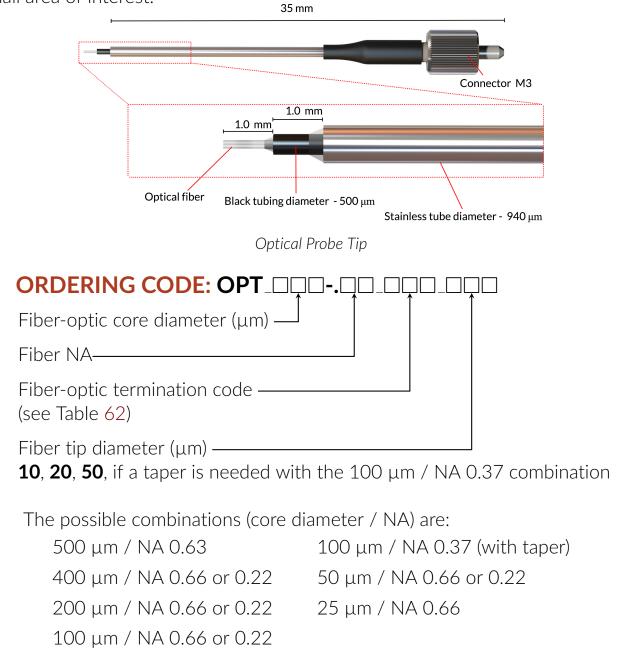


Fiber NA -

Possible fiber combinations are : 500 μm / NA 0.63 for LED sources, 200 μm / NA 0.22 for laser sources and 50 μm / NA 0.22 for small area laser illumination.

Optical Probe Tips

Like fiber-optic cannulas, the optical probe is an optical fiber of the specific NA and core diameter with an M3 connector at one end while its loose end is much longer and suitably protected and strengthened to keep its direction. The fiber tip can even be pulled to diameters smaller than the original fiber diameter. When used with micro-manipulators and its holders, it can precisely illuminate a very small area of interest.



<u>Note</u>: For the 100 μm / NA 0.37 combination, tapered tips down to 10 μm are available upon request.

Opto-electric Probes

Opto-electric Probe Holder

BNC or **pin**

The Opto-electrical probe holder has a similar function as the optical probe holder with the addition of an electrical contact. In order to reduce cable congestion around the specimen, the opto-electrical probe holder permits bringing the optical and electrical contact to the back of the holder. It is recommended to plug a shielded cable to the BNC cable.



ORDERING CODE: OEPH OEPH OEPH OEPH OEEPH OEEPH

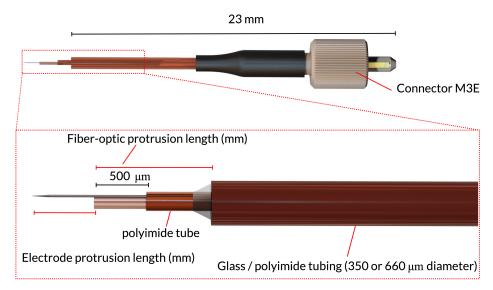
Opto-electric Probe Holder

Notes:

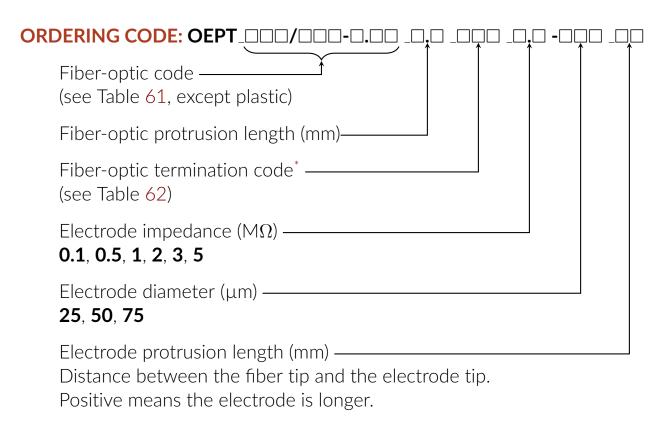
- The length of the optical fiber and the electrical cable is measured from the coming out of the rod to the tip of the connector.
- The rod has a standard length of 150 mm.
- An optional clamp (**SCL**) can be used to fix the holder on stereotaxic apparatus.

Opto-electric Probe Tips

Like optical probe tips, the *Opto-electric Probe Tips* have an optical fiber of the specific NA and core diameter with an M3 connector at one end while its loose end is much longer and suitably protected and strengthened to keep its direction. In addition, this probe has an electrical wire that goes along the optical fiber, from near the tip to the groove in the zirconia ferrule.



Opto-electric Probe Tip



*Angled (A45, A60) and mirror (MA45) tips are not offered with Opto-electric Probe Tips (see Table 62).

Single-cell Recording Opto-electric Probe

Single-cell recordings require an optical fiber core diameter at the fiber end comparable with the size of the cell under observation. The electrode tip has to be of the similar size and in close proximity to the fiber core. One way to achieve those specifications is by making a dual core optical fiber having the light guiding core and the capillary within its cladding^{*}[†], and pulling or tapering one fiber end into the small diameter tip. When the capillary is filled with electrolyte, the fiber end becomes a usable single-cell opto-electric interface smaller than the cell itself.

These Single-cell Recording Opto-electric Probes are perfectly suited for *in vivo* single-cell electrophysiological recordings, optogenetic stimulation and photometry monitoring (see animation by Stuart Jantzen from University of Toronto).

<u>NB:</u> This particular probe is used for *in vivo* experiments with head-fixed animals.



Single-cell Recording Opto-electric Probe

Single-cell Recording Opto-electric Probe Systems

This section contains complete Single-cell Recording Opto-electric Probe systems bundled with a single ordering code for convenience.

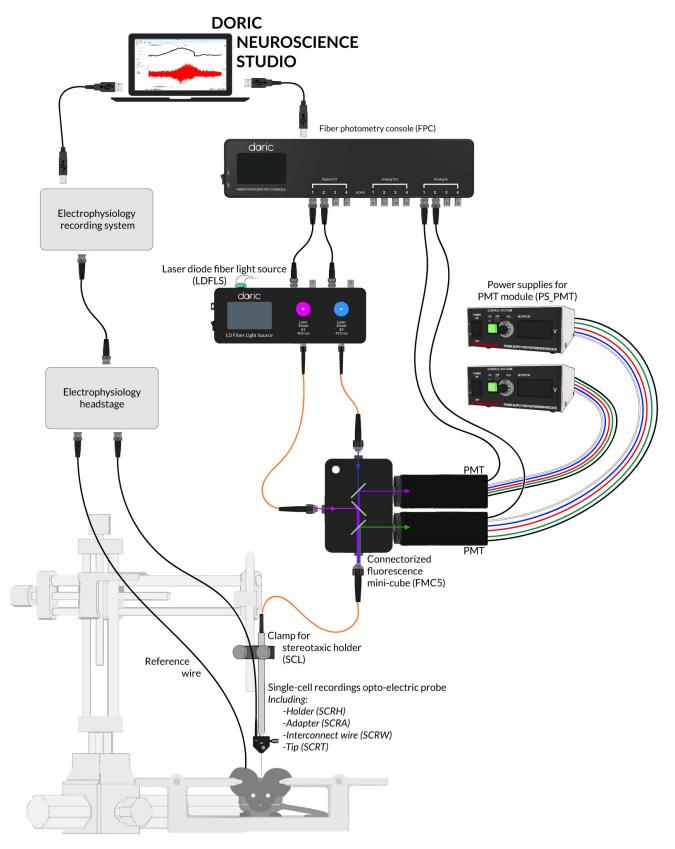
Single-cell Photometry and Electrophysiology Recording System

This system is designed to do extracellular electrophysiology recordings and photometry detection at a single-cell resolution in a head-fixed configuration. Its optical sensitivity allows detection of standard fluorophores (GFP, mCherry, quantum dot, etc.) or functional fluorophores (GCaMP, Oregon Green, FURA-2, etc). Extracellular electrophysiology is possible by filling the fiber probe hollow core with an electrolyte solution to get an electrode impedance in the range of 1 to 20 M Ω , allowing a single-unit or multi-unit spikes detection.

This system includes specifically:

- Fiber Photometry Console
- Connectorized Fluorescence Mini Cube (single or multiple wavelengths with appropriate filters)
- Photosensor Module (1 or 2x)
- Power Supply for PMT Module (1 or 2x)
- Laser Diode Fiber Light Sources (1 or 2x)
- Single-cell Opto-electric Probe Holder
- Clamp for Single-cell Opto-electric Probe Holder
- Single-cell Opto-electric Probe Adapter
- Single-cell Opto-electric Probe Interconnect Wire (5x)
- Single-cell Opto-electric Probe Tips (20x)
- Extracellular Electrophysiology Recording System (under development)
- All electrical cables and optical patch cords

ORDERING CODE: SCRS-PE



Single-cell Photometry and Electrophysiology Recording System

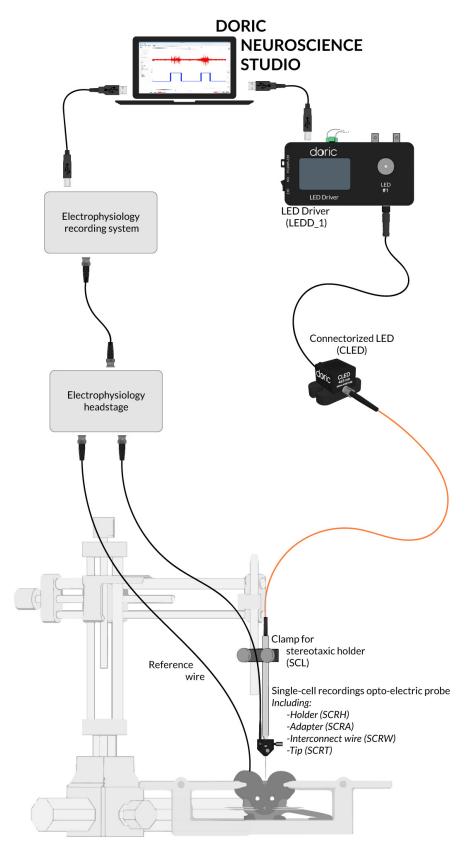
Single-cell Optogenetic Illumination and Electrophysiology Recording System

Optogenetics and electrophysiology are combined in this system to allow recordings of synaptic events at a single-cell resolution in a head-fixed configuration. It is designed to activate optogenetic proteins such as channelrhodopsine or to inhibit light-gated ion pumps like halorhodopsin. By adjusting the illumination output power, it is possible to illuminate a single cell or a group of neurons. Extracellular electrophysiology is possible by filling the fiber probe hollow core with an electrolyte solution to get an electrode impedance in the range of 1 to 20 M Ω , allowing a single-unit or multi-unit spikes detection.

This system includes specifically:

- Connectorized Single LED (1x or 2x)
- LED Driver
- Single-cell Opto-electric Probe Holder
- Clamp for Single-cell Opto-electric Probe Holder
- Single-cell Opto-electric Probe Adapter
- Single-cell Opto-electric Probe Interconnect Wire (5x)
- Single-cell Opto-electric Probe Tips (20x)
- Extracellular Electrophysiology Recording System (under development)
- All electrical cables

ORDERING CODE: SCRS-OE



Single-cell Optogenetic Illumination and Electrophysiology Recording System

Single-cell Recording Opto-electric Probe Holder

The Single-cell Recording Opto-electric Probe Holder consists of a fiber-optic patch cord in a rigid tubing. It offers an appropriate optical connection between the probe core in the Single-cell Opto-electric Probe Adapter and the light module. It can be secured in a stereotaxic apparatus with our Stereotaxic Clamp.



Single-cell Recording Opto-electric Probe Holder

ORDERING CODE: SCRH_550/600/3000-0.22____FCA

Fiber-optic code -

Optical fiber length (m) — **1 to 1.5** m is recommended.

Fiber-optic patch cord termination –

Notes:

- The length of the optical fiber is measured from the coming out of the rod to the tip of the connector.
- The rod has a standard length of 150 mm.
- An optional clamp (**SCL**) can be used to fix the holder on stereotaxic apparatus.

Single-cell Recording Opto-electric Probe Adapter

The link between the Single-cell Opto-electric Probe Holder and the Single-cell Opto-electric Probe Tip is assured by the Single-cell Opto-electric Probe Adapter. This junction component maintains and aligns the two optical cores together allowing a maximum amount of light to be guided towards the tip. Its electrical output ensures the electrical connection between the electrophysiological recording



Single-cell Recording Opto-electric Probe Adaptor

system and the electrolyte filled core. The electrical ends with a standard BNC connector can be modified upon request.

Electrical connector — **BNC** or **pin**

Cable length (m) –

0.2 m is standard. Short length is recommended.

Single-cell Recording Opto-electric Interconnect Wire

The Single-cell Opto-electric Interconnect Wire makes the connection between the electrolyte solution within the optical fiber hollow core and the electrophysiology system. The part inserted in the Singlecell Opto-electric Probe Tip is a 100 μ m silver wire and the other side is connected to the probe adaptor with a pip. Other types of wires could be offered



Single-cell Recording Opto-electric Interconnect Wire

tor with a pin. Other types of wires could be offered on demand.

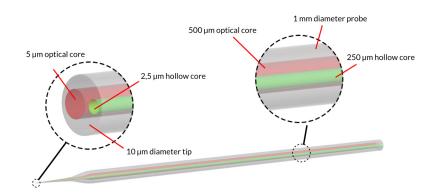
Type of wire ______ Silver wire 100 μm (**AG100**) or stainless steel wire 50 μm (**SS50**)

Single-cell Recording Opto-electric Probe Tips



A Single-cell Recording Opto-electric Probe Tip is a piece of dual core optical fiber with a 500 μ m optical core and a 250 μ m hollow core for electrolyte filling with one end pulled and cut to a 10 μ m diameter tip as illustrated above. The probe tip is simply inserted into the dedicated Single-cell Recording Opto-electric Probe Holder. A black coating can be added to minimize the light output/input in the shoulder of the tapered part of the probe.

Single-cell Recording Opto-electric Probe Tip



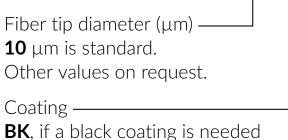
Single-cell Recording Opto-electric Probe Tip of 10 μ m diameter Table 78: Single-cell Recording Opto-electric Probe Tips Specifications

SPECIFICATION	VALUE	NOTE
Optical fiber diameter	1 mm	
Optical fiber core diameter	500 µm	Off-Centered
Optical fiber NA	0.23	
Hollow core diameter	250 µm	For electrolyte filling, off-centered
Pulled tip diameter	10 µm	Optical core and hollow core ratio is preserved
Shank (taper) length	3-7 mm	
Total length	30 - 35 mm	

Note: Opto-electric probe tips are sold in lots of 20 units.

SCRT_10_

ORDERING CODE:



DR, IT a black coating is needed

Single-cell Recording Opto-electric Probe - Raw Fiber

Silica raw dual core fiber in lengths of 55 mm are available for proficient users who prefer to pull and cut the tip by themselves. Suitable with Sutter Instrument P-2000 Laser-based micropipette puller.

ORDERING CODE: SCRF_55

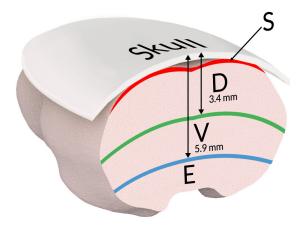
Fiber length (mm) — **55** mm is standard.

Single-cell Recording **Opto-electric Probe** - Raw Fiber

Miniaturized Fluorescence Microscopy

Until recently, fluorescence microscopy was dominated by large microscope installations, sometimes referred to as the rigs. The observations of neural circuitry in freely moving animals like mice or rats require a wearable fluorescence microscope attached to imaging cannulas chronically implanted in the animal's brain. To make this microscope mice-wearable, the smallest fluorescence microscope body ever was built. It easily snaps into a chronically implanted imaging cannula via a self-centering latching mechanism. The snap-in microscope body is electrically pigtailed and optically connectorized. In the middle of the visible spectrum, the scattering through the brain tissue limits imaging to about 150 μ m. The imaging limited to those depths from the brain surface can be performed without insertion of all-glass relay lenses. At larger brain depths, it is absolutely necessary to use relay lens systems that may consist of homogeneous or gradient-index glass rods or lenses that bring the image into focus of the microscope objective and effectively reduce the optical path through the brain tissue.

Here are some simple rules for selecting the appropriate microscope body and imaging cannula design when imaging different brain tissue zones:

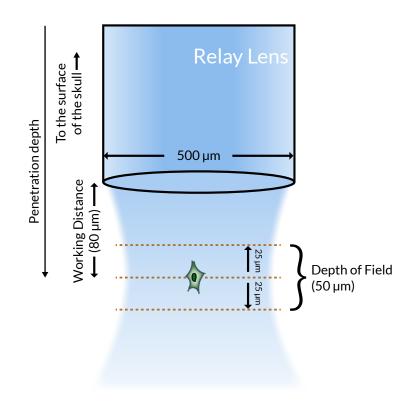


- S: Surface (up to 150 µm deep) D: Deep V: Very deep
- E: Extremely deep

Brain Zones	Microscope Body Model	Cannula Model
(S) 0 to 150 μm below the brain surface	S	S
(D) 0 to 3.4 mm below the skull surface *	L	L type D
(V) 3.0 to 5.9 mm below the skull surface *	L	L type V
(E) 5.4 to 8.3 mm below the skull surface*	L	L type E

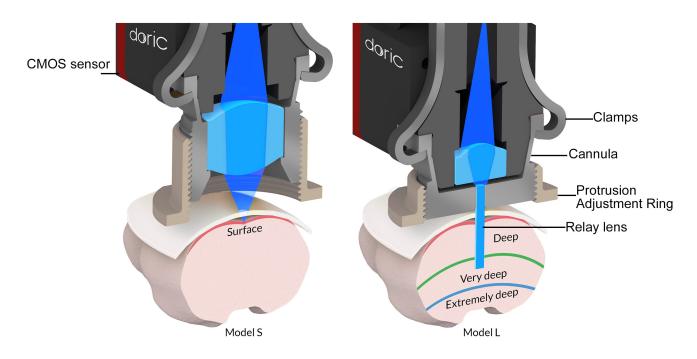
Table 79: Microscope Body and Imaging Cannula models for specific brain zones

According to different experimental requirements, Doric Lenses provides several options of miniature fluorescence microscope systems assemblies. Different systems achieve desired working distance (WD) by different means. While in surface imaging systems, depth adjustments can simply be done by using different sized protrusion rings, in Snap-in systems (deep brain) the position of GRIN lens in cannula is custom aligned to achieve desired working distance. Our standard snap-in system has 80 μ m WD, other WD can be custom ordered.

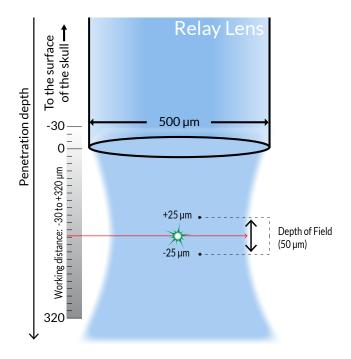


Snap-in L Imaging Cannula working distance and depth of field

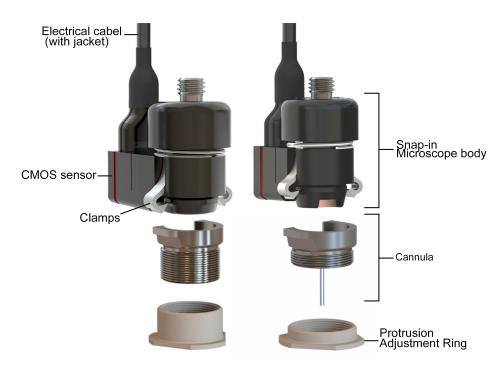
^{*} Including the thickness of the skull.



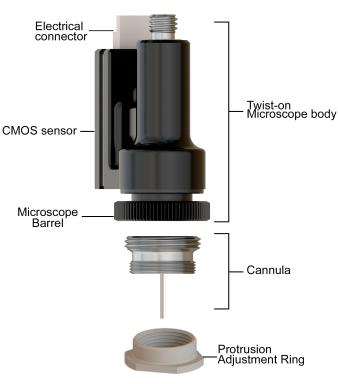
In efocus systems, we use electrowetting lenses (liquid lenses) for electronic focus adjustments. The working distance has an adjustment range of $350 \mu m$: from $-30 \mu m$ (inside the GRIN rod lens) to $320 \mu m$.



Twist-on efocus Imaging Cannula working distance and depth of field



Snap-in Fluorescence Microscope Model S (left), and Model L (right)



TwistOn Fluorescence Microscope Model

Note:

The electrical cable jacket can be customized with a lighter and more flexible cable, the *Ultralight Fiberglass Jacket*, or **UFGJ**, or a more robust but heavier one, the *Lightweight Metal Jacket*, or **LWMJ**.

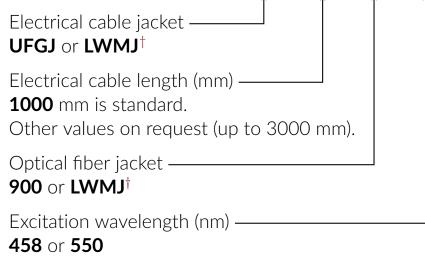
Miniaturized Fluorescence Microscopy Systems

Basic Fluorescence Microscopy Systems

For Surface Imaging (<150 μ m depth)

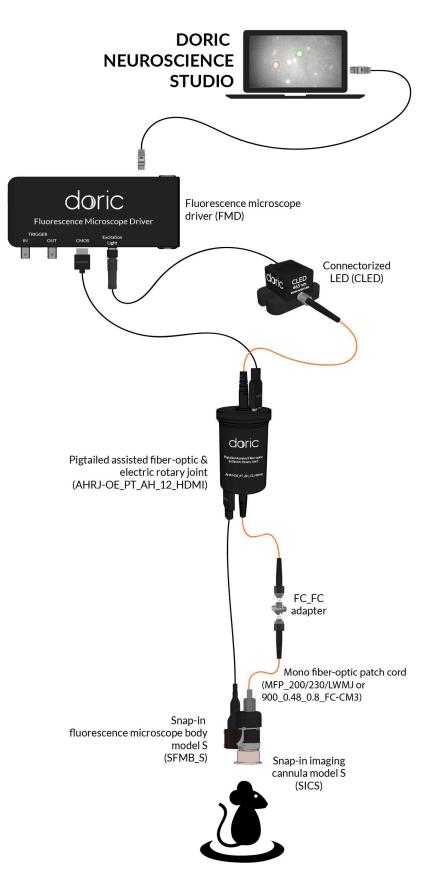
This system contains all the items necessary to do surface brain calcium imaging of freely-moving animals. This system includes specifically:

- Connectorized LED or Ce:YAG Optical Head
- Fluorescence Microscope Driver
- Snap-in Fluorescence Microscope Body Model S
- Snap-in Imaging Cannula Model S (3x)
- Protrusion Adjustment Ring Set Model S
- Pigtailed Assisted Fiber-optic & Electric Rotary Joint
- Fluorescence Microscope Holder
- Clamp for Fluorescence Microscope Holder
- Fluorescence Microscope Snapping Tool
- Dummy Microscope
- Doric Neuroscience Studio for system control and analysis
- All required electrical cables and optical patch cords*



^{*}The optical fiber length is adjusted to fit the desired electrical cable length.

[†]The Ultralight Fiberglass Jacket (UFGJ) and the 0.9 mm Hytrel Jacket (900) are lighter and more flexible, while the Lightweight Metal Jacket (LWMJ) is more robust but heavier.



Basic Fluorescence Microscopy System for Surface Imaging of GCaMP6

2-color Fluorescence Microscope Systems

For Surface Imaging ($<150 \ \mu m \ depth$)

This system contains all the items necessary to do surface brain calcium imaging with GFP-like and RFP-like fluorophores of freely-moving animals.

This system includes specifically:

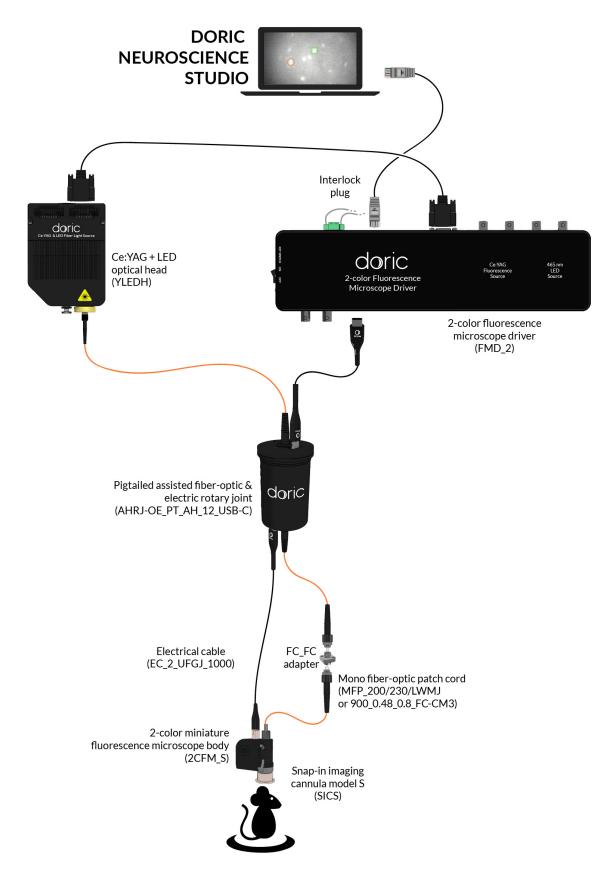
- Ce:YAG + LED (465 nm) Optical Head
- 2-color Fluorescence Microscope Driver
- 2-color Fluorescence Microscope Body Model S
- Snap-in Imaging Cannula Model S (3x)
- Protrusion Adjustment Ring Set Model S
- Pigtailed Assisted Fiber-optic & Electric Rotary Joint
- Fluorescence Microscope Holder 2
- Clamp for Fluorescence Microscope Holder 2
- 2-color Fluorescence Microscope Snapping Tool
- 2-color Dummy Microscope
- Doric Neuroscience Studio for control and analysis
- All required electrical cables and optical patch cords*

ORDERING CODE: 2CMS-S_0_1000_0_458/561

Electrical cable jacket ———— UFGJ or LWMJ [†]	
Electrical cable length (mm) — 1000 mm is standard. Other values on request (up to 3000 mm).	
Optical fiber jacket — 900 or LWMJ [†]	
Excitations wavelengths (nm)	

^{*}The optical fiber length is adjusted to fit the desired electrical cable length.

[†]The Ultralight Fiberglass Jacket (UFGJ) and the 0.9 mm Hytrel Jacket (900) are lighter and more flexible, while the Lightweight Metal Jacket (LWMJ) is more robust but heavier.



2-color Fluorescence Microscopy System for Surface Imaging

For Deep-brain Imaging (150 μm to 8 mm depth)

This system contains all the items necessary to do deep-brain calcium imaging with GFP-like and RFP-like fluorophores of freely-moving animals.

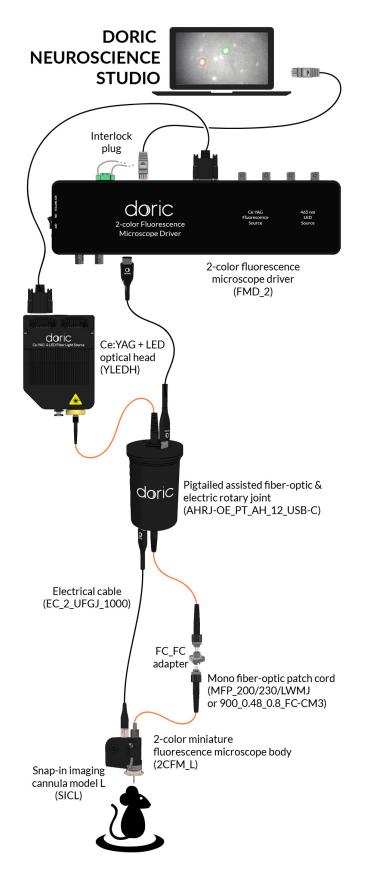
This system includes specifically:

- Ce:YAG + LED (465 nm) Optical Head
- 2-color Fluorescence Microscope Driver
- 2-color Fluorescence Microscope Body Model L
- Snap-in Imaging Cannula Model L (3x)
- Protrusion Adjustment Ring Set Model L
- Pigtailed Assisted Fiber-optic & Electric Rotary Joint
- Fluorescence Microscope Holder 2
- Clamp for Fluorescence Microscope Holder 2
- 2-color Fluorescence Microscope Snapping Tool
- 2-color Dummy Microscope
- Doric Neuroscience Studio for control and analysis
- All required electrical cables and optical patch cords*

Î	\uparrow \uparrow	î	\sim
2-color Fluorescence Microscope Body Optimized for D or V			Ì
Electrical cable jacket ——— UFGJ or LWMJ [†]			
Electrical cable length (mm) – 1000 mm is standard.			
Optical fiber jacket ——— 900 or LWMJ [†]			
Excitation wavelengths (nm) – Excitation 1 / Excitation 2			

^{*}The optical fiber length is adjusted to fit the desired electrical cable length.

[†]The Ultralight Fiberglass Jacket (UFGJ) and the 0.9 mm Hytrel Jacket (900) are lighter and more flexible, while the Lightweight Metal Jacket (LWMJ) is more robust but heavier.



2-color Fluorescence Microscope System for Deep-brain Imaging

Twist-on efocus Fluorescence Microscopy System

For Deep-brain Imaging (150 μ m to 8 mm depth)

This system contains all the items necessary to do deep-brain calcium imaging of freely-moving animals with the possibility of electronic adjustment of the distance between the implant tip and the focal plane.

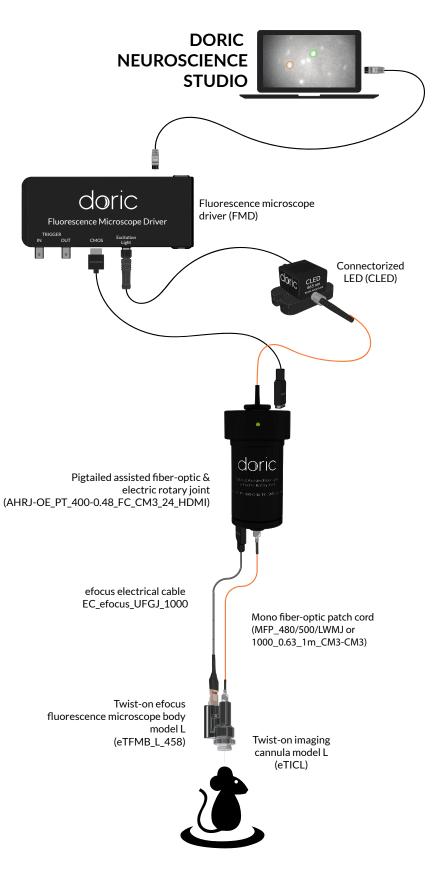
This system includes specifically:

- Connectorized LED (465 nm)
- Fluorescence Microscope Driver
- Twist-on efocus Fluorescence Microscope Body Model L
- Twist-on efocus Imaging Cannula Model L (3x)
- Protrusion Adjustment Ring Set Model L
- Pigtailed Assisted Fiber-optic & Electric Rotary Joint
- Fluorescence Microscope Holder
- Clamp for Fluorescence Microscope Holder
- Twist-on efocus Dummy Microscope
- Doric Neuroscience Studio for control and analysis
- All required electrical cables and optical patch cords*

Electrical cable jacket UFGJ or LWMJ [†]
Electrical cable length (mm) 1000 mm is standard. Other values on request (up to 3000 mm).
Optical fiber jacket 900 or LWMJ [†] Excitation wavelength (nm)
458 Twist-on cannula type D, V or E
GRIN diameter (μm) 500 or 1000

^{*}The optical fiber length is adjusted to fit the desired electrical cable length.

[†]The Ultralight Fiberglass Jacket (UFGJ) and the 0.9 mm Hytrel Jacket (900) are lighter and more flexible, while the Lightweight Metal Jacket (LWMJ) is more robust but heavier.



Twist-on efocus Fluorescence Microscopy System for Deep-brain Imaging of GCaMP6

Twist-on Optogenetically Synchronized Fluorescence Microscopy System

For Deep-brain Imaging (up to 8 mm depth)

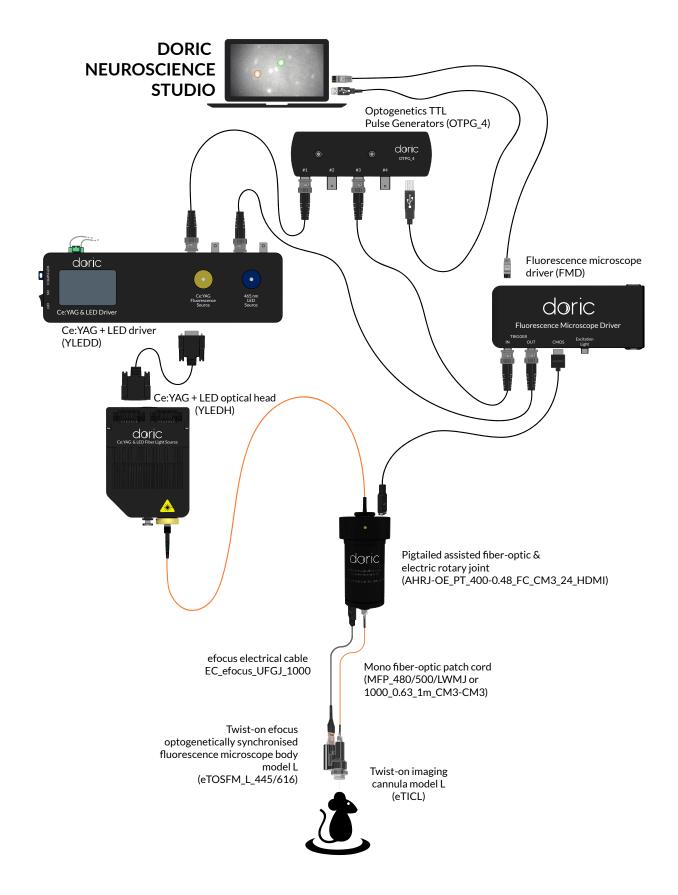
This system contains all the items necessary to do deep-brain calcium imaging synchronized with opsin activation of freely-moving animals with the possibility to adjust electronically the focus position at the tip of the implant. This system includes specifically:

- Ce:YAG + LED (465 nm) Optical Head & Driver with Bandwidth Filter
- Optogenetics TTL Generator 4-channel
- Fluorescence Microscope Driver
- eTOSFM Microscope Body Model L
- Twist-on efocus Imaging Cannula Model L (3x)
- Protrusion Adjustment Ring Set Model L
- Pigtailed Assisted Fiber-optic & Electric Rotary Joint
- Fluorescence Microscope Holder
- Clamp for Fluorescence Microscope Holder
- Twist-on efocus Dummy Microscope
- Doric Neuroscience Studio for control and analysis
- All required electrical cables and optical patch cords^*

	5 16 _Ç	
Electrical cable jacket		
Electrical cable length (mm) — 1000 mm is standard.		
Optical fiber jacket 1100 or LWMJ [†]		
Excitation/Activation wavelength (nm) —————— 445/616		
Twist-on cannula type — D, V or E (see Twist-on efocus Imaging Cannula Model L		
GRIN diameter (μm) — 500 or 1000		

^{*}The optical fiber length is adjusted to fit the desired electrical cable length.

[†]The Ultralight Fiberglass Jacket (UFGJ) and the 1.1 mm Hytrel Jacket (1100) are lighter and more flexible, while the Lightweight Metal Jacket (LWMJ) is more robust but heavier.



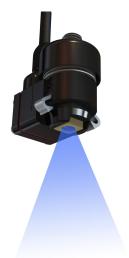
Twist-on efocus Optogenetically Synchronized Fluorescence Microscopy System for Deep-brain Imaging of GCaMP6

Fluorescence Microscope Bodies

Snap-in Fluorescence Microscope Bodies

Basic Snap-in Fluorescence Microscope Bodies

The Basic Snap-in Fluorescence Microscope Body is offered in two models: S or L. Both models have the dichroic beam-splitter, M3 optical connector, CMOS sensor etc. Each CMOS has a serial number stored within its cable that points to a specific set of mask correction filters recognizable to our software package. Model L has a 0.5 NA objective lens within its body while Model S has a plan-parallel plate instead and relies on the objective lens within the model S imaging cannula to create an image on the CMOS. When used for deep brain imaging, the fluorescence microscope body is used with an implantable imaging cannula that transfers the image from its bottom to its top.



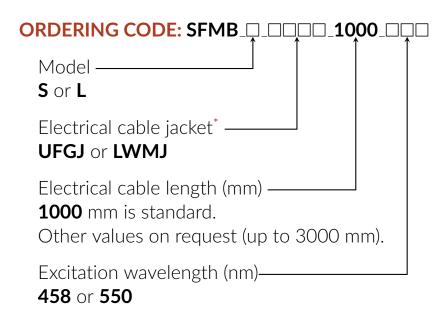
Basic Snap-in Fluorescence Microscope Body

	SPECTRUM (nm) [*]	
SFMB Bodies	Excitation	Detection
GCaMP6	458/35	525/40
RCaMP2	549/15	609/57

Table 80: Basic Snap-in Microscope Body Excitation and Detection Spectra

Table 01. Da	sia Cia ana ina Fluxaracaana	na Miaracana Dar	line Charifications
added E ba	sic Snap-in Fluorescend	.e Milcioscode Boc	Hes Specifications
101010 0 11 001			

	Basic Microscope Bodies		
SPECIFICATION	Model S	Model L	
Mass without cables (g)	2.2		
Dimensions without cables in mm (W \times L \times H)	8.8 x 13.9 x 16.6		
Frame rate (fps)	45		
Objective lens NA [†]	0.5		
FOV at image plane (pixel)	630 x 630		
FOV at object plane (μ m)	700 x 700 350 x 350		
Lens magnification	3.3x	6x	



Notes:

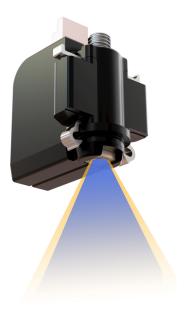
- The optical fiber length is adjusted to fit the desired electrical cable length.
- Every microscope body comes with a protective cap.

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^{*}The Ultralight Fiberglass Jacket is lighter and more flexible, the Lightweight Metal Jacket is more robust but heavier.

2-color Fluorescence Microscope Bodies

The 2-color fluorescence microscope body combines two CMOS sensors for simultaneous imaging of two different fluorophores. Due to the chromatic aberrations of GRIN lenses, the position of each image sensor is adjusted to correct the chromatic shift and image the same object plane in both colors. As the chromatic shift is related to the length of the GRIN lens, the correction is valid for one specific length of GRIN lens. For now, Green (GFP like) + Red (RFP like) systems are offered: two systems optimized for Snap-in Imaging Cannulas type D and type V, and one system optimized for surface imaging.



2-color Fluorescence Microscope Body

	SPECTRUM (nm) [*]		
2-color Bodies	Excitation	Detection	
CMOS 1	465	520/35	
CMOS 2	561	615/45	

Table 82: 2-color Microscope Bodies Excitation and Detection Spectra

Table 83: 2-color Fluorescence	Microscope	Bodies	Specifications

	2-color Microscope Bodies		
SPECIFICATION	Model S	Model L	
Dimensions without cables in mm (W \times L \times H)	18 x 17 x 9.5		
Frame rate (fps)	45		
Objective lens NA	0.5		
FOV at image plane (pixel)	600 x 600		
FOV at object plane (μ m)	730 x 730	330 x 330	
Lens magnification	Зx	6.5x	

145

ORDERING CODE: 2CFM___458/561

Model — **S**. **LD** or **LV**

Excitation wavelengths (nm)—— Excitation 1 / Excitation 2

Notes:

- The Electrical Cable for 2-color Fluorescence Microscope Bodies is required for the use of this device.
- The optical fiber length is adjusted to fit the desired electrical cable length.
- Every microscope body comes with a protective cap.

Twist-on Fluorescence Microscope Bodies

Twist-on efocus Fluorescence Microscope Body

The Twist-on efocus Fluorescence Microscope enables users to visualize larger brain areas in freely behaving animals studies. The large field of view up to 650 x 650 microns and the electronic depth adjustment of 300 microns allows calcium imaging at cellular resolution of a larger brain area. With a simple twist connector, the microscope offers an optimized way to secure the microscope body to the imaging cannula. The attachment/detachment is now easier and does not require tools.

Notes:

- The Electrical Cable for efocus Fluorescence Microscope Bodies is required for the use of this device.
- The optical fiber length is adjusted to fit the desired electrical cable length.
- Every microscope body comes with a protective cap.
- The Twist-on efocus Fluorescence Microscope Body is only compatible with the Twist-on efocus Imaging Cannulas.



Twist-on efocus Fluorescence Microscope Body

Brain Zones*	Cannula Model	Lens Diameter
0 to 2.6 μm below the skull surface	L type D	1000 mm
0 to 3.3 mm below the skull surface	L type D	500 mm
2.7 to 5.7 mm below the skull surface	L type V	500 mm
5.1 to 8.1 mm below the skull surface	L type E	500 mm

Table 84: Twist-on Microscope Body and Imaging Cannula models for specific brain zones

Table 85: Twist-on efocus Fluorescence Microscope Bodies Excitation and Detection Spectra

	SPECTRUM (nm) [†]		
eTFMB Body	Excitation	Detection	
GCaMP6	458/35	525/40	

Table 86: Twist-on efocus Fluorescence Microscope Bodies Specifications

SPECIFICATIONS	Model L
Mass without cables (g)	3.0
Frame rate (fps)	45
Cannula working distance (adjustable; µm)	0-300
Objective lens NA	0.4
FOV at image plane (pixel)	630 x 630
FOV at object plane (µm)	650 x 650
Lens magnification	3.5x

ORDERING CODE: eTFMB_L_458

Model ———

Excitation wavelengths (nm)-

^{*} Including the thickness of the skull. [†]Center wavelength/bandwidth

Twist-on efocus Optogenetically Synchronized Fluorescence Microscope Body

The Twist-on efocus Optogenetically Synchronized Fluorescence Microscope combines optogenetic stimulation/inhibition capabilities and fluorescence imaging with electronic focus adjustment within the miniature fluorescence microscope. The large field of view up to 650 x 650 microns and the electronic depth adjustment of 300 microns allows calcium imaging at cellular resolution of a larger brain area. With a simple twist connector, the microscope offers an optimized way to secure the body to the imaging cannula. The attachment/detachment is now easier and does not require tools.

ORDERING CODE: eTOSFM_L_445/616

Model _____



Twist-on efocus Optogenetically Synchronized Fluorescence Microscope Body

Notes:

- The Electrical Cable for efocus Fluorescence Microscope Bodies is required for the use of this device.
- The optical fiber length is adjusted to fit the desired electrical cable length.
- Every microscope body comes with a protective cap.
- The Twist-on efocus Optogenetically Synchronized Fluorescence Microscope Body is only compatible with the Twist-on efocus Imaging Cannulas.

Table 87: eTOSFM Microscope Body Excitation and Detection Spectra

	SPECTRUM (nm) [*]		
eTOSFM Body	Opsin activation	Fluorophore Excitation	Detection
GCaMP6 + NpHR3.0	616/76 (0-55 mW/mm ²)	445/62	525/40

*Center wavelength/bandwidth

SPECIFICATIONS	Model L
Mass without cables (g)	3.0
Frame rate (fps)	45
Cannula working distance (adjustable)	0-300 µm
Objective lens NA	0.4
FOV at image plane (pixel)	630 x 630
FOV at object plane (µm)	650 x 650

Table 88: Twist-on efocus Optogenetically Synchronized Fluorescence Microscope Body Specifications

Table 89: Twist-on Microscope	Bodv and	Imaging Cannula	models for specific brai	n zones

Brain Zones [*]	Cannula Model	Lens Diameter
0 to 2.6 μm below the skull surface	L type D	1000 mm
0 to 3.3 mm below the skull surface	L type D	500 mm
2.7 to 5.7 mm below the skull surface	L type V	500 mm
5.1 to 8.1 mm below the skull surface	L type E	500 mm

¹⁴⁸

^{*} Including the thickness of the skull.

Imaging Cannulas

Ordinary fiber-optic cannulas send light along the optical fiber but do not create or capture an image. The imaging cannula can transfer an image but only over a very short distance in highly turbid media like brain tissue. For areas near the brain surface use the *Imaging Cannula Model S*. For deeper brain regions use the *Imaging Cannula Model L* with image guiding gradient-index rod lens that brings the image from inside the brain to the skull surface. Each *Snap-in Imaging Cannula* comes with a protective cap and it is a good practice to put it on the implanted cannula when the microscope body is not in place.

Snap-in Imaging Cannula Model S

The Snap-in Imaging Cannula Model S looks inside brain tissue with an objective lens that brings the image from inside the first 150 μ m of the brain to the microscope camera.

Note: One Protrusion Adjustment Ring Model S is included with each Snap-in Imaging Cannula Model S. If more rings are needed, a set can be purchased separately (**PARS_S**).

ORDERING CODE: SICS_2_2,4

Lens Magnification (x) \Box

Lens Working distance – in air (mm)

Snap-in Imaging Cannula Model L

Standard Imaging Cannula Model L

As the choice of these lenses is quite limited, different depth ranges of brain tissue are accessed with different lens lengths while fine focusing is done with the protrusion adjustment ring that comes with each cannula. As cannulas might be re-used it is advisable to get a set of these rings as spare parts. The working distance of D, V and E imaging cannulas is 80 μ m.

<u>Note</u>: Each Standard Imaging Cannula Model L is provided with one of each of the five models of Protrusion Adjustment Rings Model L. If more rings are needed, a set can be purchased separately (**PARS_L**). Snap-in Imaging Cannula Model S and Protrusion Adjustment Ring



Snap-in Imaging Cannula Model L and Protrusion Adjustment Ring



ORDERING CODE: SICL___500_80

Model ______ **D**, **V**, **E** (see Table 90) Lens diameter (μm) _____ Working distance in water (μm) _____

Table 90: Snap-in Imaging Cannula Model L Specifications

Cannula Type	Range of Penetration Depth (mm)
D	0 - 3.46
\vee	2.87 - 5.96
E	5.30 - 8.39

efocus Imaging Cannula Model L

For deep brain regions (0 mm to 8 mm deep) use the efocus Imaging Cannula Model L that brings the image from inside the brain to the skull surface with an image guiding gradient-index rod lens. efocus Imaging Cannulas are compatible with efocus Fluorescence Microscope Bodies only.



efocus Imaging Cannula Model L and Protrusion Adjustment

<u>Note</u>: Each efocus Imaging Cannula Model L is provided with one of each of the five models of Protrusion Adjustment Rings Model L. If more rings are needed, a set can be purchased separately (**PARS_L**).

ORDERING CODE: eSICL___500_80

Model ———

D, V, E (see Table 90)

Lens diameter (μ m) –

Working distance in water (µm) —

Reduced Footprint Imaging Cannula Model L

To improve the stability of the cannula on the animal, the base of the standard snap-in imaging cannula has been defined with a larger diameter. In opposition, the *Reduced Footprint Imaging Cannula* has been designed with an outer diameter as small as 3.5 mm to image exiguous area of the brain (e.g. olfactory bulb). In the *Reduced Footprint Imaging Cannula*, the GRIN lens protrusion length is user defined with steps of 250 microns. There is no protrusion adjustment ring.

ORDERING CODE: RFICL _____80

Model ______ **D** for deep, **V** for very deep or **E** for extra deep

GRIN lens protrusion length ——— **0.25 mm increments** (see Table 91)

Working distance in water (µm) –

Cannula Type	GRIN lens protrusion length (0.25 mm increments)
D	0 - 3.00
\vee	2.43 - 5.43
E	4.85 - 7.85

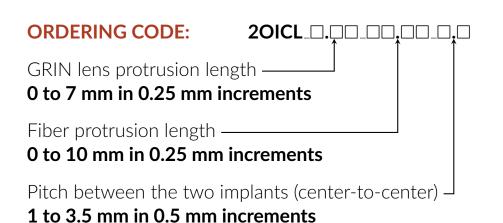
<u>Note</u>: The protrusion length of the GRIN lens is determined from the base of the imaging cannula to its tip.



Reduced Footprint Imaging Cannula

2-channel Optogenetics and Imaging Cannula Model L

The 2-channel Optogenetics and Imaging Cannula has two parallel implants, a GRIN lens for deep-brain imaging, and a 200 μ m diameter optical fiber for optogenetic stimulation of another brain area. For now, this imaging cannula is used for experiments not requiring the use of a rotary joint. The development of a compatible rotary joint is in progress.



2-channel Optogenetics and Imaging Cannula model L

<u>Note</u>: The protrusion length of the GRIN lens and the fiber is determined from the base of the imaging cannula to their tips.

Imaging Cannula Model L with Prism

While the standard Imaging Cannula Model L images horizontal plane sections of the brain (0-8 mm depth), the Imaging Cannula Model L with Prism allows the imaging of sagital-coronal plane sections. The GRIN lens and the right-angle prism at its tip bring images to the skull surface. This imaging configuration has the advantage of leaving the brain tissue intact above the region of interest. Four orientations of the prism are available depending on the brain region of interest.



Imaging Cannula Model L with Prism

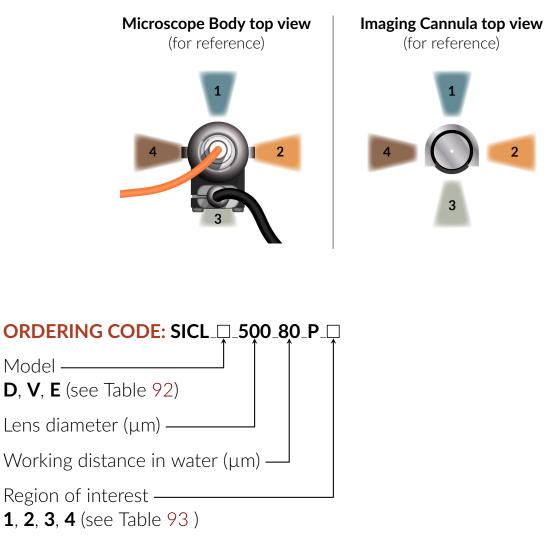
Notes:

Cannula Type	Range of Penetration Depth (mm)
D	0 - 3.5
\vee	2.9 - 5.9
E	5.4 - 8.3

Table 92: Imaging Cannula Model L with Prism Specifications

- The range of penetration depth is determined from the surface of the skull or the bottom of the focusing ring to the lower tip of the prism.
- Each Imaging Cannula Model L with Prism is provided with one of each of the five models of Protrusion Adjustment Rings Model L. If more rings are needed, a set can be purchased separately (**PARS_L**).

Table 93: Regions of interest observed with the Imaging Cannula Model L with Prism



Twist-on Imaging Cannula Model L

Twist-on efocus Imaging Cannula Model L

For deep brain regions (0 mm to 8 mm deep) use the Twist-on efocus Imaging Cannula Model L that brings the image from inside the brain to the skull surface with an image guiding gradient-index rod lens. The Twist-on efocus



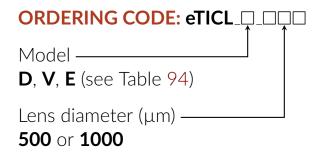
Twist-on efocus Imaging Cannula Model

Imaging Cannulas are compatible with Twist-on *L and Protrusion Adjustment Ring* efocus Fluorescence Microscope Bodies only. For a larger field of view, it is possible to use a GRIN diameter of 1000 μm instead of 500 μm.

<u>Note</u>: Each Twist-on Imaging Cannula Model L is provided with one of each of the five models of Protrusion Adjustment Rings Model L. If more rings are needed, a set can be purchased separately (**PARS_L**).

GRIN diameter (µm)	Cannula Type	Range of Penetration Depth (mm)
500	D V E	0 - 3.3 2.7 - 5.7 5.1 - 8.1
1000	D	0 - 2.6

Table 94: Twist-on Imaging Cannula Model L Specifications



Twist-on Imaging Cannula Model L with Prism

While the standard Imaging Cannula Model L images horizontal plane sections of the brain (0-8 mm depth), the Twist-on Imaging Cannula Model L with Prism allows the imaging of sagital-coronal plane sections. The GRIN lens and the right-angle prism at its tip bring images to the skull surface. This imaging configuration has the advantage of leaving the brain tissue above the region of interest intact. Four prism orientations are available depending on the brain region of interest.



Twist-on Imaging Cannula Model L with Prism

GRIN diameter (µm)	Cannula Type	Range of Penetration Depth (mm)
500	D V E	0 - 3.3 2.7 - 5.7 5.2 - 8.2
1000	D	0 - 2.7

Table 95: Twist-on Imaging Cannula Model L with Prism Specifications

Notes:

- The penetration depth range is determined from the surface of the skull or the bottom of the focusing ring to the lower tip of the prism.
- Each Twist-on Imaging Cannula Model L with Prism is provided with one of each of the five models of Protrusion Adjustment Rings Model L. If more rings are needed, a set can be purchased separately (**PARS_L**).

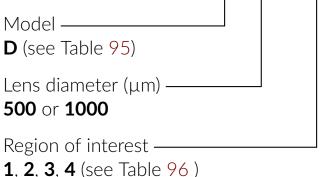
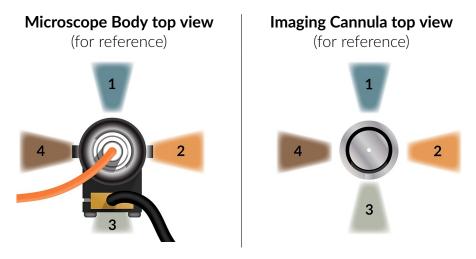


Table 96: Regions of interest observed with the Twist-on Imaging Cannula Model L with Prism



Fluorescence Microscope Drivers

Fluorescence Microscope Driver

This driver has been designed for the Basic Snap-in and the Optogenetically Synchronized Fluorescence Microscope Bodies. It allows for computer control over the excitation LED light source, image capturing and its broadcast at video rate to single or multiple computers via high speed Ethernet communication. It can be triggered or synchronized with external recording devices and it can trigger other devices.



Fluorescence Microscope Driver

ORDERING CODE: FMD

2-color Fluorescence Microscope Driver

The 2-color Fluorescence Microscope Driver allows for computer control over excitation light sources (blue LED and yellow Ce:YAG source), images capturing from both CMOS and the broadcast at video rate to single or multiple computers via high speed Ethernet communication. It is compatible with the 2-color microscope body.



2-color Fluorescence Microscope Driver

ORDERING CODE: FMD_2

Fluorescence Microscope Accessories

Electrical Cable for Fluorescence Microscope Bodies

Electrical Cable for 2-color Fluorescence Microscope Bodies

2-color Fluorescence Microscope Bodies are connectorized in order to allow for more flexibility. Like Basic and OSFM Fluorescence Microscope Bodies, the jacket of the electrical cable can be customized with a lighter and more flexible Ultralight Fiberglass Jacket (UFGJ) or a



Electrical cable for 2-color Fluorescence Microscope Bodies

more robust but heavier Lightweight Metal Jacket (LWMJ).

ORDERING CODE: EC 2 000 1000

Electrical cable jacket -**UFGJ** or **LWMJ**

Electrical cable length (mm) -1000 mm is standard.

Other values on request (up to 3000 mm).

Note: The 2-color Electrical Cable is required for the use of the 2-color Fluorescence Microscope Bodies.

Electrical Cable for efocus Twist-on Fluorescence Microscope Bodies

efocus Fluorescence Microscope Bodies are connectorized in order to allow for more flexibility. The jacket of the electrical cable can be customized with a lighter and more flexible Ultralight Fiberglass Jacket (UFGJ, recommended for mice) or a more robust but heavier Lightweight Metal Jacket (LWMJ, recommended for rats).



Electrical cable for efocus Fluorescence Microscope Bodies

ORDERING CODE: EC_efocus_00_1000

Electrical cable jacket ______ UFGJ or LWMJ Electrical cable length (mm) _____ 1000 mm is standard.

Other values on request (up to 3000 mm).

Note: The efocus Electrical Cable is required for the use of the efocus Twist-on Fluorescence Microscope Bodies.

Fluorescence Microscope Holders

Fluorescence Microscope Holder

For *in vitro* and head-fixed observations, it is desirable to have a *Fluorescence Microscope Holder* coaxial with microscope that fits stereotaxic instrumentation or micromanipulators. As our optical connector has axial position, we constructed the holder to be an optical interface as well. The microscope holder allows for imaging while implanting the cannula from a stereotaxic frame.



Fluorescence Microscope Holder

<u>Note</u>: The Fluorescence Microscope Holder is only compatible with Single-color Fluorescence Microscope Bodies.

ORDERING CODE: FMH

Fluorescence Microscope Holder 2

A specific pigtailed microscope holder has been designed to fit the 2-color fluorescence microscope bodies. This holder can be used to image the brain during the implantation of the cannula or during experiments requiring a head-fixed in vivo configuration. The microscope holder is compatible with



Fluorescence Microscope Holder 2

stereotaxic instruments and connects to the microscope body via a CM3 connector. The input patch cord has a standard length of 1.0 m.

<u>Note</u>: The Fluorescence Microscope Holder 2 is compatible with all our Fluorescence Microscope Bodies.

ORDERING CODE: FMH_2

Clamp for Fluorescence Microscope Holder

This adaptor allows for an easy fit between our Fluorescence Microscope Holders and most stereotaxic frames.

ORDERING CODE: CLAMP_FMH



Clamp for Fluorescence Microscope Holder

Dummy Microscope

Model I

Dummy Microscopes

Dummy Microscope

The dummy microscope is a look-alike, inexpensive replica of the Snap-in Fluorescence Microscope Body that fits any snap-in imaging cannula. It is meant to be secured to the rodent's head to habituate it to the feel and weight of a microscope before using the real microscope body. The dummy microscope has an M3 connector and can be connected to a CM3 optical patch cord.

Table 97: Dummy Microscope Ordering Code

Microscope Body Model	Ordering Code
Basic and OSFM model S	DSMB-S
Basic and OSFM model L	DSMB-L

2-color Dummy Microscope

The 2-color dummy microscope has the same shape and weight as the 2-color fluorescence microscope body model S and L. This dummy microscope is compatible with all cannula types.

Model ·		
S or L		



2-color Dummy Microscope Model L

Twist-on efocus Dummy Microscope

The Twist-on efocus Dummy Microscope is a look-alike, inexpensive replica of the Twist-on efocus Fluorescence Microscope Body that fits any Twist-on efocus Imaging Cannula. It is meant to be secured to the rodent's head to habituate it to the feel and weight of a microscope before using the real microscope body. The Twist-on efocus Dummy Microscope has an M3 connector and can be connected to a CM3 optical patch cord.

ORDERING CODE: eDTMB-L

Fluorescence Microscope Snapping Tools

Fluorescence Microscope Snapping Tool

This tool is used to easily detach the Basic Snap-in and the OSFM microscope body from the imaging cannula.

ORDERING CODE: FMST



Twist-on efocus Dummy Microscope Model L



Fluorescence Microscope Snapping Tool

2-color Fluorescence Microscope Snapping Tool

The 2-color Fluorescence Microscope Snapping Tool comprises two different pairs of tweezers, one for attaching and one for detaching the 2-color fluorescence microscope body from the imaging cannula.



Protrusion Adjustment Ring Set

As the point of observation can be anywhere within the brain, a set of protrusion adjustment rings of different heights is available. By combining an imaging cannula model L with the right protrusion adjustment ring it is possible to cover most parts of the brain. The height of the rings is 2.0 mm, 2.7 mm,

3.4 mm, 4.2 mm and 4.9 mm. Within the set, there are 8 rings for each height. For the model S, one protrusion adjustment ring (height of 4.5 mm) is required to observe the brain from the surface to 1.1 mm deep. The set is composed of 10 identical rings.

Table	98:	Protrusion	Adiustment	Ring S	et Ordering (Codes
TUDIC	/0.	11001031011	, ajastinent	10100		200005

Microscope Body Model	Ordering Code
S	PARS_S
L	PARS_L

External Relay Lens Accessory

Our snap-in imaging cannulas create an image at the bottom part of its mechanical body that is perfect for our miniaturized microscopes. However, for conventional and specialized microscopes like the two photon microscope, the image is simply not accessible. To correct this situation we have made a relay lens accessory that fits the interior of



External Relay Lens Accessorv



2-color Fluorescence Microscope Snapping Tool





Protrusion Adjustment Ring Set for

Microscope Body Model L

the cannula and brings the image to the top of the cannula where access to the image is not obstructed. The optics used in our relay lens accessory is a gradient index (GRIN) lens having the following specifications:

SPECIFICATION	VALUE	
Diameter	1 mm	
Numerical aperture	0.5	
Magnification	1:1	
Design wavelength	520 nm	

Note: the object and image working distances can be adjusted by changing the distance between the relay lens and the microscope objective.

ORDERING CODE: ERLA_1

Lens magnification (x) —

Fiber Photometry

In neuroscience, fiber photometry denotes a method whereby a single or multimode chronically implanted optical fiber delivers excitation light to neurons labeled with a fluorescent reporters typically calcium indicator(s) such as GCaMP and collects their overall activity-induced fluorescence. Within the field of view, while the fluorescence microscopy indicates activity of each tagged neuron, the fiber photometry sums up the activity-induced fluorescence of all neurons expressing the indicator(s).

Distinguishing the very weak fluorescence variations from relatively high noise levels requires careful selection of components within the system, from light sources to detectors. The connectorized LED module (CLED) as excitation source offers stable power,speckle-free illumination and sufficient spectral intensity for most fluorescent markers. Another interesting alternative is a combination of UV or blue LEDs with the Ce:YAG source filtered to a required wavelength. The latter offers all the advantages of LED illumination, along with higher intensity in the 500-600 nm range. Laser sources could be considered when using small diameter core fibers with low NA and/or multiple color excitations requiring narrow spectral filtering.

The heart of the Fiber Photometry System is the Fluorescence Mini Cube (FMC) that directs excitation light into an optical fiber leading to the fiber-optic cannula. The fluorescence of the sample captured by the cannula is returned into the FMC, filtered and redirected into a detection fiber that goes to the high sensitivity photodetector. The opto-mechanical design of the fluorescence mini-cube, the filter selection and the coupling optics alignment play an important role in increasing the signal to noise ratio.

Typically, excitation optical power in mW range produces fluorescence responses in nW range. The detection of such a low level signal requires a low-noise amplified photodetector. As the optical isolation of each component is essential in this power range, the optical fibers must have protective jackets to avoid possible effects of ambient light on the measurement. The worst DC noise might be coming from the autofluorescence of the probe or the patch cord itself. To keep this noise in check, low autofluorescence optical fibers must be used and their length kept to a bare minimum. To prevent injecting light into optical fiber cladding, the fiber optic collimator must under fill the fiber NA and the light spot on the fiber's end face should be smaller than its core diameter.

Fiber Photometry Systems

1-site Fiber Photometry Systems

GCaMP Isosbestic and Functional Excitations - 405 and 465 nm

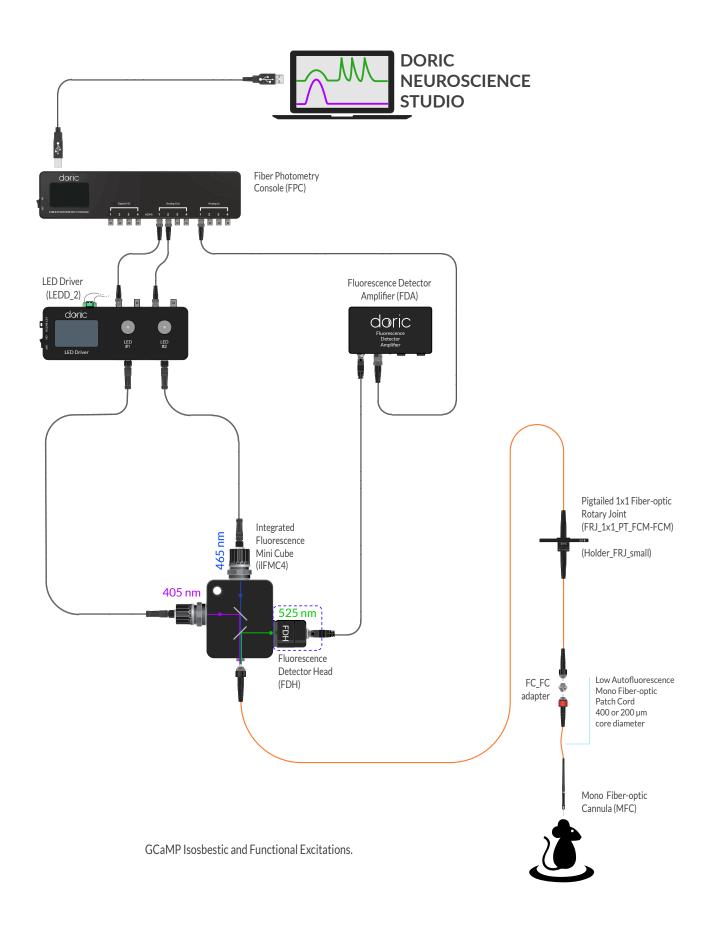
This 1-site Fiber Photometry System measures the 405 nm (isosbestic point) excited GCaMP fluorescence, and the 465 nm excited calcium-dependent GCaMP fluorescence, on a single photodetector. The fluorescence emission can be demodulated by lock-in detection, or by sequential acquisition. The GCaMP Isosbestic and Functional Excitations - 405 and 465 nm System contains:

- 2-channel LED Driver
- Integrated Fluorescence Mini Cube with 4 ports Lock-in or Sequential Detection of GCaMP Isosbestic and Functional Excitations
- Pigtailed 1x1 Fiber-optic Rotary Joint
- 1x1 Fiber-optic Rotary Joint Holder
- Rotary Joint Gimbal Holder
- Mono Fiber-optic Cannulas (10x)
- Fiber Photometry Cannula Holder
- Fiber Photometry Console for data acquisition and illumination control
- Doric Neuroscience Studio Software
- Fiber Photometry Rack to mount the whole system
- All required electrical cables and optical patch cords

ORDERING CODE: FPS_1S_405/GFP_00.57

Cannula fiber diameter (μm) — 400 or 200
Cannula numerical aperture

<u>Note</u>: Other light sources and different fluorophore combinations are possible. Please do not hesitate to request your preferences.



GCaMP Isosbestic and Functional Excitations, and Red Fluorophore Fluorescence

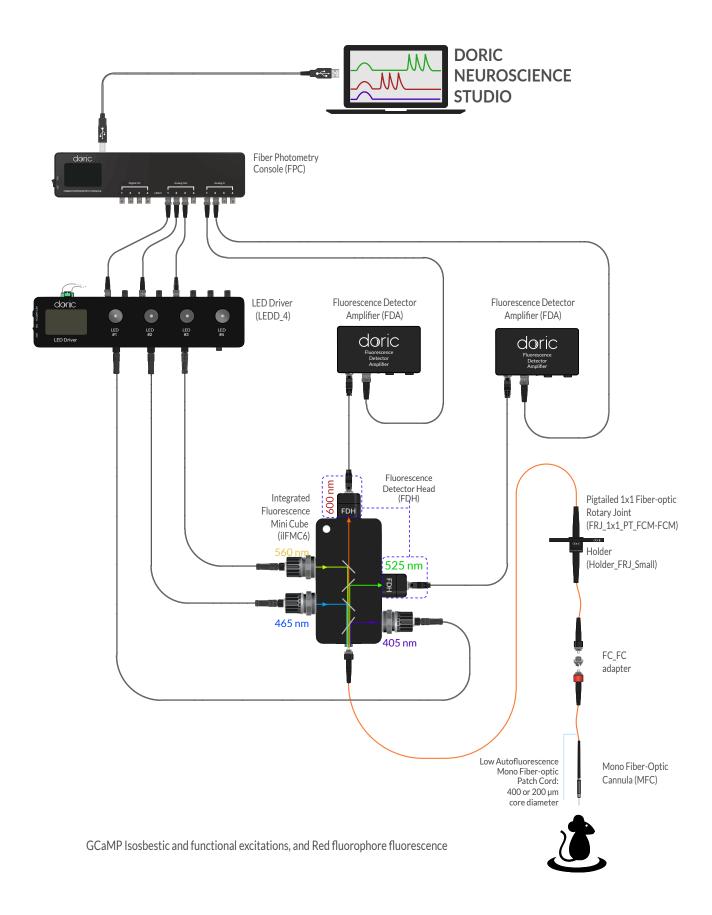
This 1-site Fiber Photometry System contains all the items necessary to do photometry measurements of two independent colors for instance GFP-like and RFP-like fluorophores in freely-moving animals. It also measures the 405 nm (isosbestic point) excitation of GCaMP fluorescence. The GCaMP fluorescence emission can be demodulated by lock-in detection or by sequential acquisition. It is also possible to use lock-in demodulation to ensure a proper spectral separation of the green and red fluorophore emissions. The GCaMP Isosbestic and Functional Excitations, and Red Fluorophore Fluorescence System contains:

- 4-channel LED Driver
- Integrated Fluorescence Mini Cube with 6 ports Two Fluorophores Fluorescence and GCaMP Isosbestic Excitation
- Pigtailed 1x1 Fiber-optic Rotary Joint
- 1x1 Fiber-optic Rotary Joint Holder
- Rotary Joint Gimbal Holder
- Mono Fiber-optic Cannulas (10x)
- Fiber Photometry Cannula Holder
- Fiber Photometry Console for data acquisition and illumination control
- Doric Neuroscience Studio Software
- Fiber Photometry Rack to mount the whole system
- All required electrical cables and optical patch cords

Note: Other light sources and different fluorophore combinations are possible. Please do not hesitate to request your preferences.

ORDERING CODE: FPS_1S_405/GFP/RFP_00.57

```
Cannula fiber diameter (μm) ______Ť
400 or 200
Cannula numerical aperture _____
```



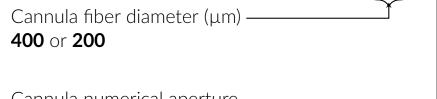
GCaMP Isosbestic and Functional Excitations and Opsin Activation

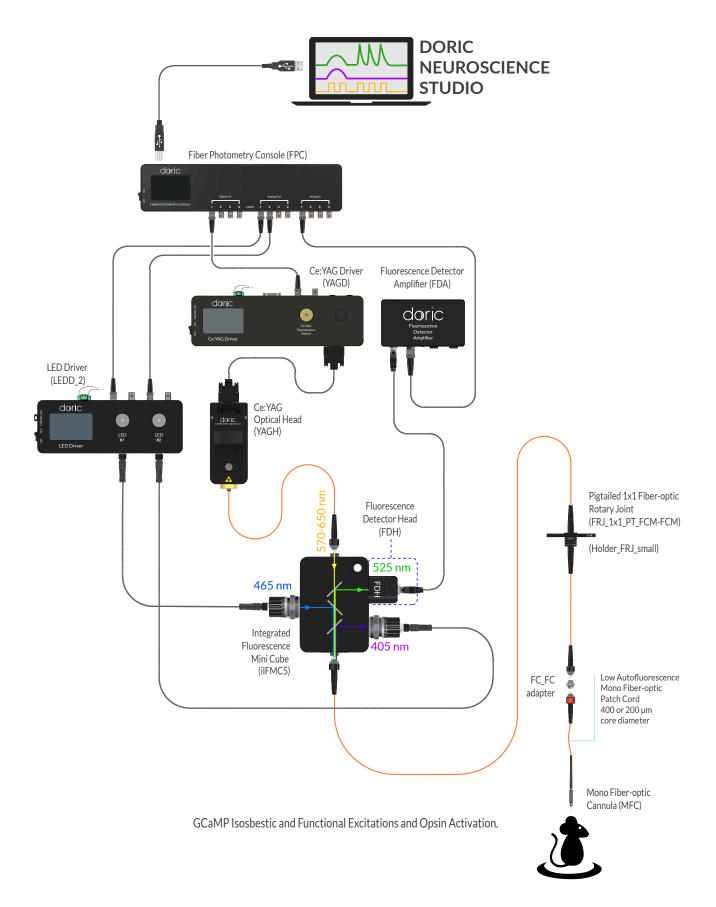
This 1-site Fiber Photometry System measures the GCaMP isosbestic point and the GCaMP functional calcium-dependent fluorescence, on a single photodetector. The fluorescence emission can be demodulated by lock-in detection, or by sequential acquisition. In addition, this system allows the activation of an opsin (570-650 nm). The GCaMP Isosbestic and Functional Excitations, and Opsin Activation System contains:

- 2-channel LED Driver
- Ce:YAG Optical Head
- Ce:YAG Driver
- Bandpass Filter For Ce:YAG Fiber Light Sources
- Integrated Fluorescence Mini Cube with 4 ports Excitation, Fluorescence and Opsin Activation
- Pigtailed 1x1 Fiber-optic Rotary Joint
- 1x1 Fiber-optic Rotary Joint Holder
- Rotary Joint Gimbal Holder
- Mono Fiber-optic Cannulas (10x)
- Fiber Photometry Cannula Holder
- Fiber Photometry Console for data acquisition and illumination control
- Doric Neuroscience Studio Software
- All required electrical cables and optical patch cords

Note: Other light sources, such as laser diodes, and different fluorophore combinations are possible. Please do not hesitate to request your preferences.

ORDERING CODE: FPS_1S_405/GFP/opsin_00.57





2-site Fiber Photometry Systems

2-site GCaMP Isosbestic and Functional Excitations

This 2-site Fiber Photometry System measures the 405 nm (isosbestic point) excited GCaMP fluorescence, and the 465 nm excited calcium-dependent GCaMP fluorescence, on a single photodetector. The fluorescence emission can be demodulated by lock-in detection, or by sequential acquisition. The measures are done in two different brains areas using distinct optical paths and two detectors.

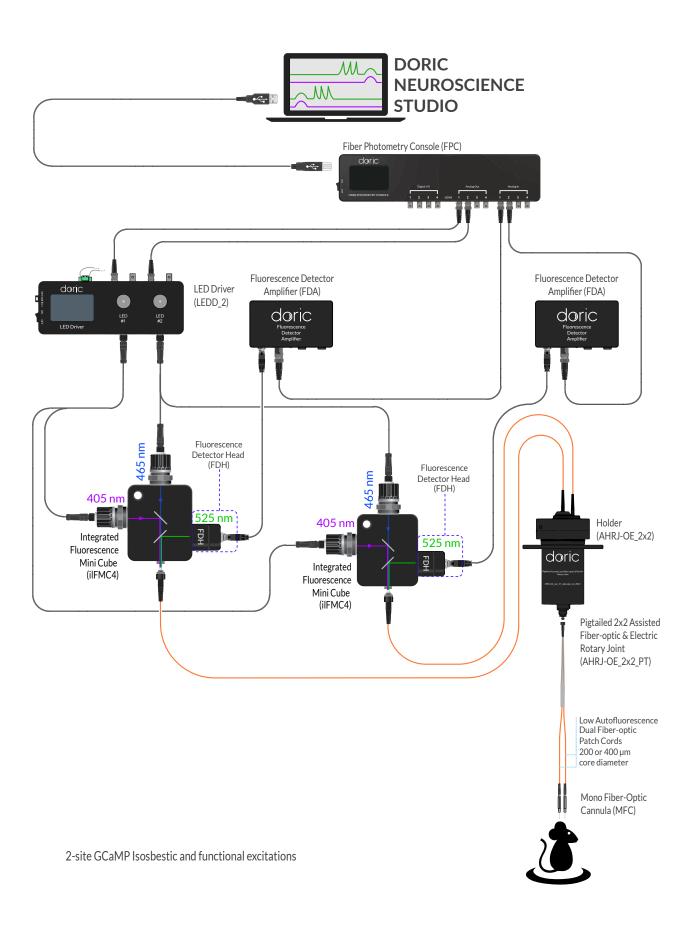
The 2-site GCaMP Isosbestic and Functional Excitations contains:

- 2-channel LED Driver
- Integrated Fluorescence Mini Cube with 4 ports Lock-in or Sequential Detection for GCaMP Isosbestic and Functional Excitations (filter set optimized for 405 nm excitation and GFP; (2x)
- Pigtailed 2x2 Assisted Fiber-optic & Electric Rotary Joint
- 2x2 Fiber-optic Rotary Joint Holder
- Mono or Dual Fiber-optic Cannulas (10x)
- Fiber Photometry Cannula Holder
- Fiber Photometry Console for data acquisition and illumination control
- Doric Neuroscience Studio Software
- Fiber Photometry Rack to mount the whole system
- All required electrical cables and optical patch cords

ORDERING CODE: FPS_2S_405/GFP_00-0.57

Cannula fiber diameter (μm) —— **400** or **200**

Cannula numerical aperture —



Bundle-imaging Fiber Photometry Systems

Scaling up fiber photometry measurement with photodiode based systems to a large number of samples can be difficult. This is due to the increasing number of components required, such as mini cubes, cables and sensors. With some compromises on time resolution and sensitivity, an alternative approach is to relay the image of an optical fiber bundle to an image sensor. The opposite end of the bundle can be split, with each connected to different samples. On the camera, the signal of each sample can be distinguished by measuring the average count coming from each individual fiber in the bundle.

GCaMP Isosbestic & Functional Excitations

The GCaMP Isosbestic & Functional Excitations Bundle-imaging Fiber Photometry System contains all items necessary to perform fiber photometry measurements on a large number of animals and sites with GFP-like fluorophores. It also measures the 405 nm (isosbestic point) excitation of GCaMP fluorescence. The GCaMP fluorescence emission can be demodulated by sequential acquisition. The base GCaMP Isosbestic & Functional Excitations Bundle-imaging Fiber Photometry

System contains the following elements.

- 2-channel LED driver
- Bundle-imaging Fluorescence Mini Cube with 4 ports Isosbestic and Functional Excitations
- Bundle-imaging Fiber Photometry Driver
- 4-port USB3.0 Hub
- Photometry Rack for BFPS
- All required electrical cables

<u>Note</u>: Cannulae, rotary joints and other elements used are to be defined for each application, and are not specified here.

ORDERING CODE: BFPS_405/GFP

GCaMP Isosbestic & Functional Excitations, & Red Fluorophore Fluorescence

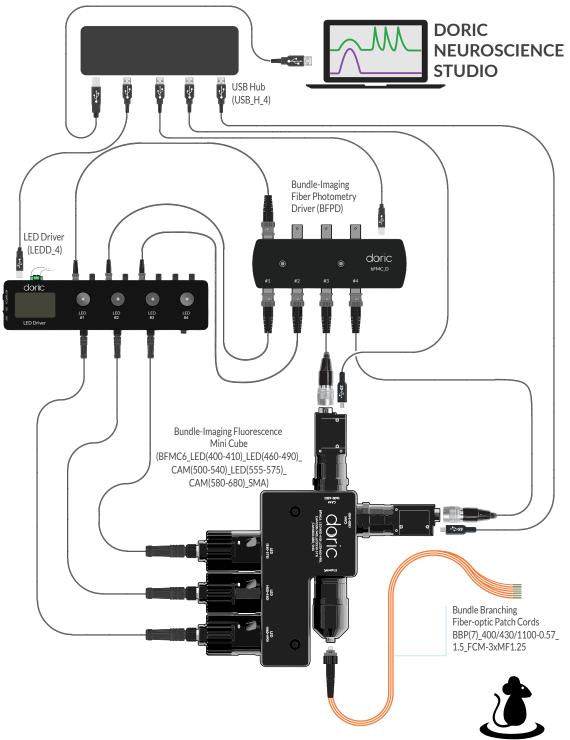
The GCaMP Isosbestic & Functional Excitations, & Red Fluorophore Fluorescence Bundleimaging Fiber Photometry System contains all items necessary to perform fiber photometry measurements on a large number of animals and sites with GFP-like and RFP-like fluorophores. It also measures the 405 nm (isosbestic point) excitation of GCaMP fluorescence. The GCaMP fluorescence emission can be demodulated by sequential acquisition.

The base GCaMP Isosbestic & Functional Excitations, & Red Fluorophore Fluorescence Bundle-imaging Fiber Photometry System contains the following elements.

- 4-channel LED driver
- Bundle-imaging Fluorescence Mini Cube with 6 ports GCaMP Isosbestic and Functional Excitations, & Red Fluorophore Fluorescence
- Bundle-imaging Fiber Photometry Driver
- 4-port USB3.0 Hub
- All required electrical cables
- Photometry Rack for BFPS

<u>Note</u>: Cannulas, rotary joints and other elements used are to be defined for each application, and are not specified here.

ORDERING CODE: BFPS_405/GFP/RFP



GCaMP Isosbestic & Functional Excitations, & Red Fluorophore Photometry System

Fiber Photometry Console



Fiber Photometry Console

This FPGA based data acquisition unit synchronizes the control of excitation light and the detection of the induced fluorescence. This device seamlessly integrates with the Doric Neuroscience Studio that provides user interface for multi-channel photometry experiments. The software interface enables control over the CW excitation light pulses, or the sinusoidal waveform trig of an external source (i.e. LED driver) with 4 TTL and 4 analog voltage outputs. The software interface displays real-time recording data of up to 4 detector input signals. Signal processing such as averaging, subtraction, multiplication to calculate the Δ F/F₀ and other new functionalities are being developed. Updates will be freely available as they are released. Main features:

- 4 Digital Input/Output TTL, 25 MS/s, via 4 BNC connector; In : 3 k Ω , Out : 30 Ω
- 4 Analog Input ± 10 V, 17 bits, 15 kS/s, via 4 BNC connector; 124 k Ω
- 4 Analog Output \pm 5V, 16 bits, 25 MS/s, via 4 BNC connector; 6 Ω
- 1 digital communication SPI and LVDS via custom pinout HDMI connector
- USB2 connection to computer, cable included
- Compatible with Doric Neuroscience Studio with photometry-oriented interface
- All software updates included

ORDERING CODE: FPC

Connectorized Fluorescence Mini Cubes

The fiber photometry experiments may require a varied number of excitation and detection channels, an optional optogenetically synchronized activation/silencing channel etc., directly affecting the number of fluorescence cube ports. The sample itself requires fixed or rotating port. As there are number of different possibilities of assigning these ports, our cube classification is based on a number of ports. So far, we offer fluorescence mini cube models with 3, 4, 5, 6 and 7 ports where each port is assigned one of the following functions: E for tagged neurons excitation band or IE for GCaMP isosbestic excitation band, F for fluorescence band, O for optogenetics activation or silencing and S for the sample. For extremely low light level applications, the fluorescence port code letters F, F1, F2, etc., can be replaced by PMT, meaning that the fiber-optic receptacle is replaced by a photomultiplier tube attached directly to the mini cube.

Fluorescence Mini Cube with 3 ports

Single excitation band fiber photometry measurements use a *Fluorescence Mini Cube with 3 ports*, with one port for the excitation light, one for the fluorescence detection and one for the sample being tested. The cube has a dichroic mirror to separate the excitation light from the fluorescence emission and may incorporate narrow bandpass filters that limit the excitation or fluorescence spectrum. Currently we offer configurations for GFPlike or RFP-like fluorophores. The 3 ports' mini cube filters can be customized on request.

On the image E is for excitation, F for fluorescence and S is for the fixed sample port.



Fluorescence Mini Cube 3 Table 100: Fluorescence Mini Cube 3 ports Ordering Codes ports

Filter Set	Excitation Band (nm)	Detection Band (nm)	Ordering Code
GFP-like	460-490	500-550	FMC3_E(460-490)_F(500-550)_S [*]
RFP-like	540-570	580-680	FMC3_E(540-570)_F(580-680)_S [*]

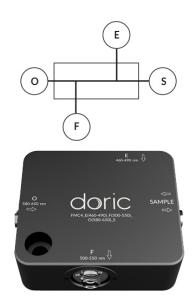
^{*}To use with a PMT, in the ordering code replace **F** for **PMT**, *e.g.* FMC3_E(460-490)_PMT(500-550)_S

Fluorescence Mini Cube with 4 ports

• Excitation, Fluorescence and Opsin Activation

This cube is for measurements involving an excitation, an optogenetic activation/silencing, fluorescence detection and sample ports. Such a cube can be used for GCaMP fluorescence measurements combined with the activation of red opsins in the 580-650 nm band.

On the image, E is for excitation, F for fluorescence, O for opsin activation/silencing and S is for the fixed sample port. The numbers in the brackets of the ordering code are for the corresponding wavelength bands.



FMC4, Excitation, Fluorescence and Opsin Activation

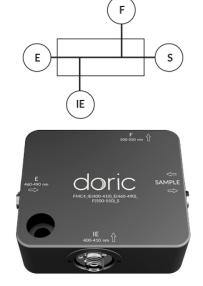
ORDERING CODE:

FMC4_E(460-490)_F(500-550)_O(580-650)_S

• Lock-in or Sequential Detection for GCaMP Isosbestic and Functional Excitations

This cube permits excitation of the GCaMP isosbestic point with 400-410 nm light and Ca²⁺dependent functional excitation with 460-490 nm light. The single detector measures both signals within the fluorescence detection window of 500-540 nm. The separation of GCaMP isosbestic and functional excitations is possible if both excitations are modulated.

On the image IE and E are ports for excitations, F is for fluorescence detection and S is for fixed sample port. The numbers in the brackets of the ordering code denote the corresponding wavelength bands.



FMC4, Lock-in or Sequential Detection for GCaMP Isosbestic and Functional Excitations

ORDERING CODE: FMC4_IE(400-410)_E(460-490)_F(500-550)_S^{*}

^{*}To use with a PMT, in the ordering code replace **F** for **PMT**, *e.g.* FMC4_E(460-490)_PMT(500-550)_O(580-650)_S

• FRET Cube (One Excitation and Two Fluorescence Detection Ports)

This cube is used to excite the donor fluorophore with a 420-445 nm excitation wavelength band. The donor fluorophore loses part of that energy to fluorescence in the 460-500 nm band, while the rest is transferred in a distance dependent radiationless manner to the acceptor fluorophore. The fluorescence emitted by the acceptor is detected in the 528-556 nm window.

ORDERING CODE: FMC4_E(420-445)_F1(460-500)_F2(528-556)_S^{*}



On the image E is for excitation, F1 and F2 for two spectrally different fluorescences and S is for the fixed sam-

FMC4, FRET Cube

ple port. The numbers in the brackets are for the corresponding wavelength bands.

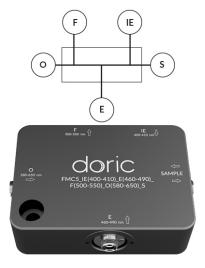
Fluorescence Mini Cube with 5 ports

• GCaMP Isosbestic and Functional Excitations, and Opsin Activation

The GCaMP isosbestic point is excited by 400-410 nm bandwidth light and the 460-490 nm spectral window excites its functional fluorescence. The 500-550 nm spectral window collects the functional fluorescence of GCaMP. At the same time, an opsin is excited with 580-650 nm light.

ORDERING CODE:

FMC5_IE(400-410)_E(460-490)_F(500-550)_O(580-650)_S*



FMC5, GCaMP Isosbestic and Functional Excitations, and Opsin Activation

^{*}To use with a PMT, in the ordering code replace **F** for **PMT**, *e.g.* FMC5_IE(400-410)_E(460-490)_PMT(500-550)_O(580-650)_S

Separated Two Fluorophores Fluorescence

This cube is used for green and red fluorophore excitation and their respective detection wavelengths. Other fluorophore combinations are possible.

ORDERING CODE:

FMC5_E1(460-490)_F1(500-540)_E2(555-570)_F2(580-680)_S*

Fluorescence Mini Cube with 6 ports

• Two Fluorophores Fluorescence and GCaMP Isosbestic Excitation

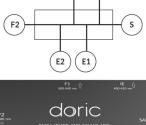
This cube can be used to detect the fluorescence from two calcium indicators and the GCaMP isosbestic point. The separation of functional and isosbestic fluorescence signals of GCaMP is possible if the light sources are modulated.

ORDERING CODE:

FMC6_IE(400-410)_E1(460-490)_F1(500-540)_E2(555-570)_F2(580-680)_-S^{*}

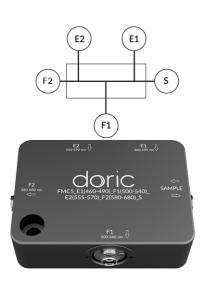
^{*}To use with a PMT, in the ordering code replace **F** for **PMT**,

e.g. FMC5_E1(460-490)_PMT(500-540)_E2(555-570)_PMT(580-680)_S





FMC6, Two Fluorophores Fluorescence and GCaMP Isosbestic Excitation



FMC5, Separated Two Fluorophores Fluorescence

Fluorescence Mini Cube with 7 ports

• Three Fluorophores Fluorescence

This mini cube separates three different indicators simultaneously.



FMC7, Three Fluorophores Fluorescence

ORDERING CODE: FMC7_E1(400-410)_F1(420-450)_E2(460-490)_F2(500-540)_E3(555-570)_F3(580-680)_S^{*}

Integrated Fluorescence Mini Cubes

In order to optimize signal detection and simplify usage, it is desirable to minimize the number of fiber optic connections to a Fluorescence Mini Cube. By taking advantages of the new *Doric Fluorescence Detector*, and our *Built-in LED Optical Heads*, it is possible to have an *Integrated Fluorescence Mini Cube*. The *Integrated Fluorescence Mini Cubes* include a number of built-in detectors and LED light sources on various ports to improve *Fluorescence Mini Cube* performance.

- The Built-in Fluorescence Detector Head allows for an increase in signal transmission of 30% due to the proximity of the detector active surface and signal source, as well as the reduced number of optical connections. A Fluorescence Detector Amplifier is provided for each detector head. All Fluorescence Mini Cube F ports come with a Built-in Fluorescence Detector Head.
- The Built-in LED Optical Head removes the need for light source patch cords. The Intensity Adjustment Ring allows additional fine control of light intensity, ideal for low-power fiber photometry experiments. All Fluorescence Mini Cube E and IE ports come with a Built-in LED Optical Head.
- All ports used for opsin activation/silencing, identified with **O**, have an FC receptacle.
- Any built-in device can be replaced by an FC Receptacle if requested. To order a cube without Built-in LED Optical Heads, use the ordering code for the iFMC: e.g. iFMC3_E(460-490)_F(500-550)_S for the iIFMC3_E(460-490)_-F(500-550)_S



Integrated Fluorescence Mini Cubes with 4 ports - LED & Doric Fluorescence Detector Head

Integrated Fluorescence Mini Cubes with 3 ports - LED & Doric Fluorescence Detector Head

Single excitation band fiber photometry measurements use a *Fluorescence Mini Cube with 3 ports*, with one port for the excitation light, one for the fluorescence detection and one for the sample being tested. The cube has a dichroic mirror to separate the excitation light from the fluorescence emission and may incorporate narrow bandpass filters that limit the excitation or fluorescence spectrum. Currently we offer configurations for GFP-like or RFP-like fluorophores. The 3 ports' mini cube filters can be customized on request.



Integrated Fluorescence Mini Cube 3 ports

On the image E is for excitation, F for fluorescence and S is for the fixed sample port.

Filter Set	Excitation Band (nm)	Detection Band (nm)	Ordering Code
GFP-like	460-490	500-550	ilFMC3_E(460-490)_F(500-550)_S
RFP-like	540-570	580-680	ilFMC3_E(540-570)_F(580-680)_S

Table 101: Integrated Fluorescence Mini Cube 3 ports Ordering Codes

Integrated Fluorescence Mini Cubes with 4 ports - LED & Doric Fluorescence Detector Head

• Excitation, Fluorescence and Opsin Activation

This cube is for measurements involving an excitation, an optogenetic activation/silencing, fluorescence detection and sample ports. Such a cube can be used for GCaMP fluorescence measurements combined with the activation of red opsins in the 580-650 nm band.

On the image E is for excitation, F for fluorescence, O for opsin activation/silencing and S is for the fixed sample port. The numbers in the brackets of the ordering code are for the corresponding wavelength bands.

ORDERING CODE:

iIFMC4_E(460-490)_F(500-550)_O(580-650)_S

• Lock-in or Sequential Detection for GCaMP Isosbestic and Functional Excitations

This cube permits excitation of the GCaMP isosbestic point with 400-410 nm light and fluorophores with 460-490 nm light. The single detector measures both signals within the fluorescence detection window from 500-550 nm. The separation of GCaMP isosbestic and functional excitations is possible if both excitations are modulated.

On the image IE and E are ports for excitations, F is for fluorescence detection and S is the for fixed sam-

ple port. The numbers in the brackets of the ordering code denote the corresponding wavelength bands.





iIFMC4, Excitation, Fluorescence and Opsin Activation

ilFMC4, Lock-in or Sequential Detection for GCaMP Isosbestic and Functional Excitations

ORDERING CODE: iIFMC4_IE(400-410)_E(460-490)_F(500-550)_S

• FRET Cube (One Excitation and Two Fluorescence Detection Ports)

This cube is used to excite the donor fluorophore with a 420-445 nm excitation wavelength band. The donor fluorophore loses part of that energy to fluorescence in the 460-500 nm band, while the rest is transferred in a distance dependent radiationless manner to the acceptor fluorophore. The fluorescence emitted by the acceptor is detected in the 528-556 nm window.



ilFMC4, FRET Cube

ORDERING CODE:

iIFMC4_E(420-445)_F1(460-500)_F2(528-556)_S

On the image E is for excitation, F1 and F2 for two spectrally different fluorescences and S is for the fixed sample port. The numbers in the brackets are for the corresponding wavelength bands. Fiber Photometry

Integrated Fluorescence Mini Cubes with 5 ports -LED & Doric Fluorescence Detector Heads

• Separated Two Fluorophores Fluorescence

This cube is used for green and red fluorophore excitation and their respective detection wavelengths. Other fluorophore combinations are possible.

ORDERING CODE:

iIFMC5_E1(460-490)_F1(500-540)_E2(555-570)_F2(580-680)_S



ilFMC5, Separated Two Fluorophores Fluorescence Cube

• GCaMP Isosbestic and Functional Excitations, and Opsin Activation

The GCaMP isosbestic point is excited by 400-410 nm bandwidth light and the 460-490 nm spectral window excites its functional fluorescence. The 500-550 nm spectral window collects the functional fluorescence of GCaMP. At the same time, an opsin is excited with 580-650 nm light.

ORDERING CODE:

iIFMC5_IE(400-410)_E(460-490)_F(500-550)_O(580-650)_S

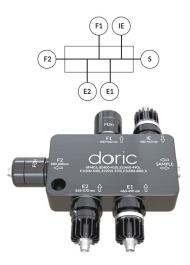


iIFMC5, GCaMP Isosbestic and Functional Excitations, and Opsin Activation

Integrated Fluorescence Mini Cubes with 6 ports -LED & Doric Fluorescence Detector Heads

• Two Fluorophores Fluorescence and GCaMP Isosbestic Excitation

This cube can be used to detect the fluorescence from two calcium indicators and the GCaMP isosbestic point. The separation of functional and isosbestic fluorescence signals of GCaMP is possible if the light sources are modulated.



ilFMC6, Two Fluorophores Fluorescence and GCaMP Isosbestic Excitation Cube

ORDERING CODE:

iIFMC6_IE(400-410)_E1(460-490)_F1(500-540)_E2(555-570)_F2(580-680)_S <u>Notes</u>: The E2(555-570) Built-in LED Optical Head has no Intensity Adjustment Ring.

Integrated Fluorescence Mini Cubes with 7 ports - LED & Doric Fluorescence Detector Heads

• Three Fluorophores Fluorescence

This mini cube separates three different indicators simultaneously.



ilFMC7, Three Fluorophores Fluorescence

ORDERING CODE:

ilFMC7_E1(400-410)_F1(420-450)_E2(460-490)_F2(500-540)_E3(555-570)_F3(580-680)_S <u>Notes</u>: The E3(555-570) Built-in LED Optical Head has no Intensity Adjustment Ring.

Rotary Fluorescence Mini Cubes

Fluorescence recording of freely moving subjects is a challenge for neuroscience researchers due to low signal and external sources of noise. Here we present a modified version of *Doric Integrated Fluorescence Mini Cube* that can be mounted on the rotor of a specialized version of our 24-channel Assisted Electrical Rotary Joint. This configuration increases collection efficiency by removing one optical connection while reducing motion related signal variation.



Unmounted Rotary Fluorescence Mini Cube with 6 ports

- LED light sources are built directly into the fluorescence mini cube, and are optimized to deliver low excitation power.
- The integrated detector is an amplified photodiode circuit with comparable specifications to our *Doric Fluorescence Detector* (See section).
- To connect these devices to an LED Driver and a Fluorescence Detector Amplifier, a 24-channel Assisted Electrical Rotary Joint for Fluorescence Mini Cube (See section) is required. Each rotary joint can hold up to two fluorescence mini cube.
- The *Rotary Fluorescence Mini Cubes* are modular, allowing them to be removed from a rotary joint and replaced. They can be ordered individually or added onto an existing system for added flexibility.



Two Rotary Fluorescence Mini Cubes with 6 ports- Integrated on an Assisted Electrical Rotary Joint

Rotary Fluorescence Mini Cubes with 3 ports

• 1 LED & 1 Integrated Fluorescence Detector-For GFP-like Fluorophore

Single excitation band fiber photometry measurements use a *Fluorescence Mini Cube with 3 ports*, with one port for the excitation light, one for the fluorescence detection and one for the sample being tested. The cube has a dichroic mirror to separate the excitation light from the fluorescence emission and may incorporate narrow bandpass filters that limit the excitation or fluorescence spectrum. Currently we offer configurations for GFP-like fluorophores.

ORDERING CODE: RFMC3_E(460-490)_F(500-550)

Rotary Fluorescence Mini Cubes with 4 ports

2 LED & 1 Integrated Fluorescence Detector-Lock-in or Sequential Detection for GCaMP Isosbestic and Functional Excitations

This cube permits excitation of the GCaMP isosbestic point with 400-410 nm light and fluorophores with 460-490 nm light. The single detector measures both signals within the fluorescence detection window from 500-550 nm. The separation of GCaMP isosbestic and functional excitations using lock-in demodulation or sequential detection.

ORDERING CODE: RFMC4_IE(400-410)_E(460-490)_F(500-550)_S

• 1 LED & 2 Integrated Detectors-FRET (Fluorescence Resonance Energy Transfer)

This cube is used to excite the donor fluorophore in a 420-445 nm excitation wavelength band. The donor fluorophore loses part of that energy to fluorescence in the 460-500 nm band, while the rest is transferred in a distance dependent radiationless manner to the acceptor fluorophore. The fluorescence emitted by the acceptor is detected in the 528-556 nm window.

ORDERING CODE:

RFMC4_E(420-445)_F1(460-500)_F2(528-556)_S

Rotary Fluorescence Mini Cubes with 6 ports

 3 LED & 2 Integrated Detectors-GCaMP Isosbestic & Functional Excitations, & Red Fluorophore Fluorescence

This cube can be used to detect the fluorescence from two calcium indicators and the GCaMP isosbestic point. The separation of functional and isosbestic fluorescence signals of GCaMP is possible if the light sources are modulated using lock-in demodulation or sequential detection.

ORDERING CODE:

RFMC6_IE(400-410)_E1(460-490)_F1(500-540)_E2(555-570)_F2(580-680)_S

Bundle-imaging Fluorescence Mini Cubes



6-port Bundle-imaging Fluorescence Mini Cube

Performing fiber photometry on a large number of sites and animals can be inconvenient due to the large number of photoreceivers and patch cords required. By bundling the connective patch cords together and using a camera, it is possible to measure a great number of sites or animals at once.

The *Bundle-imaging Fluorescence Mini Cube* is able to perform such measurements. This is done by imaging a fiber bundle onto a camera using a microscope objective. These otherwise function as any fluorescence mini cube.

- The light source ports can be shipped with integrated LED light sources (**LED** ports) or with FC receptacles (**FC** ports).
- The camera ports can be shipped with integrated cameras (**CAM** ports) or with standard C-mounts (**CM** ports).
- To accomodate larger fiber bundles, an SMA fiber optic receptacle (**SMA** ports) is used on the sample port.

Bundle-imaging Fluorescence Mini Cubes with 4 ports-Sequential Detection for GCaMP Isosbestic and Function

This cube permits excitation of the GCaMP isosbestic point with 400-410 nm light and fluorophores with 460-490 nm light. The camera measures both signals within the fluorescence detection window from 500-540 nm. The separation of GCaMP isosbestic and functional excitations is possible if both excitations are interleaved.

ORDERING CODE:

• Integrated Cube:

BFMC4_LED(400-410)_LED(460-490)_CAM(500-550)_SMA

• Connectorized Cube:

BFMC4_FC(400-410)_FC(460-490)_CM(500-550)_SMA

Bundle-imaging Fluorescence Mini Cubes with 6 ports-GCaMP Isosbestic & Functional Excitations, & Red Fluorophore Fluorescence

This cube can be used to detect the fluorescence from two calcium indicators and the GCaMP isosbestic point. The separation of functional and isosbestic fluorescence signals of GCaMP is possible if the light sources are interleaved. This cube contains two cameras, one for each fluorescence port. **ORDERING CODE:**

• Integrated Cube:

BFMC6_LED(410-420)_LED(460-490)_CAM(500-540)_LED(555-575)_CAM(580-680)_SMA

• Connectorized Cube:

BFMC6_FC(410-420)_FC(460-490)_CM(500-540)_FC(555-575)_CM(580-680)_SMA





4-port Bundle-imaging Fluorescence Mini Cube

Photodetectors

Doric Fluorescence Detector

This photodetector system designed specifically for use in fiber photometry experiments consists of the detector head and the detector amplifier. Its high gain and low noise allow for detection of signals in the sub-picowatt to nanowatt range. When used with a modulated light source and a lock-in amplifier it can detect signals in the femtowatt range.



Doric Fluorescence Detector

Fluorescence Detector Head

When the detector is not integrated into the iFMC, the Fluorescence Detector Head is fitted with an FC adapter to connect it to a signal source. The detector uses a specialized shielded cable to connect to the amplifier, keeping noise to a minimum.

Fluorescence Detector Amplifier

The Fluorescence Detector Amplifier amplifies the signal coming from the detector head and transmits it to a recording system using a BNC output. The amplifier has two detection modes (AC/DC), each selectable for all three (1x, 10x, 100x) amplification levels, allowing 6 different amplification configurations. The DC detection

mode is optimal for use with interleaved measurements or during large-bandwidth continuous measurements. The AC detection mode is optimal for lock-in modulation/demodulation and the elimination of low-frequency noise. Its output analog voltage (±5 V) can be monitored with a DAQ board for data recording or an oscilloscope for live signal.

Notes:

- The Fluorescence Detector Head and the Fluorescence Detector Amplifier are always shipped together with a Detector Cable.
- A battery pack is included with each Fluorescence Detector for cordless use.

SPECIFICATION	VALUE		
Wavelength Range (nm)		320-1100	
Peak Sensitivity Wavelength (nm)	960		
Peak Responsivity (A/W)		0.6	
Output Impedance (Ω)		50	
NEP (W/√Hz)		<12	
CW Saturation Power (nW) 4.75			
DC Bandwidth (Hz)	2) 0-1000		
AC Bandwidth (Hz)	30-1000		
Output Connector	Male BNC		
Detector Material	Si		
Detector Size (mm x mm)		1.1 × 1.1	
Amplification level	1x	10x	100x
Transimpedance Gain (V/A)	2×10 ⁹	2x10 ¹⁰	2x10 ¹¹
Maximum Conversion Gain (V/W)	1.2×10^{9}	1.2×10^{10}	1.2×10^{11}

 Table 102: Doric Fluorescence Detector Specifications

ORDERING CODE: DFD_FOA_FC

Newport Visible Femtowatt Photoreceiver Module

This battery-operated photoreceiver has high gain and detects CW light signals in the sub-picowatt to nanowatt range. When used in conjunction with a modulated light source and a lock-in amplifier to reduce the measurement bandwidth, it achieves sensitivity levels in the femtowatt range. For this Newport product Doric offers an add-on fiber-optic adapter



Newport Visible Femtowatt Photoreceiver Model 2151 + Doric FC Adapter that improves coupling efficiency between the large core, high NA optical fibers used in fiber photometry and the relatively small detector area. Its output analog voltage (0-5 V) can be monitored with an oscilloscope or with a DAQ board to record the data with a computer.

Table 103: Newport Visible Femtowatt Photoreceiver Module Specifications

SPECIFICATION	VALUE
Model	2151
Wavelength Range (nm)	320-1050
Bandwidth (-3 dB)	DC-750 Hz (DC), 30-750 Hz (AC)
Conversion Gain, Maximum (V/W)	1×10^{11}
Responsivity (Peak)	0.5 A/W
Transimpedance Gain (V/A)	$2 \times 10^{10} \& 2 \times 10^{11}$
Output Impedance (Ω)	100
NEP (W/\sqrt{Hz})	16 f
Saturation Power CW	0.5 nW
Output Connector	Male BNC
Detector Material	Si
Detector Diameter (mm)	1.0
Power Requirements	Internal 9 V battery
PRODUCT Newport Photoreceiver Module + Doric FC Adapter Doric FC Adapter only	Ordering Code NPM_2151_FOA_FC FOA_2151_FC

Photosensor Module H10722-20

The Hamamatsu H10722-20 Photosensor Module is compatible with our cubes and is the most sensitive detector we offer for very low light level detection. Unlike other ports of our mini cubes that have receptacles and a focusing lens, the port for this sensor has a lens that adjusts the beam size to fit the size of the PMT and instead of a receptacle, it has a thread that matches the thread on the Doric adapter for the photosensor. The



Hamamatsu H10722-20 Photosensor Module with a FC connector be easily damaged if exrequires a power supply

photomultiplier tube (PMT) is highly sensitive and can be easily damaged if exposed to high optical power. The photosensor module requires a power supply model C10709.

	Minimum optical power detected (W)		
Photodetector	in CW	with lock-in	
Newport 2151	0.4×10^{-12}	4.2×10^{-15}	
Hamamatsu H10722	6.0×10^{-15}	not tested in lock-in	

Table 104: Limit of Detection for each Photodetectors



Hamamatsu H10722-20 Photosensor modules directly attached to the mini cube

ORDERING:

To get PMT ready cube, replace fluorescence port code from the Fluorescence Mini Cube, F, F1 or F2 with PMT (*e.g.* FMC3_E(460-490)_**PMT**(500-550)_S).

Power Supply for PMT Module C10709

This Power Supply unit can drive photomultiplier tube modules. Both drive voltages and control voltages can be supplied from this one unit.



Power Supply for PMT Module C10709

ORDERING CODE: PS_PMT

Fiber Photometry Accessories

Fiber Photometry Rack for FMC5

This small *Fiber Photometry Rack* can fit the Fiber Photometry Console, the 2-channel LED driver, 2 LEDs, 2 photodetectors (Doric or Newport) and up to 2 FMC5. It is suitable for use with the following systems: all the 1-site Fiber Photometry Systems and the 2-site Fiber Photometry Systems for GCaMP recording with two excitation wavelengths.



Fiber Photometry Rack for FMC5

ORDERING CODE: PR_5

Fiber Photometry Rack for FMC7

This rack-mount (19") *Fiber Photometry Rack* allows for the housing of more complex systems requiring the bigger 4-channel LED driver. It is suitable for use with systems using FMC6 or FMC7, or for multi-site fiber photometry systems as the 2-site Fiber Photometry Systems for GCaMP Isosbestic and Functional Excitations.



Fiber Photometry Rack for FMC7

Bundle-imaging Fiber Photometry Driver

ORDERING CODE: PR 7

The Bundle-imaging Fiber Photometry Driver is used to coordinate the many devices used by a Bundle-imaging Fiber Photometry System. These include LED light sources and cameras. The driver is connected to a computer or the HUB using a USB cable, provided with the system.

ORDERING CODE: BFPD

4-port USB3.0 Hub

The 4-port USB3.0 Hub is used to transfer the high-density data throughput involved in using the Bundle-imaging Fiber Photometry System. The hub is provided with all necessary electrical cables.

ORDERING CODE: USB_H_4

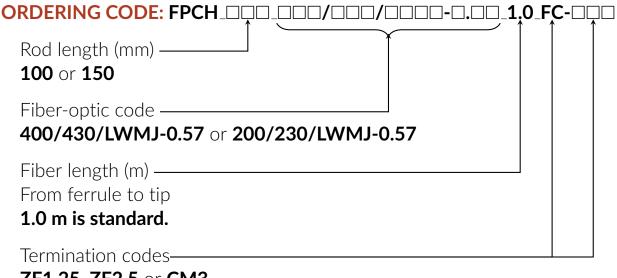
Fiber Photometry Cannula Holders

The Fiber Photometry Cannula Holder is designed to enable the recording of the fluorescence during the implantation of the cannula. It is a stainless steel rod having an FC receptacle on one end that allows a light delivery patch cord to be plugged in and at the other end a receptacle where a cannula can be screwed on. The two receptacles are mutually connected with an internal optical fiber housed



Fiber Photometry Cannula Holder

within the 6.35 mm diameter rod that fits most popular micro-manipulators. To avoid unnecessary optical losses, the selected optical fiber parameters such as the core diameter and NA match those used in the fiber photometry system.



ZF1.25, **ZF2.5** or **CM3** (see Table 48)

Notes:

- The Fiber Photometry Cannula Holder is compatible with Mono Fiber-optic Cannulas.
- A holder compatible with Dual Fiber-optic Cannulas is available on request.
- An optional clamp (**SCL**) can be used to fix the Fiber Photometry Cannula Holder on stereotaxic apparatus.

Behavioral Tracking

Technically speaking, the behavior study of freely-moving animals resembles the filmmaking or film production process involving scriptwritting, choreography, recordings, editing etc. From the neuroscientist stand point it requires:

- A) the recording of neuronal activity of the specific brain region using calcium imaging, fiber photometry or electrophysiology,
- B) the behavioral tracking or simply filming of the animal activity in a given situation synchronized with the recordings of neural activity,
- C) the behavior triggers that can be external in the form of the stage event or internal in the form of light or electric signals directed to specific brain region.

Optogenetics enable precise triggering or silencing of the brain cells with light. The electrophysiology can be used to deliver electrical trigger signals and to record the neuronal activity. The chronically implanted fluorescence microscopes and fiber photometry probes can monitor the neuronal activity. The filming of the animal is complementary information needed to establish correlation between the neuronal activity of the specific brain region and the animal behavior. The Doric Neuroscience Studio software seamlessly integrates neuronal imaging, fiber photometry, electrophysiological recording, optogenetics stimulation and behavioral tracking of the freely-moving animals. Another first from Doric.

Behavior Tracking Cameras

USB 3.0 Behavior Tracking Camera

These Doric Color and B&W cameras use an USB 3.0 interface standard typical of highperformance industrial cameras. This interface provides a framework for streaming high-speed video and related control data. The camera control and the image acquisition are done through the Doric Neuroscience Studio software. The system includes a Trigger cable to synchronize with external devices. The power is feed directly to the camera by the USB cable. An ar-



USB 3.0 Behavior Tracking Camera with Wide-angle Lens

ticulated holder is also included with the Behavior Tracking Camera.

The purchase of the USB 3.0 Behavior Tracking Camera includes:

- Camera (B&W or Color)
- C-Mount camera lens for 1/2" sensor, 1.5MP
- Articulated holder
- Trigger cable

Table 105:	USB 3.0	Behavior	Tracking	Camera	Specifications
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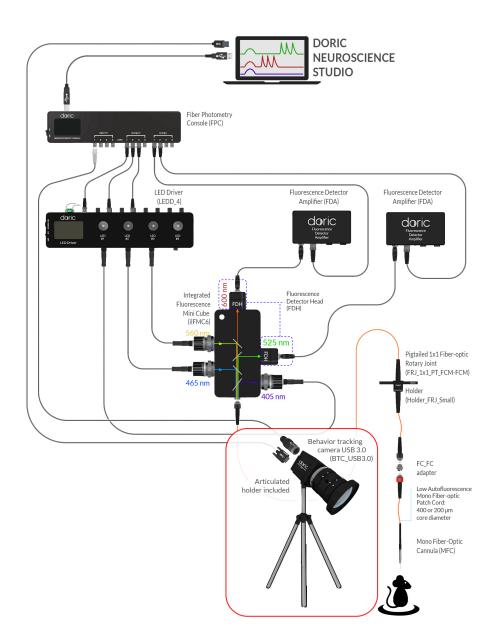
SPECIFICATION	VALUE
Video formats	B&W 1920 x 1080 Y800
	Color 1920 x 1080 RGB32
Frame rate @ full resolution	60
Resolution	H: 1920, V: 1080
Format	1/2.8 "
Pixel size	2.9 μm x 2.9 μm
Lens mount	C/CS
Interface	USB 3.0
Exposure	20 µs to 30 s
Gain	0 to 72 dB

Table 106: Behavior Tracking Camera Lens Specifications

Focal Length (mm)	Iris Range	MOD [*] (m)	FOV @ 1 m
5	F1.4 - 16C	0.10	1.0 × 1.0

Table 107: Behavior Tracking Camera Ordering Codes

CHROMA	Ordering Code
Color (RGB32)	BTC_USB3.0_CO
B&W (Y800)	BTC_USB3.0_BW



USB3 Behavioral Tracking Synchronized within a GCaMP Isosbestic and Functional Excitations and Red Fluorophore Fluorescence System

GigE Behavior Tracking Camera

The Doric Color and B&W cameras use a GigE Vision interface standard typical of highperformance industrial cameras. This interface provides a framework for streaming high-speed video and related control data over Ethernet networks. The camera control and the image acquisition are done through the Doric Neuroscience Studio software. The system includes a Power/Trigger cable to synchronize with ex-



GigE Behavior Tracking Camera with Wide-angle Lens

ternal devices. If the system is coupled with a fluorescence microscope driver, a Gigabit PoE+ Switch is included to the system. This switch allows the connection of multiple Ethernet devices to a single Ethernet port on the computer and feeds the camera power directly through the Ethernet cable. An articulated holder is also included with the Behavior Tracking Camera.

The purchase of the GigE Behavior Tracking Camera includes:

- Camera (B&W or Color)
- C-Mount camera lens for 1/2" sensor, 1.5MP
- Articulated holder
- Power/trigger cable
- PoE+ Switch (when coupled with a microscope driver)

SPECIFICATION	VALUE
Video formats	B&W 1920 x 1200 Y800
	Color 1920 x 1200 RGB32
Frame rate @ full resolution	50
Resolution	H: 1920, V: 1200
Format	1/2.8 "
Pixel size	2.8 μm x 2.8 μm
Lens mount	C/CS
Interface	GigE
Exposure	20 µs to 30 s
Gain	0 to 48 dB

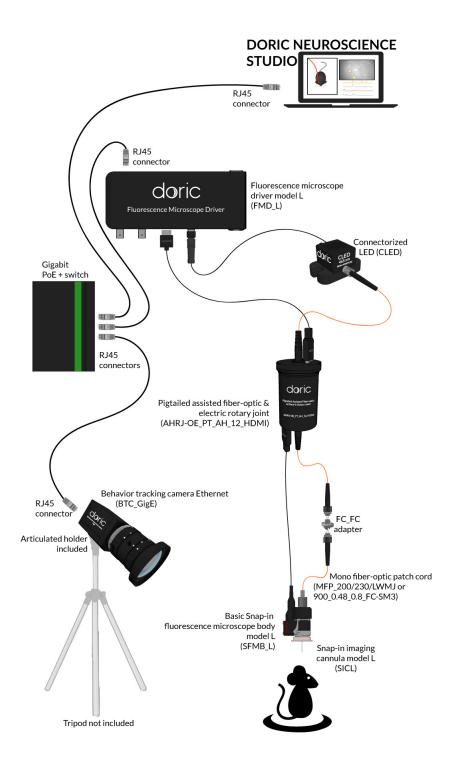
Table 108: GigE Behavior Tracking Camera Specifications

Table 109: Behavior Tracking Camera Lens Specifications

Focal Length (mm)	Iris Range	MOD [*] (m)	FOV @ 1 m
5	F1.4 - 16C	0.10	1.0 × 1.0

Table 110: Behavior Tracking Camera Ordering Codes

CHROMA	Ordering Code
Color (RGB32)	BTC_GigE_CO
B&W (Y800)	BTC_GigE_BW



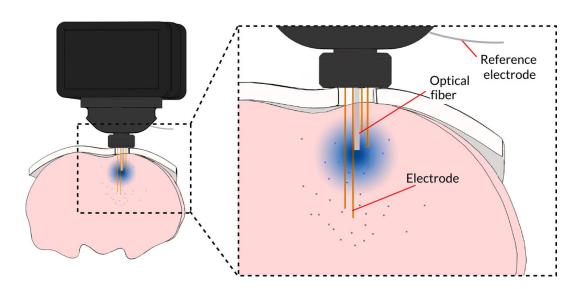
Ethernet Behavioral Tracking Synchronized to the Basic Miniature Fluorescence Microscopy System

Optogenetically Synchronized Electrophysiology (OSE)

The systems that combine optogenetics with electrophysiological recordings open up new possibilities for neuroscience. They require delivery of appropriate optical signals to the point of interest within the neural tissue and detection and processing of the electrical spikes from neural activity. The system definition starts from the chronically implanted opto-electric cannula for behaving animals or from the opto-electric probes for *in vitro* experiments or *in vivo* head-fixed configuration. For studies involving freelymoving animals , there is the wireless/fiberless option.



Detachable Fiberless & Wireless Headstage and Opto-electric Cannula



Detachable Fiberless & Wireless Headstage and the Opto-electric Cannula implanted in the brain

Optogenetically Synchronized Electrophysiology Systems

Fiberless & Wireless (Fi-Wi) OSE System

Tethering lab animals with fibers and wires compromises their "freely-moving" status for behaviour studies. Going wireless and fiberless effectively removes those limitations. *Fi-Wi OSE System* features the opto-electric cannula with one LED and up to four recording electrodes, the fiberless and wireless headstage^{*} for communication between the cannula and the control console, and finally the electrophysiology console that configures the stimulation and recording parameters and displays the real-time data. The recording tips of the opto-electric cannulas are custom made for specific experiments which ensure flexibility in recording and illuminating different brain areas.

This system contains all the items necessary to record synchronized electrophysiological signals with optogenetic stimulation of freely-moving animals.

- Fiberless & Wireless Headstage (2x)
- Fi-Wi Opto-electric Cannula (3x)
- Electrophysiology Console
- Fi-Wi Headstage Charger
- Fi-Wi Dummy Headstage
- Fi-Wi Cannula Implantation Holder
- Fi-Wi Test Cannula
- Doric Neuroscience Studio Software
- All required cables

ORDERING CODE:



Fiberless & Wireless headstage — OE, O or E (see table 112)

^{*}A Wireless Optogenetic Headstage with Multichannel Electrophysiological Recording Capability, Gagnon-Turcotte G, et al., Sensors 2015, 15(9), 22776-22797

Fiberless & Wireless Headstages

Fiberless & Wireless Headstage

The Fiberless and Wireless Headstage controls the activation of the implanted LED, records electrophysiological data from brain electrodes, and transfers all the information to the electrophysiology console. It is an electronic component that could be placed or removed from a chronically implanted cannula on the head of an animal. Typically, one needs at least two headstages so that one can be charged while the other is in use. Each headstage includes a 60 mAh



Detachable Fiberless & Wireless Headstage

battery, an LED driver, an electrophysiology recording system and a radio frequency transmitter.

SPECIFICATION	VALUE
Transmission range	3 meters (2 meters with 2 simultaneous headstages)
Continuous operating time (10% duty cycle LED + 4 ephy channels)	1.5 hours
Sample rate	4 ch @ 14.3 KHz
Size	19 x 15 x 10 mm
Weight (including battery)	2.8 g (2.5 g for FiWi_HS-O)
Battery	60 mAh, 1.2 g

Table 112: Fiberless & Wireless Headstages Ordering Codes

ТҮРЕ	Ordering Code
Opto-electric	FiWi_HS-OE
Optic only	FiWi_HS-O
Electric only	FiWi_HS-E

Notes:

- Simplified headstage versions are also available with only optical or only electrical features (see Table 112). Note that the choice of Fi-Wi Cannulas must match the type of the chosen headstage.
- The *Fi-Wi Headstage Charger* can recharge the headstage battery in less than 1 hour (**FiWi_HSC**).

Fiberless & Wireless Cannulas

Fi-Wi Opto-electric Cannula

The *Fi-Wi Opto-electric Cannulas* are opto-electric devices designed to be chronically implanted on the skull of an animal. The base of the receptacle is smaller to facilitate implantation. The optical fiber is connectorized to an LED, that brings the light directly into a specific brain area, with 1 to 4 electrodes recording the activity of the brain. The LED activation is done by the headstage and the delivered light intensity can be modulated throughout the experiment (0-100%, DC to 1000 Hz, different pulsed shapes). The position of the fiber and the electrodes is customized within a 100 μ m tolerance.



Fi-Wi Opto-electric Cannula with 1 optical fiber and 4 electrodes

The tungsten electrodes and the impedance could be chosen between 0.2 and 1.2 $\mbox{M}\Omega.$

The cannula has an electrical connector to hold the fiberless and wireless headstage in place. Between experiments, the headstage can be easily disconnected from the cannula and reconnected at the appropriate time.

SPECIFICATION	VALUE	
	465 nm - 8 mW @ 150 mA (256 mW/mm ²) 525 nm - 3.5 mW @ 150 mA (112 mW/mm ²)	
LED	595 nm - 1.5 mW @ 150 mA (48 mW/mm ²)	
	635 nm - 4.0 mW @ 150 mA (128 mW/mm ²)	
Optical fiber	250 μm diameter, NA 0.66	
Electrodes	25 μm diameter, 0.7 to 1.2 M Ω	
	50 μm diameter, 0.2 to 0.3 M Ω	
Size	10 x 10 x 7 mm	
Weight	0.3 g	

Table 113: Fi-Wi Opto-electric Cannula Specifications

ORDERING CODE: FiWi_C-OE_	FL1
LED central wavelength (nm) 465 , 525 , 595 or 635	
Number of electrodes 1, 2, 3 or 4	
Optical fiber length (mm) ——	
Optical fiber termination FLT	

Notes:

- Simplified cannula versions are also available with only optical or only electrical features. Note that the choice of Fiberless & Wireless Headstage must match the type of chosen Fi-Wi Cannulas.
- Choose the ordering code according to the type of cannula required. Use the PDF on our website to indicate the position of each electrode/optical fiber in the cannula and their length.

Electrophysiology Console

Electrophysiology Console

The *Electrophysiology Console* is an FPGA based component that controls bi-lateral wireless communication between the computer and the Fi-Wi headstage. The electrophysiological recording parameters and the LED stimulation sequence for optogenetics



4-channel Electrophysiology Console

stimulation patterns are defined in the Doric Neuroscience Studio software and sent to the headstage via the console. The headstage can be also trigged by any external source (optical gate, tracking software, etc.). After the stimulation, the data collected are transferred via RF frequency over a distance up to 5 m. The *Electrophysiology Console* can handle up to 2 headstages at the same time and each headstage can stream live 4 electrophysiological recording traces. The console and the headstage are in continuous communication which allows the activation, the cessation or the modification of the recording and stimulation parameters within 30 ms. The recorded data can be displayed, commented, saved and recalled within our software.

Main features:

- 2 antennas allowing the control/recording from up to 2 headstages at the same time
- 4 Digital Input/Output TTL, 25 MS/s, via 4 BNC connectors (could be used as triggers)
- 4 Analog Output 5 V, 16 bits, 25 MS/s, via 4 BNC connectors (IN/OUT)
- 1 Digital communication SPI and LVDS via custom pinout HDMI connector
- USB2 connection to computer, cable included
- Compatible with Doric Neuroscience Studio with Fi-Wi interface
- All software updates included

ORDERING CODE: EPC

Optogenetically Synchronized Electrophysiology System Accessories

Fi-Wi Headstage Charger

The *Fi-Wi Headstage Charger* is a device that can recharge the headstage battery in less than 1 hour. It can also inactivate unused headstages.



FiWi Headstage Charger

ORDERING CODE: FiWi_HSC

Fi-Wi Dummy Headstage

The Fi-Wi Dummy Headstage has the same shape and weight as the Fiberless & Wireless Headstage. The Fi-Wi Dummy Heastage can be secured on the implanted Fi-Wi Cannulas and allows the habituation to the weight and the feel of the real Fiberless & Wireless Headstage. This dummy is compatible with all the Fi-Wi Opto-electric Cannulas.



Fi-Wi Dummy Headstage

ORDERING CODE: FiWi_DHS

Fi-Wi Test Cannula

The Fi-Wi Test Cannula is a replica of the Fi-Wi Opto-electric Cannula that can be connected to any Fiberless & Wireless Headstage. The Fi-Wi Test Cannula can be used to test the headstage functions and the illumination patterns in the experiment location.



Fi-Wi Test Cannula

ORDERING CODE: FiWi_TC

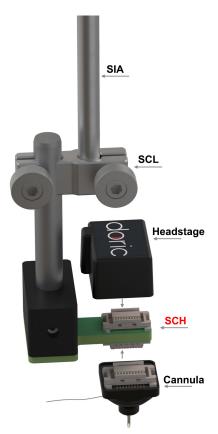
Fi-Wi Cannula Implantation Holder

The *Fi-Wi Cannula Holder* is jig designed to secure the Fi-Wi Opto-electric Cannula during the implantation. With cannula on its bottom side and fibrelesswireless headstage on the top, the holder allows for optical activation and electrical recording guidance during cannula implantation and positioning. This holder offers the possibility of installing the cannula with the headstage in two different orientations.

Notes:

- An adapter of 10 cm long can be added at one end of the holder (**SIA**; see Table 74).
- An optional clamp (**SCL**) can be used to secure the Fi-Wi Cannula Holder on stereotaxic apparatus.

ORDERING CODE: SCH_FiWi



Fi-Wi Cannula Implantation Holder

Doric Neuroscience Studio

A wide range of instruments are used in neurophotonics experiments. Light sources, cameras, detectors, microscopes and data acquisition units are few of the many devices that can be required. To ensure optimal usability of our devices, we have developed the *Doric Neuroscience Studio*, a complimentary software provided with equipment manufactured by Doric Lenses.

The *Doric Neuroscience Studio* is designed for integrated control of all devices required to perform a neurophotonics experiment. This allows convenient synchronization of device control, output signal generation, data acquisition and data processing. In addition, it comes with a suite of tools to perform basic analysis of behavior, photometric, electrophysiological and image-based data.

Software Modules

The primary purpose of the *Doric Neuroscience Studio* is the control and synchronization of the devices we manufacture. The software contains a module for each product requiring computer control. In addition, there are analysis modules for most forms of data acquired by these devices.

- **Light source** device modules are easy to use, as the software allows the generation of complex pulse patterns in many formats from the light source driver itself. These additional functions are otherwise inaccessible when the light sources are used as stand-alone devices.
- **Behavior tracking camera** device module allows the monitoring of an experimental subject while synchronized with other devices. Our analysis modules can be used to synchronize behavior video with photometric and electrophysiological data.
- **Photometry** device modules synchronize a multitude of input and output signals from electronic devices using our *Fiber photometry console*. This includes the generation of TTL and analog pulse sequences to control light sources,

as well as data acquisition from photodetectors and cameras. Our analysis module allows simple signal processing of photometric data.

- Miniaturized fluorescence microscopy device modules are used for light source control and data acquisition from *Miniature fluorescence microscopes*. Our **Image analysis** module performs basic image processing as well as automated cell detection for recorded microscope images.
- **Electrophysiology** device modules are used to send and receive signal from electrodes implanted in experimental subjects. These devices include both wired and wireless cannulas. Our analysis module does basic signal processing of electrophysiology data. Also included is an optrode simulation module that evaluates light propagation from an opto-electric cannula.

Accessories

LED Illumination Accessories

Fan Power Adapter

The renewed line of Doric LED Drivers (LEDD) has a new connector pinout that does not include pins for fan power. It is thus essential to use a *Fan Power Adapter* when using *Connectorized Multi LEDs* or *Multi LEDs + Fiber-optic Rotary Joints*. This power adapter is suitable for up to 4 channels and sold with corresponding M8 cables.

Table 114: Fan Power Adapter Ordering Codes

Adapter	Compatible with	Ordering Code
	 LEDC2 LEDC3 LEDC4 LEDFRJ (Multi LEDs) 	FPA

Optical Breadboard for Connectorized LED

An Optical Breadboard for Connectorized LED is available to mount systems including two Connectorized LEDs.

Breadboard	Compatible with	Ordering Code
cloric	CLEDCLDM	LEDB

Ce:YAG Fluorescent Illumination Accessories

Filter Holder for Ce:YAG Fiber Light Source

Doric standard bandpass filters (see Table 11) are sold already mounted in a *Filter Holder* and each Ce:YAG optical head is delivered with one empty filter holder model. Additional or replacement *Filter Holder* can be purchased using the following ordering code.

Filter holder Compatible with		Ordering Code	
	 YBPF_525/030 YBPF_549/015 YBPF_559/034 YBPF_582/075 YBPF_593/040 YBPF_612/069 	YFH	

Rotary Joints Accessories

Holders for Rotary Joints

Rotary Joint Holders	Compatible with	Ordering Code
dorice	FRJ_1x1FRJ_1x1_PT	Holder_FRJ_small
Gorie	 FRJ_1x2 FRJ_1x4 ERJ HRJ-OE LEDFRJ (1ch) 	Holder_FRJ_large
	• FRJ_2x2	Holder_FRJ_2x2
	AHRJ-OEAHRJ-OE_PTAERJ	Holder_ARJ
	AHRJ-OE_2x2AHRJ-OE_2x2_PT	Holder_AHRJ-OE_2x2

Rotary Joint Holders	Compatible with	Ordering Code
	 ERJ + FRJ_1x1 ERJ + FRJ_1x2 	Holder_ERJ
	 AERJ + FRJ_1x1 AERJ + FRJ_1x2 	Holder_AERJ

Table 118: Holders for Rotary Joints Combinations - Ordering Codes

Gimbal and Cable Holders for Rotary Joints

Table 119: Orderin	ng Codes for Rotary	/ Joints Gimbal an	d Cable Holders
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Holder	Compatible with	Ordering Code
	 FRJ_1x1 FRJ_1x1_PT FRJ_1x2 FRJ_1x4 ERJ HRJ-OE LEDFRJ (1ch) 	GH₋FRJ
	ERJHRJ-OE	НСН

Adapters for Rotary Joints

Harwin 12 to Omnetics 12 Adapter	Compatible with	Ordering Code
	ERJAERJHRJ-OEAHRJ-OE	ADAPTER_HO12

Cannulas Accessories

Polyethylene Tubing for Opto-fluid Cannulas

This *Polyethylene Tubing* is used to connect the Opto-fluid Cannulas to a liquid delivery system. The tubing is attached on the 25-gauge stainless insert of the OsFC or to the fluid injector in the case of the OmFC or the iOFC. The clear tube makes fluid flow visible. The 2-meter polyethylene tube has an inner diameter of 0.4 mm and an outer diameter of 0.8 mm.

OFC Polyethylene Tubing	Compatible with	Ordering Code
	OsFCOmFCiOFC	PT_OFC_2

Table 121: OFC Polyethylene Tubing Ordering Codes

Mating Adapters

Table 122	Mating Adapters	Ordering	Codes
		010101110	000100

DESCRIPTION	PRODUCT	Ordering Code
Zirconia Sleeve ID 1.25 mm	Ŵ	SLEEVE_ZR_1.25
Zirconia Sleeve ID 2.5 mm		SLEEVE_ZR_2.5
Zirconia Sleeve ID 1.25 mm with Black Cover		SLEEVE_ZR_1.25_BK
Zirconia Sleeve ID 2.5 mm with Black Cover		SLEEVE_ZR_2.5_BK
Bronze Sleeve ID 1.25 mm	<u>II</u>	SLEEVE_BR_1.25
Bronze Sleeve ID 2.5 mm		SLEEVE_BR_2.5
Bronze Sleeve ID 1.25 mm with Black Cover		SLEEVE_BR_1.25_BK
Bronze Sleeve ID 2.5 mm with Black Cover	N	SLEEVE_BR_2.5_BK
FC/FC Mating Adapter - Square	COM:	ADAPTER_FC_SQ
FC/FC Mating Adapter - Round		ADAPTER_FC_RO
M3/M3 Mating Adapter - Square		ADAPTER_M3_SQ
M3/M3 Mating Adapter - Rectangular		ADAPTER_M3_RC
SMA/SMA Mating Adapter		ADAPTER_SMA

Connectors

M3 Connectors offer a secured, light and small connection for multimode fibers. The standard material of the flange and the screw is titanium. Alternative to the titanium is the peek plastic.



M3 Connector - parts included: ferrule, screw, strain relief

	Ordering Code	
Ferrule Inner Diameter (µm)	Titanium	Peek Plastic [*]
125	CM3_125	CM3(P)_125
127	CM3_127	CM3(P)_127
230	CM3_230	CM3(P)_230
235	CM3_235	CM3(P)_235
245	CM3_245	CM3(P)_245
330	CM3_330	CM3(P)_330

Table 123: M3 Connectors Ordering Codes

^{*}Peek plastic can be used instead of metal for MRI compatibility.

Dust Caps

DESCRIPTION	PRODUCT	Ordering Code
SMA Receptacle Cap		CAP_SMA
FC Receptacle Cap		CAP_FC
Ferrule 1.25 mm Cap		CAP_Ferrule_1.25
Ferrule 2.5 mm Cap		CAP_Ferrule_2.5
M2 Receptacle Cap		CAP_M2
M3 Receptacle Cap		CAP_M3
Stainless Steel M3 Receptacle Cap		CAP_M3_S
Guiding Socket Receptacle Cap		CAP_GS

Table 124: Dust Caps Ordering Codes

All our products are supplied with the appropriate dust caps.

Cables

Table 125: Cables Ordering Codes

DESCRIPTION	PRODUCT	Ordering Code
M8 male / M8 female, 1.5 m long		Cable_M8-M8
BNC / BNC, 0.6 m long		Cable_BNC-BNC

Other Accessories

Table 126: Other Accessories Ordering Codes

DESCRIPTION	PRODUCT	Ordering Code
Cleaver		Cleaver
1.25 mm Fiber-optic Swab (25/bag)		Swab

For any questions or comments, do not hesitate to contact us by:

Phone 1-418-877-5600

Email sales@doriclenses.com

doric

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Integrated Fluorescence Mini Cube

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